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On Movement of Muscles by Galen of Pergamon

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ABSTRACT Galen begins with definitions, giving the special differentiating characteristics of nerves, muscles, tendons, aponeuroses, fascia, and other connective tissue elements. He demonstrates that contraction is the specific activity of muscles. It occurs voluntarily through impulses from the brain carried by the nerves. Protagonists and antagonists are described and demonstrated by experiments with severing individuals and groups as well as with a model made of articulated bones and thongs attached in proper places to represent muscles. Attention is called to passive motion of muscles. The action of gravity is related to muscular action. Sustained contraction of muscles is called tonic activity. The effect of habit on muscles and the subconscious control of voluntary muscles are explained. Visceral organs and blood vessels are under the control of nature rather than the brain, and are thus contrasted with the muscles.

A translation of this treatise is important because it gives the reader an opportunity to judge for himself the validity of the criticism that Galen did not distinguish between nerves, tendons, and ligaments. Within the first few pages it becomes evident that he understood their differences both structurally and functionally to a surprising degree, although he is puzzled over the termination of the motor nerves because he had no microscope with which to follow them.

The title of the treatise is misleading to the modern reader because it does not mention the nervous system. Galen pictured the muscles as instruments of the brain, controlled through the nerves. In demonstrating this he introduced a large amount of information concerning the activity of the brain which is useful in understanding some of his other treatises, especially those dealing with psychiatry.

In the translation an attempt has been made to follow the Greek of the Kuhn (1821–1835) edition as literally as possible and at the same time convert it into an approach to easily readable English. Two words have been particularly difficult to translate, namely, PSYCHE and PHYSIS. Galen used the former to designate the organ which exerts voluntary control over the various structures of the body, especially the muscles. In so doing he contrasts these structures with others not under this control, but which are the province of nature, that is, the PHYSIS. The word PSYCHE, therefore, will be translated brain and PHYSIS nature. He occasionally used another word for the brain, ENKEPHALON, but by this he meant the dissectable structure in the head contrasted with the spinal cord. Still another word is used for the more dynamic and less coporeal concept of the brain, namely NOUS, and this is translated mind in order to emphasize this distinction.

The following typographical conventions will be used:

The numbers in square brackets refer to the pages in volume 4 of the Kühn edition (1821–1833).

Greek words taken directly into English are transliterated into Roman Capitals.

Greek words are given their Greek endings.

Titles of Galen's own works appear in quotes.

Modern anatomical terms, where appropriate, are taken from the 1965 Nomina Anatomica, italicized, and placed in parentheses.

Editorial comments and explanations are enclosed in square brackets.

BOOK ONE

Chapter I

Voluntary motion in the various parts of the body is brought about by the contraction of muscles. Their number is so great, however, that they cannot be counted with ease, for they unite with one another, some

soon as they join with the muscles, the nerves divide in many directions, some divisions go to one part, others to others, and at last, completely separating into delicate membranous fibers [INAS], they form a plexus through all the body of the muscle. The fascia by which the muscles are united and bound with the bones gives rise to the membranes which surround them and penetrate the flesh of the muscle. It appears to me, therefore, as if the muscle were a region [372] irrigated by several conduits, that is, by the first mentioned conduit, the nerve, and by two others, one the conduit for blood which is hot, thin, and vaporous, and the other for thicker and colder blood. The former of these is called the artery, the latter the vein. These same conduits, having their sources [ARCHE], from the heart, and from the liver, permeate the substance of the muscle and through them the muscle becomes not simply a place, but a vegetative being [PHYTOS = plant].

Through the third conduit, that from the great source, it becomes not simply a living thing but something in addition to a vegetative being, by acquiring sensation and the power of voluntary motion, attributes by which the animal is differentiated from the non-animal. Through these powers the muscle has become an instrument of the brain [PSYCHIKON ORGANON] exactly as it has become a natural or living one [PHYSIKON] through the arteries and veins, for the motions of the arteries and veins are simply living or natural and without impulse, but those of the muscles are voluntary and by impulse. Whether you say the movements of the muscles arise by deliberate choice or act of free will or by intention makes little difference, for the one object at which we must aim with all these terms [373] is to distinguish the notion of muscles from that of the arteries and veins. If it is not sufficient to point out the difference in words, you may demonstrate what you wish.

Why do we say that the muscle is not an organ of sensation, but solely of motion, and why does it not partake equally of both? Is it because voluntary movement would not occur in animals without the muscles, so that a muscle is the proper organ of motion, but sensation resides in all the sensitive parts and independent of muscles? For whichever of them shares the nerve, that, at any rate, has sensation. It has been clearly stated, then, what the muscle is, namely, that it is the instrument of voluntary motion. It was stated, also, where the source is from which its motion arises, as well as how it comes from the brain through the nerves. It was stated also how the latter as well as the binding tissues ramify with it.

Chapter II

It still remains for us to speak concerning the nature of the tendons in order that nothing will remain obscure for the [374] discussions to follow. It was mentioned above, the constitution of tendons is a mixture of binding tissue and nerve. The elaboration of this was set aside, but now will be presented. Tendon is harder than nerve by as much as it is softer than binding tissue, but the bulk of its substance is great enough to be formed of both. Binding tissues are all without sensation, but all nerves are sensitive. The tendon is not insensitive because it contains nerve but it is not as sensitive as nerve because it is not solely nerve. To the extent that it has the nature of binding tissue, the accuracy of sensation is blunted. The tendon arises from the tail [insertion] of the muscle. Attaching to its head, however, are both nerve and binding tissue and they are distributed to the whole muscle making it reasonable that the tendon is formed from both.

The way to learn this more distinctly is through dissection, for then you would see clearly that the origin which they call the head of the muscle is more sinewy [NEUROEIDES = nerve-like], the middle, more fleshlike, [375] is called the belly of the muscle, and thereafter it becomes more and more sinewy in proportion to the beginning until finally the lower end seems more sinewy than the head. The nerve, having reached it at its origin, is distributed in small subdivisions; these in turn dividing into others and again the latter divide into still others, until this division advances in such a degree as to end in membranous and very thin fibers. Continuing farther, in

seeming to appear single, and some, although ending in many tendons, remain entities, however many the tendons seem to be. Because of this and because they are of diverse shapes and attach to dissimilar parts, it is quite difficult to investigate the manner of their moving. This may also be to some extent because they are secured to the moveable parts at different and often *opposite places.* [368] *Thus, they insert,* some above, others below, some in front, others behind, some on one side or some on the other. Accordingly, if a whole muscle has been cut into by a wound not too slight nor near the surface, some of the motions of the part to which it is attached are interrupted. Also, when its injuries are of many kinds, it becomes most difficult to comprehend the manner in which it moves. For example, if one or another of the muscles in the legs has been cut, a person may be unable to bend, straighten, raise, lower, or turn the limb about. In a similar manner, inflammations, scars, necroses, and bruises, or indurations affecting a whole muscle have the same effect, not only on the legs but also on the hands. For involvement of some of the muscles would cause loss of power to raise, others to bend, extend, or lower the hands, or twist them to either side or turn them backward. The same thing happens if the tendons are affected. (The more recent observers call them aponeuroses of muscles because, I think, they see the muscles [369] coming to an end *in them.*) The nature of tendon is something of a mixture and is intermediate between fascia [SUNDESMOS binding tissue or connective tissue] and nerve. Fascia, at least in its specific meaning, not as the name commonly implies (probably refers to ligament), is a nerve-like substance, always seen associated with bone, intervening between either bones or muscles, clearly its name (i.e., binding tissue) comes from its purpose.

Nerve [NEURON] or sinew [TONOS], on the other hand, originates from the brain [ENKEPHALON] or spinal marrow (cord). (Although one organ, it has been called by two names from its functions because it produces bending and pulling.) The consistency of its substance may be imagined if you think of the brain as having been compressed and packed close, thereby becoming harder. The substance of the spinal cord is similar to the brain but, having been compressed, it has become hard. Indeed, within the brain itself, the posterior portion which is continuous with the spinal cord is harder than the anterior part, and some nerves which are softer than others will not seem to you to differ *from the spinal cord.*

The marrow in the other bones is not the same, for it is wet and almost liquid. You might compare it most of all [370] to fat in softness. Moreover, you will not find any nerve either soft or hard arising from this latter marrow, nor is it clothed by the meninges of the brain and spinal cord. *Arteries and veins do not form a* plexus through it as they do in the brain and spinal cord, nor is it associated with any muscle. There is an important association between the brain or spinal cord and all muscles, however, for each must receive a nerve from either the brain or the spinal cord.

This nerve appears small but it is, indeed, not slight in power. This you may observe as a result of injuries to it, for either cutting, compressing, contusing, scarring, necrosing, or binding with a ligature would eliminate all motion and sensation from its associated muscle. When it is inflamed, however, convulsions and delirium are caused in not a few persons. Some individuals, being thus afflicted and meeting one *of the more skilled physicians,* have had the nerve cut and straightway were relieved of the convulsions and delirium. Henceforth the muscle to [371] which the nerve ran was insensible and useless for motion.

Thus in the nerves, some great power resides flowing to them from the great source [ARCHE]. The power is not inherent in the nerves nor does it stem directly from them. You may demonstrate this especially well if you cut any one of these nerves or the spinal cord itself. Then whatever part is above the cut and connected to the brain [ENCEPHALON] will retain the powers of the source, while all the part below can supply no sensation or motion. The nerves, being analogous to conduits, carry power to the muscles from some fount of the brain. As

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turn, the latter join with each other and create sinews, larger in bulk than they were formerly, but smaller in number, until they become at the end of the muscle as great in number and size as are those found at the first origin. Since the tendon becomes much larger than the nerve running to the muscle, it is quite clear that it is not formed of nerve alone but appropriates something of the nature of the binding tissue, and this in no small part. For in many places it seems six times and in many places ten times as thick as the nerve [376] and then its constitution is suitable in kind and size to provide the function of both the nerve and the binding tissue. It binds the muscle to the subjacent bone to which it inserts, and in this way does not differ from the binding tissue. It is sensitive and is moved and in this way shares with nerve. It becomes greater than the nerve, being designed to move the bone. The whole tendon is implanted for the most part toward the end of the bone invested with cartilage, not indeed at its very end nor in its accustomed shape, but flattened out, and wound around the bone at its upper end, which is called the head. The tendon, attaching itself to the underlying bone, is pulled along by the contracting muscle, for the muscle needs some firm cord to move the bone and nothing would be more suitable for this than binding tissue. The nerve from the brain, being a path for the motive power, contributes its share by extending along and mixing together with the binding tissue and in this way the tendon comes from both. [377]

Chapter III

Every tendon is attached to bone, generally speaking, but not every muscle ends in a tendon. No tendon comes from the muscles moving the tongue. It is not necessary for the tongue to move bones; instead, it articulates sound, distinguishes juices, and assists in chewing and drinking. If it seems to anyone that the heart is somewhat similar, he must not have studied the substance of muscle carefully, or he would have recognized that they differ much both in thickness, moulding, texture, and hardness. Their actions, moreover, are not alike. The motion of the heart, being compounded of the double and complex perpetual motion of diastole and systole, does not require voluntary impulse for its initiation. The motions of the muscles are not the same and would not arise without voluntary impulse. There are some binding tissue cords in the cavity of the heart, exactly like tendons, about whose purpose we speak elsewhere, the name binding tissue being used here in its general meaning. [378] The lips of the mouth, composed of a careful mixture of skin and muscle without bone, are moved, just as the eyes are moved, by voluntary impulse. The latter are moved by muscles but bone is not moved with them. The skin in the forehead, eyebrows, and the greatest part of the face is moved by impulse while the bones remain still. This skin is different from the eyes and lips because it has a thin layer of muscular nature stretched under it in place of muscle, but the eyes are moved by muscles and the composition of the lips is a mixture of skin with muscle. If you consider the entrance to the stomach, which the ancients called the esophagus [OISOPHAGOS], to be muscle and to do the work of muscle in animals, this would be a muscle not ending in tendon nor operating bone. The muscle at the neck of the bladder receiving the urine has a substance exactly like muscle and performs the work of muscle, and so likewise with the perineum, whether it is fitting to call the latter a single muscle or [379] many attached together. Bones are not moved by them nor by the muscles running down to the testes and the penis.

In summary it can be said of all muscles that they are the organs of voluntary motion. On the one hand, pulling the skin at their own origin, they draw themselves together as those in the lips and the brow and those of the entire face. No tendon arises from these. On the other hand, all other muscles which move bones generally end in tendons to a greater or lesser extent. Some of the muscles which move something else have tendons and some do not. The muscles which move something other than bone are those of the eyes, of the tongue, of the testes and penis, and those of the pharynx. The muscles of the eyes have membranous but strong [380] apo-

neuroses attached to the sclera and choroid surrounding the nerve-like coat. The muscles of the penis and testes, not forming any aponeurosis, attach to the fleshy parts of their own organs. Some of the muscles of the pharynx and larynx give origin to indistinct aponeuroses, others to none at all.

One kind of motion may appear in one place, another in another. No motion exists that is not intrinsic to the tongue, for it moves up and down, forward and backward, unrolls, and moves around in a complete circle. There are four direct motions for the eyes, upward, downward, to the right and to the left and two other motions, rolling around in a circle. When the two muscles of the temple, by pulling, approximate the teeth to each other, they protrude in an arch, but during gaping, on the other hand, they slacken and are drawn in. The motion of the great muscle in the arm is sufficiently evident. When it bends the arm at [381] the elbow, it protrudes in an arch and pulls itself together, but in straightening of the elbow it slackens and is drawn in. The great muscle of the forearm has the same motions clearly evident. In bending down the fingers it protrudes into an arch and pulls together, but in extension of the fingers it is stretched out, slackened and drawn in. And so it happens with almost all the other muscles. Those in the thigh appear to have a twofold motion if you strip them of the skin, or indeed they are apparent without the skinning on account of their size. In lean muscular bodies the largest of the muscles when moving are seen clearly before they are stripped of their skin.

Some muscles have a peculiar motion because their shape is different. The muscle at the anus would acquire a motion appropriate for pouches closed by a pulling together (as by a purse string). The diaphragm is like this to some extent except that it has no hole made in it. You may observe the motion of this muscle clearly if, dividing the peritoneum, you draw aside the underlying viscera. But for observation of other muscles of the [382] thorax and the whole epigastrium, it is necessary only to raise the skin. What happens to the muscles of the epigastrium is the opposite of what happens to those of the limbs and of the face. The latter when contracting protrude and bend into an arch but the former are drawn in when contracting and protrude in an arch when relaxing.

Chapter IV

Now, do the muscles (and this was the first thing examined at the beginning) have any types of motion which are seen after they are dissected into parts or before the body is denuded or are they much less numerous than they appear? Even setting aside other reasons, it would be as extraordinary, if all the muscles did not have a single motion, as if someone said that different arteries have different motions, for nature always seems to function similarly through the same organs. That six motions exist for all muscles, as some have said already, clearly has been refuted. None of the muscles in the arm and the leg has any third motion other than lengthening and shortening [383], just as each of the two temporal muscles evidently has two motions. Even if the muscles move the whole member in six directions, it does not cause doubt that a twofold motion exists in each of them. For, if only one muscle moved the whole member, it would be necessary that whatever motions the member has would be the same as the motions of the muscle. Since there are many more than just six muscles in each of the limbs, would it not be astonishing if different motions arose from the different muscles? I think the tongue has deceived those who declare these things because they suppose that it is a single muscle. If it were a single muscle, it would demonstrate that many motions belonged to one muscle. On the contrary, since not one but many muscles move the tongue, I think it is to be concluded that there are not many motions belonging to each muscle. It would be useless to have many muscles if it were possible for all motions to be accomplished by one. But, you may say, each of the [384] eyes goes through motions in four directions. True, my friends! And, what is more, there are four rectus muscles. There would be only one for each eye if it were in accord with nature for a muscle to move in all directions. Just as, if there were one mus-

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for just as common occurrences, considered by themselves, indicate some common function, and obscure whatever is peculiar to each, when the different occurrences make a display of the function peculiar to each of the muscles, then the truth becomes evident. The occurrences peculiar to each of the muscles are these: when the internal muscle is cut through, the part remains thereafter in the position of extension; if the external, it is flexed and cannot be extended. If, taking it with your own hands, you flex the extended part or extend the flexed, you will accomplish each of them easily but forthwith the part will return to the previous state. What is demonstrated by these things? That it is flexed by the internal muscle, and extended by the external. For this reason, if the external is wounded and loses its action while the internal remains active, the latter will bend the part since the muscle intended by nature to flex would be there, unwounded. If the internal [389] is cut across, the opposite takes place; the limb is extended and no longer can flex. For what reason does the part remain motionless in each position? Is it because the motions which follow one another in succession are abolished? The muscle meant by nature to flex, being sound, flexes the first time but cannot flex a second and a third because the member does not undertake to extend again. Flexion of the extended part alone is possible. The muscle intended by nature to extend, by the same logic, extends first and cannot a second or third time, because no muscle again undertakes to flex the part, for extension of the flexed only is possible. Under these conditions if you, mimicking the action lost by the wounded muscle, with your hands extend the flexed part, you will see that the motion intended by nature to flex has been preserved. For without you it will not be flexed spontaneously by the action of the muscle placed on the internal side when it pulls, since it will not be extended again by any muscle but will always require you to do it. Likewise, if you cut the internal muscle, the part will always remain extended without you. It will not be flexed by [390] any muscle but will need you to make this motion. From what has been said, therefore, it is clear that, as the work of the internal muscle is to flex, so the work of the external is to extend.

It should no longer be obscure that to pull and draw themselves together is the inherent function of the muscles whereas it is to lengthen and be relaxed when the antagonistic muscles contract and draw themselves together. You may confirm this from other not unimportant phenomena similar to those observed at the beginning. If you take in your hands the legs of a bird completely removed from the body, and attempt to pull its tendons with your own fingers, first the internal and then the external, on the one hand you will see the limb flexed and on the other extended. Also, if the leg preserves intact its attachment to the body of the animal, and you take each of the tendons or muscles you wish to pull, you will see in turn the part being flexed by the internal and extended by the external. If you wish to cut through a muscle either in [391] the dead or living animal, you will see clearly that of its two parts, one will be drawn upward and the other downward, each toward its own extremity, and you will recognize this effect at whatever part you cut across the whole muscle. So it is clear that every part of the muscle has the inherent motion of retracting to itself. For if you cut only the head of the muscle above, all the muscle belly is carried to the tail, but if the tail is cut, the muscle is drawn up to the head. And if you cut it at each end, you will see it round up and retract toward the middle from both ends. I believe that in each of the examples presented the material is explained with sufficient clarity.

Chapter VI

On account of the efforts of the physicians and philosophers who are attempting to cast doubt on all inquiry into activity, however, not only the above material but also everything else about to be presented lies before us. When any of the muscles or tendons placed internally on the limb have been scarred, [392] the part, having become flexed, is not again extended, or when any of those placed externally are scarred, the part, having become extended, is not again flexed. This is contrary to what

cle, in this instance it would show that four motions exist for each muscle, likewise, it shows that one motion would arise from each, since four is the number of muscles, and correspondingly, when the eyes turn about, each of two muscles makes one motion.

By your own words, however, they say, it is agreed that if not many, at least two motions are seen in each of the muscles. What then, is the logic in saying there is one? It is not surprising for each has one motion as its activity and the other incidentally. It is active when it draws the moving part toward it, but it is not active when it is dragged away to the opposite side by another muscle. On account of this, no moving part is furnished with one muscle only. If some muscle attaches above, another certainly attaches opposite below; if one from the right side, then certainly another from the left. For when each of the moving parts is drawn in opposite directions by the guiding reins [385] of the muscles, the latter are alternately in contraction and relaxation. The one contracting draws the part to itself, the one relaxed is drawn by the part. Because of this, both muscles are moved in both motions. The action of the moving muscle is to contract, not to follow. It does follow passively, however, just as any other part of the limb when it is inactive.

Shall we postpone our attempt to demonstrate that all the muscles have one inherent motion, or shall we first agree among ourselves on all the phenomena observed in them? The latter seems to me to be the best way to begin. Let us mention their manifestations one after another, therefore, in order not to neglect anything. First, let us take up one point which was mentioned in the earlier chapters, namely, that when the muscles have been cut across completely, any motion in the subjacent parts is destroyed, but when cut into, it is hindered. The amount of the damage follows the amount of the cut, more of the motion being destroyed in the greater incisions, and less in the smaller. [386] Let us grant that all these effects apply to the tendons also, for if you cut the latter entirely, you paralyze the motion of the whole part, but if you cut into them, you damage them as much as you cut. If all the motions of a part were to be abolished when one muscle is cut, you establish, I think, that this one muscle is the producer of all the motions.

Again, if only one motion is abolished when one muscle is cut, you establish that the muscle cut produces the one motion. Since it happens that neither a single nor all motions are abolished, but always two, it would seem to be established that two motions arise from one muscle. On the other hand, when the muscle or tendon placed on the opposite side is cut, the same two motions are abolished and, dominated by the same logic, therefore, we say that the same two motions are governed or disabled by them both equally. Therefore, if one muscle is destroyed, either the motion of the one opposite is destroyed with it, or they are not designed by nature for the same performance, and each is responsible for its own motion only.

If either of these propositions is true, [387] we must of necessity attempt to demonstrate which one, after having first stated more clearly that if one of two motions alternating with each other is abolished, necessarily the remaining one will be abolished along with it. Let us suppose that the motion of extending the limb were destroyed. The part would be flexed first and remain thus thereafter, not having the power to extend again because it has been deprived of the extending motion. Moreover, since it can no longer be extended, it cannot again be flexed because it must be extended first before it can be flexed. In the same way, if the motion of flexing the part happens to be destroyed, first it is extended, then becoming motionless thereafter, it is no longer able to return to flexion which must precede if extension is to follow. It seems to be altogether true that the motions opposing and succeeding each other are destroyed together. It is right to inquire, therefore, whether two motions are produced by both muscles, or one by each, with the one remaining being destroyed with the first. [388]

Chapter V

How, then, will this distinction be made? By the different occurrences [SYMPTOMA],

toward itself. So you will find all the actions of the arms of wrestlers, archers, carpenters, and those doing all the other things, executed by the muscles. This now seems to me to be clear and no more discussion is necessary.

What has not yet been discussed and is not clear must, therefore, be mentioned at this time. Not all motions of the hand arise through action of the muscles and not all lack of motion through their inactivity. It is possible to discover movements in which all the muscles are inactive and absence of motion in which a great many are active. Let us speak first of the [397] movement. It will make our discourse clear to mention first two motions of the whole body which resemble each other but which do not arise in the same way. One of these is called lying down, the other falling down. It is evident that lying down takes place voluntarily, falling down involuntarily. Lying down is accomplished by the action of muscles, and for this reason is a voluntary act of the living being. Falling down is not an act but something that happens to one involuntarily and no muscle need to be acting. It is only necessary for all the muscles to be released from tension and the body, yielding to the weight, is carried downward in the direction toward which it was inclined. As falling differs from lying down, likewise bringing the arm down differs from letting it fall, for when it drops, drawn by the weight inherent in material bodies, all its muscles remain idle. When it is brought down, on the other hand, all the muscles in the axilla pull the humerus toward them. Thus we have discovered a third motion besides the two cited earlier. Of the latter, one, [398] in which they are active, represents a drawing together of the muscles on themselves, the other, in which they lie idle, being lengthened by their antagonistic muscles, takes place by some chance, and is not inherent in them.

The third motion now being brought forward resembles these latter in no way for in it no muscle shortens or lengthens. Does nothing move at all? When the whole limb is carried down by its weight, it is not to be admitted that the muscle remains motionless, being part of the limb, but it is moved bodily in this instance and is not lengthened or shortened. What kind of motion is this? The same, evidently, as that of the bones. Not being shortened or lengthened it is carried about together with the limb as if it were some inanimate body which happened to be attached to it. Contraction is the power of a muscle as it would be of an instrument of the brain [PSYCHE], lengthening would be not a power of an organ but simply a motion while the third motion just expounded does not belong to muscles alive but as inanimate and [399] completely motionless by themselves.

Passing over to the fourth motion, let us investigate the manner of it, for it seems to be quite opposite to the third. In the third manner of motion it was pointed out that the muscles remain idle while moving, but in the fourth it will be pointed out that they are active while appearing not to move in the least. We have in mind that the hand, having been raised up, is maintained in this position. We ask ourselves this: "Why is it not born down, brought downward by the weight?" We respond, thereupon, "Because the contraction of the muscles carrying it up persists." Until they are completely relaxed, the hand is not able to move back. The traction ceasing, however, if no other muscles contract and all remain inactive, it descends to that side where the weight pulls it. If some other muscle does contract, it is moved where the latter pulls. So, in keeping the arm extended, the traction of the muscles putting it there is maintained. Must one say that they are active and pull and do not move? [400] If you are too cautious to say this, you must say that they are not active. Would it not be absurd to agree that they are active in their inherent and most intimate activity and not to agree that they move? And yet, they do not appear to move. Why is it necessary to introduce contradictions when the unravelling of them would become most difficult and hard to interpret? Because, when difficulties are mentioned, we make a mistake if we lay stress upon contradictions. If, for this reason, we are not among those who, in order to prevent discovery, promote doubts, but rather are among those who, in order to investigate accurately, look about carefully in all directions, we must compromise all opposing views by leaving

takes place in wounds. In the latter conditions the part is drawn up in a direction opposite to the muscle wounded. That this is not opposed to the preceeding but greatly confirms it is apparent, for the scarring by its very nature makes everything contract. That which in the healthy muscle was caused by impulse now appears to come from the pathological process. On the one hand the motion is voluntary, caused by impulse, on the other involuntary, caused by a pathological condition. The affected parts cannot be pulled in the opposite direction by your own hands, as they could with wounds, for the scar, having replaced the muscle with a binding cord, holds against you. If it were possible for you to draw the member toward the opposite position with your hands, nothing would hinder it from being returned by the antagonistic muscles, since it would then be moved by the appropriate muscles in a natural motion. [393] The conditions caused by scars are mimicked by inflammations, for inflamed muscles and tendons pull the part toward themselves and frequently make it immobile. The induration following healed wounds no less than the above-mentioned maladies frequently immobilize it. All these conditions and those about to be mentioned no less seem to work together to prove the same point. In addition to the latter, however, many other problems are still before us to be solved.

If all the muscles have but one kind of motion, it might seem astonishing and almost impossible, that one part, such as the hand, would be extended at one time, flexed at another, or moved about from one side to the other, raised, lowered, or be turned backward toward the spine. It does not appear astonishing, however, when we know that to raise and lower the arm is the function of the muscles moving the shoulder joint; to extend and flex the forearm those at the elbow joint; to turn it to pronation or supination [394] those of the joint between the humerus and radius. In touching the spine, the motion comes from the same activity of the arm through the four joints moving together, that is, the lowering of the shoulder, flexing at the elbow, turning the forearm to pronation, and turning the wrist backward. All arise by the functioning muscles but it is not fitting to say at this time which muscle performs what, for in the treatises "The Dissection of Muscles" and "The Purpose of the Parts" as well as in "The Anatomical Procedures", their great number and the motion of the parts brought about by each of them has been described.

The discussion which lies ahead of us also contributes to the problem already mentioned, namely, that it is not astonishing that, if one manner of motion exists in the muscles, the limb may change posture in many ways. Each muscle draws the part to which it is attached toward itself; it can pull either from the right to the left, or from left to the opposite, or one flexes, the other extends. When many muscles [395] act on many joints at the same time, how is it astonishing that much diversity arises in the postures of the limbs? Muscles inserting at the head of the humerus, I think, raise it; those inserting in the forearm on the outside of the elbow, extend the ulna; those which insert internally in the forearm and which cross it obliquely, move the radius to pronation; those in the forearm which insert on it externally, extend the wrist. Each of the fingers is flexed by the internal tendons and when each of the fingers has been flexed, the posture of the hand is especially like that of a boxer thrusting out his fist. If the humerus has been raised fittingly and the ulna has been extended exactly to supination, and if those muscles in the forearm attaching externally have bent the radius and the wrist back at the same time that the digits are extended, the posture of the whole arm would become the same as when extended by someone wishing to receive something. In the same way if you keep the other joints in position but change only the supination and place the hand exactly in the midposition between supination [396] and pronation, the entire posture you make will be that of the arm which is adopted especially by the archers when they shoot arrows, as Hippocrates has said. So also in each of the other postures of the whole arm it is not difficult to discover the position of each of the joints if you remember this one point, that each muscle, when it contracts, pulls the part to which it is attached

tions by the antagonistic muscles? If it did not happen thus, someone might equally well charge the power of the brain with seeming to command the muscles to lie in repose, whenever they must be moved by the antagonistic muscles. But if we agree upon this, we see immediately that it is contrary to what we said above. For if we say that the movement of the muscles is not by the psychic pull, [405] and that lack of movement is, we find that this contradicts many of the phenomena observed.

First and foremost, when the nerve running to the internal muscle has been cut, that muscle is seen to lengthen at once and to remain in this state thereafter. If indeed the shortening were inherent in its substance and the lengthening originated by command of the brain, it would be necessary for lengthening to be abolished sooner than shortening when the communications with the source [ARCHE] are cut away. The opposite, however, takes place. It shortens when the nerve is uncut; it lengthens under the opposite condition. It would be necessary, not only that the nerve of the cut muscle abolish the lengthening but also that the one not cut preserve both the lengthening and the shortening, if indeed the muscles obtain lengthening through the nerve and shortening through themselves.

What solution do we have of this difficulty? Here among these same peculiarities of things it is necessary to find the difference between the motion of the substance of the muscles [406] and the power making use of it. What is the difference? When the external muscle or tendon is cut, the part is flexed on the spot. Even should we not wish to flex it, it would be flexed by something, that is, and one must not neglect this, by the body of the internal muscles designed by nature to retract to themselves. If the flexing of the part is not from our volition, how could it be said that it arose from the power of the brain? In order to know what the proper motion of the power is, order a wounded man to attempt to flex completely the mutilated part and you will see it obviously bent. Then command him to discontinue the impulse of bending and you will see the mutilated part extend to the point to which it was carried without impulse when it received the first flexion. From these phenomena, obviously, it is to be learned that the body of the muscle would never arrive by itself at exact and complete flexion if not urged by the power from the brain. It was in vain, someone will say perhaps, for the body of the muscles to have a constitution for shortening when it would be accomplished more completely [407] and better from the functioning of the brain. Whoever says this must be one of the lovers of difficulty and ambiguity, as they call themselves. Should I ask him gently if he believes that the part meant by nature to lengthen is the most suitable organ to have the power to contract, or the opposite? I cannot imagine how anyone could trump up an organ more unfit by nature for motion than if he made it disposed toward the opposite of the voluntary impulse. If the latter is unfit, then the opposite nerve is opposed spontaneously, then that which is disposed toward motion is most appropriate. Indeed, if the power of the brain urges the muscle to pull according to its intrinsic source, this preparation would be adapted for it, and this is how it is.

Chapter IX

Should someone inquire into this more reasonably and be puzzled as to why, speaking generally, we say that extreme shortening is the proper motion of the body of the muscle if, after complete lengthening, it shortens and after complete [408] shortening, it lengthens. Either, one must say that neither motion is proper to it but is brought about by some chance or, one must suppose that both are equally proper to it. Or again, must one suppose that it departs farthest from complete lengthening and least from complete shortening and for this reason that shortening is the more proper to it? Since there are two extreme positions, as someone might say, greatest lengthening and complete shortening, and since it is more proper for the body of the muscle to shorten than to lengthen, it would take up a position exactly intermediate between the two and thereafter would return to this position whenever released. But this does not take place, and instead,

nothing concealed. On the one hand, when the muscles are active we say that they move themselves; on the other hand, when they appear to move neither the whole limb of which they are a part nor themselves alone, we do not venture to agree that they move at all. *What solution may one find of the difficulty? Shall we hypothesize contraction called tonic* [τονικος=pulling], [401] *or find something else better, or shall we decide nothing concerning this until we have examined it carefully and then speak up? This seems to me to be much better.*

Chapter VIII

Let us do so then, and let the discussion proceed immediately as indicated above. Let us observe some inanimate object, either wood or stone, being dragged about by some force. Again, let us observe this same object pulled back contrariwise in the opposite direction by some other force and suppose that the first dragging force prevails by its might and for this reason the object follows it, but much less than if pulled back by no opposing force. Let us present a third situation for this object: where it is pulled in the opposite direction with equal force. Now, in the first situation the object made the one movement which the power of the moving force was capable of producing and it was necessary for it to proceed to the distance which the mover carried it. In the second situation, less distance was evident than in the first to the extent the other force dragged the moving object in the opposite direction. [402] In the third situation, where however much one motion drags forward, the other drags backward, the object is forced to remain in the same place. It does this, not as a completely motionless object which must remain thereafter in the same place, but because in the latter instance, it is not moved at all, whereas in the former, *it is* moved in two ways just as a swimmer striving against the current of a *river* is pulled in the opposite direction. This swimmer, if he has strength equal *to the violence* of the current, remains *always in the* same spot, not *as if motionless but because* he propels himself *forward* by his own effort *as much as he is carried* backward by the external force.

Since it is not wrong to explain an obscure point by *numerous* examples, let us *suppose that some bird in the air seems to stay in the same place.* Should it be said *that it is motionless as if it* happened to be *hung suspended* or that it is moved by its *own upward* motion to the same extent as *carried* downward by the weight of its *body? The latter seems to me to be the truest. Should you deprive it of the control of the brain or the pull of the muscles*, you *would quickly see it born down to* [403] earth. *It is* clear that the downward tendency, inherent in objects from their weight, is counterbalanced by the upward force of the power from the brain. In all similar *situations*, the object is carried alternately now downward, now upward, and, since the changes are rapid and sudden and the motions are carried out in the shortest *intervals, it seems to remain in the same place, held in one spot thereafter. This is not the fitting place to make this clear, for in the book of physics "On Motion", these points are examined more according to custom. It is enough for the present to have disclosed that there is some form of activity which is called tonic, or if you wish to call it something else, it will not matter.* It is better to know what it is than to think *that the muscles are inactive when the* arm is raised. *Indeed these four are all of the different motions seen in the muscles. They are shortened, or lengthened, or are carried about or remain taut. The fourth difference is of the same* [404] *kind as the first, for both are activities of the muscle.*

When a dead muscle, *which has none of* the influence of the brain [ψυχικος], has been cut across *entirely, you see it retract* toward *the ends. This would seem, not illogically, to be the work of the constitution of the muscular substance.* If the substance of the muscle is *intended by nature to retract to itself*, what need is there of the *psychic power* to move it unless the power *from the brain* is useful for making the *muscles give* way to each other's motions? If each carried on perpetually what it is designed by nature to perform, nothing would prevent the body from being afflicted by the disease called tetanos. What is tetanos but that the parts are drawn back involuntarily in opposite direc-

tions by the antagonistic muscles? If it did not happen thus, someone might equally well charge the power of the brain with seeming to command the muscles to lie in repose, whenever they must be moved by the *antagonistic muscles.* But if we agree upon this, we see immediately that it is contrary to what we said above. For if we say that the movement of the muscles is not by the psychic pull, [405] and that lack of movement is, we find that this contradicts many of the phenomena observed.

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Since it is not wrong to explain an obscure point by numerous examples, let us suppose that some bird in the air seems to stay in the same place. Should it be said that it is motionless as if it happened to be hung suspended or that it is moved by its own upward motion to the same extent as carried downward by the weight of its body? The latter seems to me to be the truest. Should you deprive it of the control of the brain or the pull of the muscles, you would quickly see it born down to [403] earth. It is clear that the downward tendency, inherent in objects from their weight, is counterbalanced by the upward force of the power from the brain. In all similar situations, the object is carried alternately now downward, now upward, and, since the changes are rapid and sudden and the motions are carried out in the shortest intervals, it seems to remain in the same place, held in one spot thereafter. This is not the fitting place to make this clear, for in the book of physics "On Motion", these points are examined more according to custom. It is enough for the present to have disclosed that there is some form of activity which is called tonic, or if you wish to call it something else, it will not matter. It is better to know what it is than to think that the muscles are inactive when the arm is raised. Indeed these four are all of the different motions seen in the muscles. They are shortened, or lengthened, or are carried about or remain taut. The fourth difference is of the same [404] kind as the first, for both are activities of the muscle.

When a dead muscle, which has none of the influence of the brain [PSYCHIKOS], has been cut across entirely, you see it retract toward the ends. This would seem, not illogically, to be the work of the constitution of the muscular substance. If the substance of the muscle is intended by nature to retract to itself, what need is there of the psychic power to move it unless the power from the brain is useful for making the muscles give way to each other's motions? If each carried on perpetually what it is designed by nature to perform, nothing would prevent the body from being afflicted by the disease called tetanos. What is tetanos but that the parts are drawn back involuntarily in opposite direc-

on the one hand, you cut one of the two thongs a little at a time but do not sever it completely, you will alter the articulation of the two bones definitely to one side of the middle position, but if you sever it completely, they will come nearly but not completely to the extreme position. For the extreme positions do not seem to arise except by drawing the thongs in the proper direction with the hands. [413]

The same manifestations are seen clearly with the muscles, the muscle being analogous to the thong, the brain to its movement by the hand. Without the hand, neither of the ropes can bring the articulation of the bone to an extreme position and without the impulse from the brain neither of the muscles will bring about either extreme flexion or complete extension of the part. If you deprive the muscles of the impulse from the brain, or the ropes of the hands, you will see arise the intermediate position of the bones. If you cut the external muscle you will see the part flexed beyond the middle position just as if you cut the external thong. So also when you cut the internal muscle, you will perceive that the part is extended beyond the middle position. What are the causes of these and all other conditions in the muscles? One principle especially is that the muscles have complete shortening in the extreme positions, just as in the example of the thongs. All the others [414] follow this.

Let us accept the principle on the spot, for one must take it not from some obscure hypothesis but from manifestations seen clearly in all the muscles. What is this principle? As has been said a little earlier, when the tendon has been cut from the head of the limb, the muscle shortens to an extent to which it shortens when the part is brought to complete flexion by deliberate choice. It was demonstrated clearly that the muscle seems disposed by nature to reach extreme shortening as a result of the properties of its substance. When it loses its attachment of the opposing bone, it is loosed from restraint and having become completely free, it shows its own nature. While the part is being pulled upon by the muscle arranged opposite, since they have the same nature in tending toward extreme contraction, each muscle is prevented equally by the other from retracting itself. Thus it happens [415] that each of the antagonistic muscles in the limbs is half hindered from shortening, for if both are disposed by nature always to strive toward complete shortening, and both are attached to opposite parts of the head of one bone, it would be necessary that the part, receiving independent motions of equal force, should follow neither. To follow neither would be the same as to hold the position midway between the extremes, for each of the latter arises when one of the muscles holds sway, extension when the external, flexion when the internal. Motion of equal power arises from the substance of the muscles themselves when neither one has the voluntary pull allied with it; motion of unequal power when only one has sway. So that it would be necessary for the shortening of that muscle to prevail which would be aided by the power from the brain.

We have found the causes of three phenomena, from having tried out one principle which we have taken, not from some hypothesis of ours but from visible manifestations. Since the muscles appear to undergo extreme shortening [416] when they are released from the binding tissue which secures them to the head of the limb, it becomes clear to us that they are disposed by nature to shorten completely but are hindered from it by something else. When we sought what the hindrance might be, we found immediately that it was the binding tissue. When this is cut they shorten to the extreme, and this fact we make the first cause of their not shortening. For this reason we see the muscles hindered from shortening by the binding tissue not simply because it is binding tissue, but because it is connected to the head of the bone pulled in the opposite direction. For this reason also, we shall be furnished with the causes of the second phenomenon, that is, that the member takes the middle position, when none of the muscles is moved by the power from the brain. A third phenomenon in addition to these is that flexion or extension of the part occurs when volition prevails with one muscle only; for by this prevailing the other muscle is forced to extend itself with

the limb approaches complete flexion more nearly than extension. If this be granted and stated, and it should be accepted, (for *it appears to be so*), it seems to me that one *must consider why, after extreme flexion of* the limb has been caused by impulse, the muscle, whenever we set it free, appears to *move but little and is lengthened to a very* slight extent. It was *not necessary for it to* move at all if the nature of its substance [409] is disposed toward shortening. Let whatever arguments we shall supply in favor of this lie out *in the open for con*templation by the lovers of truth, so that, if they appear to demonstrate correctly and are not to be controverted, we may say *everything has been discovered. Or, if not* quite everything, but the greatest part, then the things already investigated but still genuinely in doubt may be discovered at another time, one person finding one thing, another another.

In order that my discourse may be more clear, I need some example which not only will seem convincing to you but which you may set up for yourself if you desire. Take from whatever animal you wish two bones fastened together by a joint. *Having* twisted together several sinews into two thongs, either glue them down securely or bind them to those parts of the bones *which I shall indicate. I urge you to glue* or bind them on in the way that the muscles *attach to the bones. Since the manner* of the attachment for all muscles is double, *it is not improper for them to* be imitated in this respect by this example. One must now undertake to put into words clearly this double manner (of *attachment*), *for someone might not* [410] imitate it correctly if he had not understood it thoroughly, and it is most fitting for this to become the purpose of our discourse.

Of the bones attached to each other to form an articulation, one is moved about, while the other is placed over it like a *sheath in the manner seen* in the hinges of a door. Of necessity, therefore, the one remaining quiet has the hollow surface, the one moved, the *arched surface.* The hollow one is called the cup [KOTYLE] or the socket [GLENE], the arched one is the head [KEPHALOS] and knuckle [KONDYLOS]. When the socket is deeper, the knuckle is *more prominent*, for nature provides a place as well adapted as for a hinge. Since the moving parts are so well constructed, the parts responsible for the motion should *be much more skillfully constructed.* The instruments of motion, the muscles, take origin from the bones lying above which *have cups and they insert on the heads of the bones which lie below and which* are about to be moved. Accordingly, when the muscles contract, the heads [411] are drawn up and the whole limb is brought up *at the same time. Since one of them might* have to move a larger and another a smaller bone, the size of the moving muscle is made in proportion to the bulk of the bone *to be moved. Logically, some attach at the* heads or condyles themselves of the overlying bones, others attach a little lower than these, near the cup or socket, but not too much so, for thus it would become entirely too small and unable to move the bone lying below.

*Such is the nature of the bones articulat*ing with other bones and the muscles moving them. You will imitate it exactly if you fasten the thong to one of the bones at the part from which the muscle would originate, and attach the remaining part to the head of the other bone, as the muscle was attached, giving attention to these two particulars: one, that the thickness of the thong be sufficient to move and support the underlying bone; the other, that the thongs *not be pulled in the extreme position, but* lie as if attached to nothing in the world. Of the [412] thongs occupying the places of antagonistic muscles intended to extend and flex the part, one of the two is completely free of tension at the time when the part is in an extreme position, the exter*nal being free when there is complete* extension, the internal when there is flexion. After these preparations have been made, the *articulation will* move to *extreme extension or flexion* if one of the thongs is pulled by our hands. If these thongs are left to themselves, they will *bring about a middle position of the articu*lation of the bones and there they would remain at rest for the future. It is necessary to give *scrupulous attention* to this same position for it is exactly *midway* between extreme extension and flexion. If,

on the one hand, you cut one of the two thongs *a little at a time* but do not sever it completely, you will alter the articulation of the two bones definitely to one side of the middle position, but if you sever it completely, they will come nearly but not com*pletely to the extreme position*. For the extreme positions do not seem to arise except by drawing the thongs in the proper direction with the hands. [413]

The same manifestations are seen clearly with the muscles, the muscle being analogous to the thong, the brain to its movement by the hand. Without the hand, neither of the ropes *can bring the articulation* of the bone to an extreme position and without the impulse from the brain neither of the muscles will bring about either extreme flexion or complete extension of *the part. If you deprive* the muscles of the impulse from the brain, or the ropes of the hands, you will see arise the intermediate position of the bones. If you cut the external muscle you will see the part flexed beyond the middle position just as if you cut the external thong. So also when you cut the internal muscle, you will perceive that the part is extended beyond the middle position. What are the causes of these and all other conditions in the muscles? One principle especially is that the muscles have complete shortening in the extreme *positions, just as in the example of the* thongs. All the others [414] follow this.

Let us accept the principle on the spot, for one must take it not from some obscure hypothesis but from manifestations seen clearly in all the muscles. What *is this* principle? As has been said a little earlier, when the tendon has been cut from the head of the limb, the muscle shortens to an extent to which it shortens when the part is brought to complete flexion by deliberate choice. It was demonstrated clearly that the muscle seems disposed by nature to reach *extreme shortening as a result of* the properties of its substance. When it loses its attachment of the opposing bone, it is loosed from restraint and having become completely free, it shows its own nature. While the part is being pulled upon by the muscle arranged opposite, since they have the same nature in tending toward extreme *contraction*, each muscle is prevented equally by the other from retracting itself. Thus it happens [415] that each of the antagonistic muscles in the limbs is half hindered from shortening, for if both *are disposed by nature* always to strive toward complete shortening, and both are attached to opposite parts of the head of one bone, it would be necessary that the part, receiving independent *motions of equal force*, should follow neither. To follow neither would be the same as to hold the position midway between the extremes, for each of the latter arises when one of the muscles holds sway, *extension* when the external, flexion when the internal. Motion of equal power arises from the substance of the muscles themselves when neither one has the voluntary pull allied with it; motion of unequal power when only one has sway. So that it would be necessary for the shortening of that muscle to prevail which would be aided by the power from the brain.

We have found the causes of three phenomena, from having tried out one principle which we have taken, not from some hypothesis of ours but from visible manifestations. Since the muscles appear to undergo extreme shortening [416] when they are released from the binding tissue which secures them to the head of the limb, it becomes clear to us that they are *disposed by nature to shorten completely* but are hindered from it by something else. When we sought what the hindrance might be, we found immediately that it was the binding tissue. When this is cut they *shorten to the extreme, and this fact we* make the first cause of their not shortening. For this reason we see the muscles hindered from shortening by the binding tissue not simply because it is binding tissue, but because it *is connected to the* head of the bone pulled in the opposite direction. For this reason also, we shall be furnished with the causes of the second phenomenon, that is, that the member takes the middle position, when none of the muscles is moved by the power from the brain. A third phenomenon in addition to *these is that* flexion or extension of the part occurs when volition prevails with one muscle only; for by this prevailing the other muscle is forced to extend itself with

the entire limb. Now in addition to these, [417] let us state the causes of all the other phenomena, so that, if they agree with each other, we should trust to the arguments in favor of them, and if they should differ in one point, we should doubt all equally.

First let us consider the above-mentioned phenomenon that when the external muscle has been cut, the limb is flexed beyond the middle position, but not to the extreme. Perhaps it would seem equally logical for it to reach extreme shortening if no internal muscle pulled in the opposite direction. But he who judges thus would be forgetful of the weight of the limb which resists complete shortening of the muscle. In this regard events seem to take place in the same manner as with the thongs. If the external of these has been cut, the remaining one is not able to arrive at complete shortening, until the bone moved by it is detached. As long as there is articulation, it pulls the thong to itself. Whenever someone cuts across the internal muscle, the limb remains between the middle position [418] and complete extension, for the external muscle would be powerless to extend it without the impulse from the brain. And so, these phenomena agree with those stated above and with each other.

Chapter X

What Hippocrates wrote long ago concerning all the positions of the parts still seems to be true. But does it agree remarkably with the following? First, that we suffer pain if we flex a part completely or extend it extremely; second, that the position midway of these is free of pain; third, that we desire a quick change from the extreme positions; fourth, that we maintain the middle position longer without desiring change; fifth, that we even wish to change this sometimes; sixth, that all positions are hard for the extremely feeble to bear. The extreme positions logically are painful since one of the muscles operates to extend the other against its nature. The middle position [419] is justly agreeable since both muscles are resting not only from activity but also from excessive tension. Just as we need a change from a painful position quickly, we need a change from one not painful slowly. Why do we need a change at all from one not painful? Because even in this position the muscles are under some tension, but less than in all other positions. We say that it is painless, not as if entirely without pain, but as if the pain were very small and, from its slight intensity, nearly[1] insensible. We wish to change it when, having increased gradually, the pain becomes perceptable. That the muscles would have some tension in this position we have no need to explain, if we recall the previous discourse. We said that they are pulled by the member to which they are attached and for this reason, when freed of this tension by cutting the tendon, they readily adopt their natural shortening. No muscle will be without tension, not even in the middle position, but despising it as slight [420], we adopt it and avoid the others, not enduring them because excessive and irresistible. When we have become extremely feeble, as in gastric and cardiac syncope, we do not endure even the small tension of the middle position and for this reason, although we are able to move scarcely at all, nevertheless we move the parts from one side to the other wishing to find some painless position, but finally never being able to find one. Just so, if one of us had to carry some small stone suspended from his neck, being in good health, he could bear it without difficulty, but if he had become feeble, he would wish to reject it as a burden. In the same way if each muscle were vigorous, they would despise supporting an attached bone as one would a stone, not being aware of the slight addition to the weight, but when they are feeble, they are aware of it and carry it with difficulty, and hoping to shake off the load, pull themselves about from one position to the other. So all the manifestations in the muscles seem to agree [421] and seem to share in the common source, in which they are shown always tending of themselves toward complete shortening, but unable to reach this because they have the antagonistic muscles and the weight of the attached bones pulling against them.

BOOK TWO

Chapter I

Since the elementary principles of muscular motion have already been

pointed out, it is time for us to add the details which were omitted and thus complete our exposition. Then, if someone has absorbed all this information accurately, he will, if he wishes, be competent to carry on a discourse concerning muscle. Let us begin at the point where the first discussion ended, with the posture of the parts. Having first chosen the middle position we say that one of two conditions must exist in the muscles; one, mentioned above, [423] in which none of the antagonistic muscles are active and the other, about to be described, in which both are equally active. The first is present when we are at rest, as Hippocrates has said; the other, when we do not allow someone to extend or flex our limbs even if they use great force. The latter is what happens when the antagonistic muscles are in a state of activity called tonic, so that, if you should place the limb in a position midway of each side, there would be equal activity in both the muscles. When you move it to one of the extreme positions, however, either one muscle or the other is sufficient for such activity. It is clear that in each of the positions mentioned, the muscles employ tonic activity sometimes more and sometimes less, and thus the middle position relative to a particular action avoids either of the extremes. Now we say not merely that the middle position is without burden [ANODUNOS = painless], but that the muscles must be at rest in it. For, with tension in each of the muscles, the intermediate [424] position is as burdensome as the extremes. Since in the state of rest it may be either exactly in the middle position or not exactly so, the exact mean of all the extreme positions of the limb, not just the mean of each set of extremes alone, would be without sense of toil. As Hippocrates has said, none of the others would be strictly untiring.

Explanation of what has been said will become clear if we first distinguish the exact middle position from one not so. In this discussion the arm will be referred to particularly as a model. Therefore, there being four extreme positions in this limb, pronation, supination, extreme extension and flexion, the exact middle is the mean of all of these. It would not be exactly middle when only one of the two antitheses, either intermediate of full extension and flexion or of supination and pronation exists. Supination is the position of the hand when the hollow of this part is upward, the arch downward. [425] Pronation is the opposite of this. Midway of the two is when the hollow is inward, the arch outward, the little finger lying below the others; the ulna lying below the radius. This middle position can come about with the arm completely extended and completely flexed just as the middle position of the latter antagonists can exist with the hand supinated as well as pronated. The specific characteristic of the flexed position is that the ulna makes a right angle with the humerus and for this reason they call it the angular position.

Now the exact middle of the combination of both the above mentioned middle positions concerns us. Four other positions are all intermediate; they come not from the exact middle for the whole limb, but from one antagonism only, for each is midway of extension and flexion alone or of supination and pronation. Since each of them is double, linked in part to the extremes of the remaining antagonism, it is necessary that all four come about. [426] In each of them there is something common to all and proper to each. The common thing is for some one group of muscles to be fully active, while the other three are stretched incidentally, not being active. The thing proper to each is: of the pronated and angular, those muscles twisting about internal to the radius are active, all others being stretched passively; of the supinated and angular, those twisting about externally are active, all others being stretched passively; likewise also in the middle of pronation and supination with sharp extension only the extending muscles would be active and with flexion only the flexing muscles, all the others, idle, would be stretched.

Chapter II

For the discussion to be clear, it must first be known by which muscles, having what position and moving what articulation, these same four motions of the limb are executed. The BRACHION is the largest part of the whole arm. The boundaries of it below [427] are at the joint of the elbow;

"We call the elbow the part on which we prop ourselves," says Hippocrates. Above, likewise, it extends as far as the joint of the shoulder. The part of the arm next in size after the brachion is called the PECHYS [forearm]. It is bounded, where it is joined to the brachion, by the joint of the elbow; it *is joined to the wrist at the latter joint.* In the brachion there is one large rounded bone [humerus] *named the same as the* part. In the forearm there are two bones, one having the same *name as the whole* part, [PECHYS or ulna], the other is called KERKIS [radius]. They both articulate with the lower end of the humerus, the ulna with the middle of the condyle of the humerus; the radius, ending in a cup, surrounds the outer condyle and indeed, rotating on the latter as an axis, it guides the twistings of the hand. When it rotates inward the prone position of the hand follows, when outward, the supine position. *Extension and flexion of the arm is the* work of the joint of the ulna with the *humerus.* [428]

I do not know whether there exists such *an exact fitting together of bones as this* in any other joint. For, on the one hand, the lower end of the humerus, widening, is bounded by the condyles. The ulna, on the other hand, presents two crooked APOPHYSES [processes] opposite each other which make a hollow in between them resembling the letter sigma. *In this hollow* is embraced the middle part of the condyle of the humerus *in a manner* exactly like the parts of what we call a TROCHILIA [pulley]. When the hollow of the ulna is carried around the convexity of the humerus, extension and flexion of the whole limb take place. The brows of the middle hollow are responsible for the accurate maintenance of the joint and *for the failure* to be displaced to one side or the other, the processes of *the ulna* binding it tight. When the anterior process guides the *motion,* the limb is flexed, when the posterior, it is extended. The limit of flexion occurs when the anterior process seats against the humerus; of extension, when the posterior. Also, since the humerus [429] is arched and each of *the processes of* the ulna is elongated, there is more than ordinary danger *of* the bones coming against each other and hindering *the* motion of the limb. Nature has scooped out the bone of the humerus on each side to the depth that the processes of the ulna would extend down into the hollows in extreme extension and flexion. Since the posterior process of the ulna is greater, the corresponding hollow of the humerus is deeper. The part of the bone of the humerus between the hollows becomes thin because of their depth. Nature has avoided pene*trating it, thin as it is, in order that the joint* might not be completely loose and unstable nor the motions of the muscles excessive. If the bone were pierced it would be possible to flex the arm backward and whatever motions we might undertake with the arm in continued extension would be greatly exaggerated, and the pull of both muscles would be quite painful. If the posterior muscles were shortened by the power of the brain more than they were intended by *nature, the [430] anterior muscles would* be stretched and be in danger of being ruptured. Such is the skill of nature in regard to the precision of this joint.

Chapter III

Let us discuss next how the arrangement of the muscles already mentioned is related to the movements of the joint. Two muscles have their origins in the anterior part of the arm and two in the posterior; their insertions are into the ulna through *strong aponeuroses. The largest of them* begins at the head of the humerus, the smallest much lower, and they all four run straight to the ulna and attach mainly at the same place as the foundation of the bony processes. The very posterior process of the elbow, which the Athenians call the olekranon and the *Dorians the* KUBITOS, arises at the same part as that to which the *coronoid process is attached in front. These* muscles, both those anterior and those posterior to the humerus, act on the ulna to produce extension and flexion. Four other muscles have their origin from the ulna on each side [431] of the most bent part of the elbow, two externally and two internally. *All are oblique to the bone of the radius,* and they attach, the larger at the lower *part near the wrist joint, the smaller at the* middle part. When these muscles pull toward their origins, the radius follows

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Chapter IV

Not even during sleep are all the muscles completely in repose. Only through drunkenness, weariness, or poor health are all the parts of the body completely relaxed and the muscles inclined toward the middle position, and entirely at rest. Someone asleep cannot have any part in the extreme positions, because these require strong, robust, and intense activity of the muscles. Ordinarily we sleep in positions which are intermediate between the extremes and exactly in the middle. For if you place the member in any of these positions, you rely on the muscles in tonic activity around it to maintain it, so that many times individuals may go to sleep either sitting up or walking about. Indeed, at first hearing this, I put no faith in it, but after being obliged to walk

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throughout the whole night, [436] I learned the truth by experience, and had to believe it. I walked nearly a whole stadion asleep and, absorbed in a dream, did not waken until I stumbled upon a stone. Indeed, the lack of a tendency of those asleep to walk farther is the result of their inability to happen upon a smooth path. Although this is believable only to someone who has experienced it, sitting up while asleep may be seen by everyone every day. Also, extremely few of those lying down have each of the limbs exactly in the middle position. Those who hold something in their hands while asleep demonstrate the tonic activity especially well, for their fingers often remain carefully flexed around a small object, a piece of gold, a stone, or a coin.

Is it not clear also what happens in dropping of the lower jaw? For it only separates from the upper if someone goes to sleep drunk, or very lazy, or extremely weary. And actual snoring, for the same reason, usually occurs from the slackening downward of the jaw in a man remaining [437] in a supine position. Indeed, this very remaining supine is a sign of being relaxed. Moreover, Hippocrates found fault with either lying supine and sleeping with the mouth open, but he approved of lying on either side. How much activity there is in those lying on their sides you would perceive best if you placed the body of a dead man in that position. It will not remain even a short time, but will immediately become either supine or prone because inevitably it would slip from its weight. Indeed lying supine and snoring with the mouth open are signs of slackness, drunkenness, or laxiness. For this reason, again, Hippocrates recommends putting all the parts in painless positions for operating on dislocations and fractures, and "For operating on the lower jaw," he says, "when the man gapes moderately." What the angular posture is to the arm, moderate gaping is to the lower jaw, for this is the middle position. The extreme positions of the lower jaw are with it most widely opened and with the teeth [438] set against each other. The former of these is performed by the muscles in the chin and the neck pulling downward; the latter is performed by the muscles inside the jaw, those from the palate, those called temporal, and probably also the muscles at the sides of the lower jaw called masseters assist the action to some extent; but we discuss this elsewhere. Gaping moderately is the middle position when all the muscles enumerated are in repose. In those dying, moreover, the jaw assumes this position and with reason, for in them all the muscles are deprived of activity. Likewise, it is evident that when someone sleeps without gaping, the activity of the muscles raising the jaw is maintained. Many sleep with their arms or legs extended or flexed, thus preserving the tonic activity in their muscles.

But I speak at too great length, while I might have mentioned what is most readily at hand. For example, would anyone contend that no effort is contributed when the muscles guard against the outflow of the excretions? [439] Strong muscles are placed at the ends of their conduits like guardians at some gates, not allowing them to be evacuated before reasoning power commands. During sleep these muscles perform their task faultlessly. If one of the excretions is released involuntarily, it happens either that these muscles are paralyzed, or reason suffers as in phrenitis, or both reason and muscles are depressed, as in drunkenness; for surely, either the reason must not be in control, or the muscles be without power, or both be disabled at the same time.

Let us not be rash and deny that the brain is at rest during sleep unless we mean that rest is not complete stillness; so let us rather call it a cessation of intense application. If someone says this, they say well and we agree with them. For those sleeping are not entirely without sensation, although they sense with difficulty. Otherwise how would they hear someone calling, or arise when a light is brought in, or feel someone touching them? If you remind me that the drunkard who does not know where in the world he is, [440] and who sleeps more heavily than the torpor of Epimenides, was close to stupor before going to sleep, then I say it would be better for you to turn to watching some vigorous man asleep or someone awakening, since in them many of the activities of the brain are preserved.

Chapter V

The argument is unreliable indeed of those who say that all the activities of someone asleep and stupefied are natural [i.e., without benefit of the brain], as this is not entirely true. For example, why do they shift and move their limbs in many ways? Why *do* they utter sounds while sleeping? Do they say that these acts are natural? But we do not agree that they take place by intention, someone may say, for you maintain intentionally neither the continuous motion of your eyelids, nor the motions of all the parts when you speak publically, use oratory, converse, nor, when you walk to Athens from Peiraieus, do you apply yourself to the activities of your legs in particular. Often while deep in thought people fail to notice that they have come to the end of their journey [441] and go past the place toward which they started in the beginning. Did the walking come about *neither as the work of the brain nor* by voluntary impulse? In like manner, it seems to me that walking while absorbed or while asleep involves the motion of the moving parts on the one hand and the tonic activity of the unmoving parts on the other. Consequently, what reason would you give for those awake often not paying attention to the activities of a part of the body, and why not apply this same reason to those sleeping and stupefied and, at the same time, why *not wonder how much came* about voluntarily? When unable to find the cause readily, would it not be rash to say that none of these acts occur voluntarily? *If no truer criterion* than this can be found for deciding whether the acts are by deliberate choice or by nature, why must we not demonstrate something else concerning them? It is more appropriate to become *skeptical in these* matters than be rashly dogmatic. Since we have a most obvious criterion for the voluntary acts, taking courage, let us give evidence not only for each example already mentioned, but also [442] show that respiration itself arises voluntarily, as far as it would *seem to fall* under the criterion.

What is it now by which we judge the voluntary acts? I propose to give you many, not one *criterion, all* agreeing with each other. If, choosing deliberately, you can *stop actions* taking place and not perform them, they are voluntary. If, in addition to this, you have the power to execute them more quickly or more slowly or more often or more seldom is it not in manifold ways evident that the act is the subject of deliberate choice? Deliberate choice cannot stop nor initiate, nor make more frequent, nor more seldom, nor more rapid the motion of the arteries and the heart. For this reason, these acts appear not to be of *the brain, but happen through nature.* The power of reasoning makes the motion of the legs consistent with all these, for one can act to stop the motion happening, activate it again when quiet, and make it faster, and slower, and more seldom and more frequent. These same things occur in the case of the motions of respiration, an activity [443] of the diaphragm and muscles of the thorax, as we demonstrate in the work, "Concerning the Causes of Respiration", *they are an activity of the brain, not* of nature, since to move the muscles is an activity of the brain.

It is inappropriate to discard the things that are well known simply because we lack knowledge of their causes. Indeed, the criteria of the activities by deliberate choice obviously are known, but we overlook the cause in those instances in which we do not follow the individual acts closely. The stupid person, disbelieving what is *clearly visible, the gullible one, being at a* loss concerning the things readily displayed, and the one suspicious of the known because something is unknown, are among the lovers of doubt. Anyone would be not only suspicious, but also extremely stupid if he disregards the known because of the unknown. Now let us not willingly acknowledge ourselves to be without sense, nor admire the doubtful, nor be stupid, nor any other of these things, but, because it is right *and is befitting moderate* men, let us accept the obvious readily, and inquire [444] into the dubious at leisure. Although it is clear that volition dominates *respiration, it is difficult* to understand why we do not follow many other voluntary activities closely with our intelligence. Having established our obvius knowledge, let us now pass to an investigation of

the cause, without being drawn into it completely nor as yet pretending to discover the true cause, since the things already said may be more plausible. No one has found the cause, I maintain, but some writers reporting only the search for the difficulties, which I have just now finished discussing, assume that they have discovered the cause. And indeed we must accept them favorably, whether they find or whether they strive eagerly to find, but it is necessary, also, for us not to shrink from investigating the remainder.

Chapter VI

In our search for these causes, we shall take it as our general background, that frequently many men, after passing through some achievements, a little later forget them completely. Similarly, those beset with fear or through drunkenness or some such cause, remember nothing of what they went through while in these conditions. [445] It seems to me the reason is that they have not applied themselves attentively with all their mind [NOUS, not PSYCHE] to the experiences. For the imaginative part of the brain [PSYCHE], whatever it might be, is like this very remembering itself. If it should receive the impressions of objects clearly in these fantasies, it would preserve them always, and this is to remember. If it should receive them indistinctly and superficially, it would not preserve them and this is to forget. And for this reason, in anger and in deep meditation, drunkenness, phrenitis, fear, and very stubborn sufferings of the brain, none of them would remember their achievements at a later time. Indeed, is it extraordinary in those asleep, the brain operating indistinctly, for the impressions to become indistinct and for this reason not to be lasting? If those keeping watch or pondering some problem have almost all their attention directed toward their thoughts, scarcely having any of it concerned with their walking, is it extraordinary that the impression of the performance is received indistinctly and is forgotten forthwith, and that these individuals, therefore, do not remember whether the act was accomplished voluntarily [446]. Likewise, if we do not remember completely, we cannot recall accurately anything that happened, and therefore we do not know what the things were which we do not remember, because it is necessary to keep them in the memory first, in order to observe their quality afterward. At any rate, to me it does not appear extraordinary that, although breathing during sleep is voluntary, when we arise, we cannot say whether we breathed voluntarily, and it would be the same if those moving their feet and hands and uttering sounds while asleep should forget and say the motion of the limbs and the voice arose involuntarily.

Patients in delirium prattle, walk about, and perform all the voluntary motions, but when they recover from their PARANOIA, they remember nothing of what they have done. At any rate, for 13 days I saw someone in a delirium carry on as follows: He thought he lay sick in Athens, not in Rome. Continually calling his accustomed boy, he ordered him [447] to carry the necessary things to the gymnasium, and a little later, "Oh you," he said, "to the Ptolemaion, I say, for I wish to be free in it for a long time." And then sometime afterward he leaped up and, donning his clothes, would straightway have gone forth from the outer court door, but those within restrained him and prevented him from going out. "For what reason do you prevent me?" he demanded of them. They told him (for there was nothing to say but the truth) that he had had and was still having a fever. He replied to this altogether moderately for, he said, he knew that some of his fever remained, that this was something quite small and nothing suspicious to hinder him from bathing, for the whole fever affecting him had arisen from travel. "Do you not remember," he said turning to his boy, "how violently we traveled yesterday from Megara to Athens?" While he was speaking thus and doing these things, a massive hemorrhage through his nose overtook him and after this a sweat. He recovered forthwith but remembered nothing of the foregoing. Were not among his voluntary acts (for this is what we proposed to show from the beginning) [448] the standing up, speaking aloud, defecating or urinating (for the man accomplished all these things during those many days)? Or, is it indeed absurd (for if these acts are not voluntary,

no others are) for him not to remember them, when those sober do not remember what they performed in drunkenness?

If it is true that breathing arises voluntarily, how is it extraordinary that if we pay attention more carefully at one time, but more carelessly and *lazily at another*, we remember the activities occurring while we were intent, but otherwise we forget them? Also, since we consider that we have not accomplished what we may have forgotten completely, we do not remember if it was accomplished voluntarily. How the act of breathing may be controlled voluntarily wholly through the brain is shown by the foreign slave who had been made angry and *resolved to die. Throwing* himself to the ground and [449] holding his breath, he was motionless for a long time; later, rolling over slowly, he died. Even if it were impossible to hold the breath continuously, no one would deny that it is voluntary. Thus, it seems that some voluntary *acts are free and some are subject to the* exigencies of the body. The former we perform free of hindrance at all times, the latter only at certain times and to a limited extent. Thus, walking toward something, talking, or taking and receiving something are completely free; but defecating and urinating are responses to the urgencies of the body. Now someone may be silent for a whole year or a longer time if he wishes; but holding the feces or the urine is not carried out in a similar manner because no one can do so for a month, not even for a few days. There would be pressure, often with distress, or the weight of fullness or a stinging with such sharpness that [450] there is not time enough to reach the accustomed place. Breathing is the same as these but much more pressing with more immediate necessity, for there is danger of death without breathing. It is exceedingly painful to be choked and it is not extraordinary that holding the breath is extremely difficult. All people are zealous not to die, but should they be afflicted by a myriad evils, and wish to die, they would not hasten into this, unless they wished to go out painfully. Simply because we are able to remain continuously silent but are unable *to hold* our breath, however, no one should consider that only the speaking is a voluntary act and breathing an involuntary and natural one. I think I have demonstrated this clearly.

Chapter VII

What has been left incomplete in our discourse concerning the middle positions should be added next. Since the so-called angular posture of the arm, being precisely middle between full extension and flexion, is most free of burden, it seems likely that a similar condition would hold true for the legs, but this is not so. The painless position without burden in the latter members is in the interval between the middle position and full extension. The reason for this involves habit, for ordinarily our legs are extended because we hold the whole body up by them when we stand or walk. Also before we are active with them, they are molded in extension by swaddling clothes. Also while sleeping and reclining for other reasons, they are extended more often than flexed. We are distressed much more in extremes of flexion than extension. Many people cannot bring their legs entirely to extreme flexion without assisting with their hands. We are unaccustomed to making this part go through this action and only those flex easily who are adept in dancing or expert in wrestling and especially accustomed to flexing completely. This intermediate position approaching extension which is completely without burden, was caused by custom of the parts over a long period of time.

These two things, nature and habit, are to be considered in all the joints, [452] if you would find the intermediate and burdenless position. It seems that everywhere habit corresponds to nature and the intermediate burdenless position is rightly called newly acquired nature. In the legs, this same intermediate position without burden would be the middle of the motions taking place in the parts and not the middle of complete extension and flexion. For you will find that we withdraw from the position without burden toward extension as much as we depart from flexing to the uttermost. In general, therefore, you will discover the intermediate and burdenless position from examining the uttermost motions in all the joints. Thus, in the joint of the elbow it is the so-called angular

position, in the knee that closer to extension, in the spine that closer to flexion and for the joint of the *wrist, the exactly* straight. We can make the spine quite fully arched or flexed. We bend the head backward *and forward from upright equally* so that the [453] precisely straight position is exactly middle between these extreme *motions*, and is the most without burden. In the spine the intermediate position is not the straight but that slightly curved, for this motion predominates. We exert ourselves more in the region of the spine when standing upright than when sitting or lying down, for, in standing, the spine must be *extended but in sitting or lying down it* is not hindered from being flexed. If you examine this in all the joints you will find *that they all make our exposition* consistent within itself.

With the muscles not associated with joints, an intermediate condition would be without burden, as with the anus, the bladder, and the tongue. For, both closing the anus extremely tightly and dilating it to the utmost are burdensome; also, protruding the tongue to the greatest extent, or flexing it, or however many other motions are carried out excessively, are burdensome. It is easy, indeed, to discover the intermediate of the extremes for the muscles of *the* tongue, as well as what is burdenless. All men, at the time when they cease their life-long activities, maintain *an intermediate* and burdenless posture in all the parts [454], "having been compelled to this by a well-ordered nature," says *Hippocrates. In the tongue* all the muscles attach in pairs, above and below, at the right and at the left so that it is *not extraordinary that it is moved* in opposite directions by antagonists. From the facts at hand, however, it is not equally *clear how the muscle of* the anus, the bladder, and the diaphragm may be moved in opposite directions (for the single muscle *in each of these is circular, no muscle* apparently being antagonistic), but let us look into this.

Chapter VIII

The action of the muscle of the bladder and the anus is *not to eliminate* the excretions of the food but to retain them. Many have been mistaken straight off in this very thing, believing that these muscles were created for the sake of expelling the excreta, because they have not been able to observe that after paralysis, elimination takes place involuntarily. When the muscle *in the anus* has been cut many times [455] excessively through wretched surgery, the feces flow by it involuntarily, as if, I think, the organs which prevent the flowing did not remain. *This muscle is strictly* not the primary organ of expulsion, but in order that what happens when it is cut or paralyzed *may not occur continually* throughout life, nature has placed it as a guard against the ill-timed exodus of the excreta. *We turn our attention, therefore, not* toward something to prevent the expulsion, but rather toward the structures *accomplishing this activity.*

What, therefore, are the organs of this expulsive *activity?* They are many as *individuals but of two kinds, those of the brain* [PSYCHE] and those of nature [PHYSIS], and, therefore, the organs of the brain always *act voluntarily, those of nature involuntarily.* The diaphragm and all the muscles of the epigastrium are organs of the brain. *The whole system of the intestines and the* stomach are organs of nature, but the activity of these has been written about in *other books. Let us speak now concerning* the muscles, since the present discussion is an explanation of their motion. [456] All *the muscles in the epigastrium, when they* pull actively, press into the organs of nature inside. These, if the diaphragm *yields, go back up into the latter's place,* giving way before the force of the muscles. If the diaphragm does the opposite, the *contents of their cavities would be pressed* out as if squeezed by two hands, outside by the muscles and inside by the diaphragm. *The obliquity of the diaphragm aids this* greatly, since it has one end at the cartilage in the breast lying above, the other behind *at the lumbar spine.* The *compression of* the excreta in the intestines takes place by the activity of the muscles on two sides, outside by those of the epigastrium, inside by the diaphragm. The obliquity of the diaphragm is *the cause of the* descent of the compressed. The muscle in the anus is idle during this time. The muscles in the abdomen *are numerous, and all* contract

during these emptyings; those in the hypochondrium contract more, those below less, contrary to what takes place in urination. For in the latter those below act more, [457] those in the hypochondrium less. both, not as organs of urination or defecation for this is extremely illogical, but, since the pull of the diaphragm must be as strong as the pull of the muscles in the abdomen, it would be impossible for one muscle by itself to compete with the many large ones. The danger in this is that if they overpowered the diaphragm everything would be forced into the cavity of the thorax. For this reason the muscles between the ribs contract, binding them together from all sides. The lax thorax easily yields to the pushing diaphragm, as it is possible to observe when the muscles in the abdomen contact, and especially the inferior ones, while all those inside the thorax relax. Almost the whole content of the abdomen, in these conditions, is pushed into the cavity of the thorax along with the diaphragm itself. In order that this may not happen and the act of defecation be abolished, the entire thorax is held securely fast on all sides. It is evident from all that has been said [458] that the muscles placed at the outlet for the feces are destined to shut them in. This is their proper action, and they cannot expel them except inadvertently when they cease acting. Just as opposing muscles promote opposite actions in all other regions, so it is here, for retention of the feces comes from the activity of the latter muscles, and expulsion from the muscles in the epigastrium and diaphragm.

Chapter IX

Which muscles in the diaphragm have analogy to the antagonistic muscles of other parts? The antagonistic muscles cannot be stated simply because first, as an organ for expulsion of feces, it has those retaining them, and second, especially, in another kind of antagonism, it has those of the epigastrium. As an organ of respiration the latter antagonism is in part true and in part not entirely true, for in expiration no muscle is wholly agent, since this action of the thorax, it seems, is more like the passive condition of falling [459] mentioned by us earlier. Blowing out is a massive carrying out of the air caused by the activity of the intercostal muscles. All the muscles between the ribs inside [internal intercostals] are agents for blowing out. The former is like the action of the thorax in making someone lie down with respect to his whole body already mentioned, the latter is like the laying down of each of the parts. Inspiration is opposite to expiration, blowing out to violet inspiration (for it does not have a specific name). The diaphragm alone presides over the earlier antithesis, over the other antithesis the intercostals with the muscles in the thorax coming down from the scapula and the neck preside. Especially in need of this are flute players, trumpeters, and heralds when they are about to call out their monotonous cries, and, not least, those blowing into leather wine skins and other organs or, to speak simply, anyone who wishes particularly to make expanded and contracted changes in the thorax. Someone might rightly say that the external muscles of the thorax are [460] antagonistic to the internal intercostals. These things concerning the muscles of the thorax are pointed out not only in this treatise but also in the treatise "On the Causes of Respiration," and "On the Voice," and "On the Purpose of Respiration." But now indeed the continuity of the discussion is at an end.

The diaphragm has certain peculiarities different from the other muscles due to its situation and form. When it ceases to act and becomes more lax, its convexity is inclined toward the spine on the one hand and the stomach on the other, but much more easily toward the spine. In all of a man's positions except lying prone, the diaphragm is above and the spine below, so that logically it is inclined toward the latter because it is under pressure from the abdominal viscera in front, from the softest and most nimble of all the splanchna, the lungs, behind. [461] Its convexity inclines forward, however, in the prone position, and when the intercostal muscles are active, those in the paunch are in repose and it is evident that the abdomen under these circumstances raises itself. Gymnasts seek this position habitually after exercise. Just as the intercostal mus-

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cles are active so also are those in the abdomen, during what they call holding the breath. For this it is necessary that the upper end of the larynx be closed, since if it is open when the muscles mentioned are active, there is blowing out. If again, with them, the muscles of the pharynx and larynx extend, there is not blowing out but the voice comes forth.

A double tension exists for all the muscles. The first is when, being active, they pull themselves together, the second is when they are extended by the antagonistic muscles. The first holds true for the diaphragm in non-violent inspirations. The other has a double origin, as has been said above, either when the muscles in the belly are active or only the intercostals alone. [462] In non-violent expirations which specifically we call expirations to distinguish them from blowings out, neither of them holds. But in all others, that condition which we say comes in the middle of the extreme motions, only in the case of the diaphragm is double. It inclines toward the spine in all positions but the prone, in which position only it inclines toward the stomach.

The intercostal muscles and those in the abdomen are forever curved out due to their form having been made like the underlying organs. When they are in repose, naturally they are so. When they are active they retract and become less convex while nearly all the other muscles, those which move the parts, are the opposite. The latter, being straight in repose, become convex when they are active. The reason for their difference is clear. For, since the substance of the bones, hard and rigid, is placed under some of them, and some hollow yielding space under the others, logically all those lying on the bones [463] during contraction gain as much in depth and width as they lose in length and hold prominent the whole bulk of their body. In those which conceal the softness of their background, the greater part of their bodies is hidden away during contraction. Indeed, it is not extraordinary, if nearly all the muscles in the limbs curve out when they are active, that those in the thorax and epigastrium only retract, for they alone have the yielding hollow space underneath. Certainly when the stomach is gorged so much that it is painfully distended, they do not retract. For, what happens with the others at all times, happens in those of the epigastrium in gorging of the stomach due to the rigidity of the underlying background. The latter clearly is filled by engorgings, hydropsy or in whoever is fat. In those with the stomach empty, the muscles, before they become active, are curved in resemblance to the underlying organs, for they are pushed out according to the curving of the latter, but when they become active they retract for they easily compress [464] the underlying cavity. And now also the muscles in the thorax, attached in between the boney parts, before they are active, have a form similar to the whole ribs, being convex outward and concave inward. When they are active, compressing first and especially and the membrane lying under them called the pleura and through it the lungs which are soft and spongey, they give way inward to the extent that the yielding substance of the underlying organs permits.

Anyone knowning these generalities concerning the motion of muscles can discover everything else in individual parts.

DISCUSSION

The source of error which has caused the misunderstanding of Galen's comprehension of nerves and tendons stems from the early Latin translations. The able scholars of the Renaissance, because their knowledge of anatomy was greatly inferior to their knowledge of Greek, decided that the Greek work SYNDESMOS should be translated ligamentum in Latin regardless of the context. It has been the tradition through the succeeding centuries to Latinize the Greek, and for the most part modern translators have slavishly followed the Latin by translating the word ligament. In this treatise Galen, being an extraordinarily accurate anatomical observer, hoped to avoid this mistake by giving a detailed definition and explanation of his use of SYNDESMOS. It seems clear that he understood fibrous connective tissue to range from the delicate areolar tissue of the endomysium to the dense bands and sheets of ligaments and aponeuroses. He

even points out that the name SYNDESMOS *follows the meaning*. The derivation of the word is from SYN, meaning with, the DESMOS meaning something that binds. *In a similar way*, electron microscopists use desmosome for a binding body.

At the end of the first paragraph of the *translation there* is a *key example* of the subtlety of the shades of meaning in Greek and the stultifying influence of *Latin translation. In Greek the* SYNDESMOS "is a nerve-like substance associated with bone, etc." The Latin renders this "*attached to bone," the significance of the* entire passage is lost, and the interpretation as ligament is firmly established.

No less a historian of Medicine than Charles Singer, in discussing Galen's "false concepts," makes the following *tatement in the introduction to his trans-*ation of "On Anatomical Procedures": Even greater difficulty arises from his *heory of the nature of nerves. Galen saw* hat a nerve, NEURON, passes into each nuscle and that it then divides. Knowing that many muscles end *in a whitish ten*-don, he thought that the branches of the nerves had reunited within the muscle to form this tendon, which he naturally also called NEURON." This undoubtedly refers to the second chapter of the present treatise, but it is a deliberately false interpretation, even to the statement that Galen called tendons nerves, which he certainly never did. He was much averse to loose ends in any description or explanation. He traced the nerves into the endomysium, and not having a microscope with which to see the *motor end plates, it was logical, as he says,* to carry them along with the connective tissue as it became condensed into the ten-*don.*

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Central Projections of Spinal Dorsal Roots in the Monkey [1]

I. CERVICAL AND UPPER THORACIC DORSAL ROOTS

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ABSTRACT A study of the central distribution of cervical and upper thoracic dorsal roots (DR) was made in 18 monkeys subjected to 31 dorsal rhizotomies. Degeneration was studied in multiple spinal and medullary sections stained by the Nauta technic.

Dorsal roots C2 through T7 project fibers to: (1) portions of laminae III and IV, (2) the intermediomedial nucleus, and (3) portions of laminae VII, VIII and IX. DR C5 through T1 project fibers to medial parts of lamina VI. DR fibers reach cell groups of lamina IX via different trajectories, one of which is dominant in the brachial enlargement. Fibers project to the spinal accessory nucleus from C2 through C5 DR, while portions of the dorsal nucleus receive fibers from C5 to T7 DR.

Ascending fibers in the fasciculus cuneatus: (1) arise from C1 through T7 DR, (2) exhibit only general segregation due to overlap, and (3) project somatotopically throughout the extent of the cuneate (CN) and accessory cuneate (ACN) nuclei. Fiber terminations in CN and ACN are: (1) similar for C1 through T1 DR, and (2) distinctive for T2 through T7 DR. The upper five cervical DR project fibers to: (1) the supraspinal nucleus, and (2) both the central cervical and intermediate nuclei. Cytological and degeneration studies suggest that the central cervical and intermediate nuclei constitute parts of the same cell column.

Although many studies of the central projections of spinal dorsal root fibers have been made, the majority of these investigations has utilized the Marchi method (Sherrington, 1893; Collier and Buzzard, '03; Ranson, Davenport and Doles, '32; Pass, '33; Corbin and Hinsey, '35; Ferraro and Barrera, '35b; Corbin, Lhamon and Petit, '37; Schimert, '39; Yee and Corbin, '39; Walker and Weaver, '42; Chang and Ruch, '47; Escolar, '48). More recent studies of dorsal root fibers (Torvik, '56; Liu, '56; Sprague, '58; Grant and Rexed, '58; Kerr, '61; Kuypers and Tuerk, '64; Sprague and Ha, '64; Hand, '66; Sterling and Kuypers, '67) employing silver staining methods have provided new data concerning: (1) the distribution of these fibers in the spinal gray, and (2) their mode and patterns of termination in medullary nuclei. Most investigations utilizing the Nauta staining method have been done on the cat and have been based upon section of a limited number of dorsal roots in restricted regions of the spinal cord. A notable exception is Liu's ('56) study of dorsal root afferents to the dorsal and accessory cuneate nuclei in the cat.

The purpose of this study of spinal dorsal roots in the monkey was to determine: (1) the distribution of these fibers within the spinal cord, and (2) the projection and patterns of termination of ascending fibers in medullary relay nuclei.

Material from this study has been divided into two parts. Part I concerns the central projections of the cervical and upper thoracic (i.e., C1 through T7) dorsal roots; ascending branches of these dorsal roots project primarily upon the cuneate and accessory cuneate nuclei. Part II of this study (Carpenter, Stein and Shriver, '68) presents similar data concerning the lower thoracic, lumbosacral and coccygeal dorsal roots and includes a comparison of projections to the posterior column nuclei.

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MATERIAL AND METHODS

In 18 rhesus monkeys single dorsal rhizotomies were done at different levels and on different sides. For economy of time, effort and material three or four different single dorsal roots at selected levels were sectioned in some animals. Table 1 indicates the single dorsal roots sectioned in individual animals. Although this study concerns only the cervical and upper thoracic dorsal roots, dorsal roots at other levels were sectioned in some animals (see Part II).

Surgical procedures were performed aseptically. Following rather extensive bilateral laminectomies, the dura was opened in the midline and the root to be sectioned was identified. Dorsal roots were sectioned with microsurgical scissors with the aid of the Zeiss operating microscope. Using various magnifications, it was possible to section the dorsal roots without traumatizing the spinal cord or interrupting radicular vessels. The proximal stump of the severed dorsal root was marked with a black silk suture. Following dorsal rhizotomies the dura was closed with 5-0 silk sutures, except in thoracic regions where the dural defect was covered with gelfoam.

In the thoracic region the narrow confines of the spinal canal made it difficult to close the dura and exposed portions of the spinal cord sometimes revealed evidence of trauma postoperatively. In order to avoid this, certain thoracic dorsal roots were sectioned within the dural sleeve following laminectomy. Dorsal and ventral roots were separated, and the dorsal root was cut without injury to radicular vessels.

In certain prior studies of dorsal root fibers (Liu, '56; Sprague, '58; Sprague and Ha, '64; Hand, '66) extradural section of the dorsal root was done in the intervertebral foramen. This technic was preferred by these authors because spinal cord injury frequently followed intraspinal dorsal rhizotomy. Experience from this study indicates that intralaminar dorsal rhizotomy in the monkey can be done using microsurgical technic without injury to the spinal cord or radicular vessels.

Animals were observed postoperatively in single cages for at least two weeks. Neurological examinations of each animal were made on the sixth postoperative day and prior to sacrifice. Animals were anesthetized and sacrificed by perfusion; each animal was perfused via the left ventricle

TABLE 1

Dorsal roots sectioned in eighteen monkeys

Animal Number	Spinal roots																										
	C1	C2	C3	C4	C5	C6	C7	C8	T1	T2	T3	T4	T5	T6	T7	T8	T10	T12	L1	L2	L3	L4	L5	L6	L7	S1	Co1
C-879					L																						
C-912			R						L										R							L	
C-913*		R								L										R							
C-914					R			L													R				L		
C-915*							R								L							R					
C-916	R																L							R			
C-967						R					L																
C-970*		R										L				R											
C-974					R																						
C-978*			R																								
C-981		R		L																							
C-1006		R		L																							
C-1008	B									R																	
C-1012						L												L					R				
C-1014					L			R																L			R
C-1017														L		R											
C-1024		L											R														
C-1025									L						R												

*In 18 animals 45 dorsal roots were sectioned individually on different sides and at different levels. Letters R, L and B refer to right, left and bilateral and indicate roots sectioned. Asterisks indicate that sections of the medulla were cut in horizontal planes, all others were cut transversely. Only observations on cervical and upper thoracic dorsal roots (C1 through T7) are reported here, data concerning all other dorsal roots are considered in Part II of this study.

f the heart with a liter of normal saline ollowed by an equal volume of chilled 0% neutral formalin. The brain and entire pinal cord were removed *in toto*. After dequate fixation in 10% neutral formalin he brains were sectioned. The brain stems vere cut in transverse and horizontal lanes (see table 1). Levels of the spinal ord were determined by counting spinal oots under a dissecting microscope. Transverse blocks of the spinal cord were cut and placed in separate compartments of plastic boxes.

Blocks of the brain stem and spinal cord were sectioned at 20 μ on a freezing microtome. Multiple sections of the medulla and representative sections of the spinal cord above, below, and at the level of the rhizotomy were stained by the Laidlaw modification of the Nauta and Gygax ('54) technic. Representative sections of the medulla and numerous spinal levels also were stained with cresyl violet to facilitate identification of cell groups. Drawings of the spinal cord and brain stem sections and the resulting degeneration were made for all animals.

The lower brain stems of five animals were embedded in paraffin and sectioned serially at 15 μ in horizontal (3 animals) and transverse (2 animals) planes. Every ninth section was stained with cresyl violet and every tenth section was stained according to the Weil technic. These slides were projected and nuclear groups in the medulla were outlined in serial drawings. This control material facilitated the study of the medullary nuclei receiving ascending fibers from the spinal dorsal roots.

OBSERVATIONS

Normal anatomy. The cytoarchitectonic atlas of Rexed ('52, '54, '64) and his nomenclature for cell groups of the spinal cord have been used in describing the degeneration resulting from dorsal rhizotomy. While it is realized that these studies are based on the cat, there appears to be a fairly close general correspondence for the monkey. The use of Rexed's terminology also provides a basis for more precise description. There are several nuclear groups which deserve special attention and comment.

Evidence from the study of normal cytoarchitecture in the monkey indicated that lamina II corresponds exactly with the substantia gelatinosa as described by Rexed ('52) and as shown by the ultrastructural studies of Ralston ('65). These observations differed from those of Sprague and Ha ('64) and Szentágothai ('64) in the cat who regarded the substantia gelatinosa as composed of both laminae II and III. Observations in the monkey also indicated that laminae III and IV constituted the proper sensory nucleus as described by Rexed.

Although there are numerous descriptions of the lateral cervical nucleus in the cat (Rexed and Brodal, '51; Brodal and Rexed, '53; Morin and Catalano, '55; Ha and Liu, '66), there are few detailed descriptions of this nucleus in the primate (Ha and Morin, '64). Observations in several series of Nissl stained sections revealed a small linear collection of cells lateral to the posterior horn in the upper three cervical segments. These cells, appearing as the primate equivalent of the lateral cervical nucleus of the cat, were mainly medium sized, approximately equal to those in lamina IV. This nucleus in the monkey, compared with that in the cat, contained fewer cells which form a narrower band fairly close to the posterior horn. Cells consistently appeared most numerous at the C1 level, but a few cells usually were found at C2 and C3. No dorsal root fibers were observed to project to the lateral cervical nucleus in the monkey.

A small cell group in the medial part of lamina VII near the central gray, but slightly ventrolateral to the central canal, deserves comment. This cell group, found at all levels of the spinal cord, appears to correspond to the intermediomedial nucleus as described by Rexed. This nucleus received dorsal root afferents at all levels.

The central cervical nucleus (i.e., intermediate nucleus; Cajal, '09) constituted a column of relatively large cells in medial and central portions of lamina VII in the upper four cervical spinal segments. As described by Rexed ('54), the central cervical nucleus lies almost exactly lateral to the central canal. In the monkey this

nucleus was located closer to the central canal than in the cat and frequently extended into the rostral part of the C5 segment. Even though the borders of this nucleus were not sharply delimited, its large, star-shaped cells were distinctive (fig. 1). This nucleus received dorsal root fibers from certain cervical spinal roots and appeared to be directly continuous with the intermediate nucleus of the medulla.

In order to investigate the relationship between the central cervical nucleus and the intermediate nucleus, which has been denied by Rexed ('54), serial horizontal sections of the medulla and upper cervical segments were prepared in three animals. Observations in this material indicated that the intermediate nucleus was a long cell column extending from the caudal medulla rostrally beyond the caudal pole of the dorsal motor nucleus of the vagus. Throughout its extent cells of this nucleus were somewhat ventrolateral to the central canal, or ventral to the fourth ventricle. Rostral portions of the cell column were farther from the midline than caudal parts; the most rostral parts of the nucleus lay dorsolateral to the hypoglossal nucleus and lateral to the dorsal motor nucleus of the vagus nerve (fig. 2).

Cells of the intermediate nucleus of the medulla were heterogeneous in size and shape. In the caudal half of the nucleus large star-shaped cells predominated while rostrally smaller round or spindle-shaped cells formed the bulk of the nucleus (fig. 1). Observations suggested that the central cervical nucleus and the intermediate nucleus constituted parts of the same cell column; this cell column seemed to be partially interrupted in the region of the corticospinal decussation, but some large cells usually could be seen bridging this region.

The concept that the central cervical nucleus and the intermediate nucleus are parts of the same cell column is supported by the fact that: (1) cells of these nuclei share at least one common source of afferent fibers (i.e., fibers from the upper five cervical dorsal roots), and (2) dorsal root fibers terminate in extensive pericellular arborizations in all parts of the cell column.

Cervical dorsal roots

Each of the eight cervical dorsal roots was sectioned individually. Twenty-one cervical dorsal roots were sectioned in 16 different animals.

First cervical root. The first cervical dorsal root was sectioned unilaterally (C-916) and bilaterally (C-1008; fig. 3) and was found to be absent in four other animals (C-974, C-978, C-981 and C-1006) explored surgically. All degenerated fibers contained in the slender C1 root entered the most lateral part of the fasciculus cuneatus at this level and in the caudal medulla (fig. 7). No degenerated root fibers entered the spinal trigeminal tract or nucleus, but some fibers from the fasciculus cuneatus entered the spinal gray in the cervix region of the posterior horn and coursed ventromedially to terminate about cells of the central cervical nucleus. Very sparse degeneration was seen in parts of the anterior horn. A few degenerated fibers descended in the fasciculus cuneatus through the C2 level in rhesus C-1008, but none was seen in rhesus C-916. Moderate degeneration was present in the central cervical nucleus at C2 in both animals. All other cervical segments were free of degenerated fibers.

In transverse sections of the medulla ascending degeneration in the fasciculus cuneatus formed a small bundle between the lateral border of the cuneate nucleus and the medial border of the spinal trigeminal nucleus. Rostrally, ascending degeneration in the most lateral part of the fasciculus cuneatus formed a narrow band immediately dorsal to the spinal trigeminal tract and nucleus. At caudal medullary levels degenerated fibers entered the lateral border of the cuneate nucleus, swept ventromedially along the margin of this nucleus, and entered the intermediate nucleus. Some of these fibers traversed this nucleus in their projection to dorsomedial portions of the supraspinal nucleus (fig. 36). Degeneration in the intermediate nucleus was most profuse caudally, but extended through low olivary levels of the medulla.

Ascending root fibers passing to the accessory cuneate nucleus were more numerous than those terminating in the

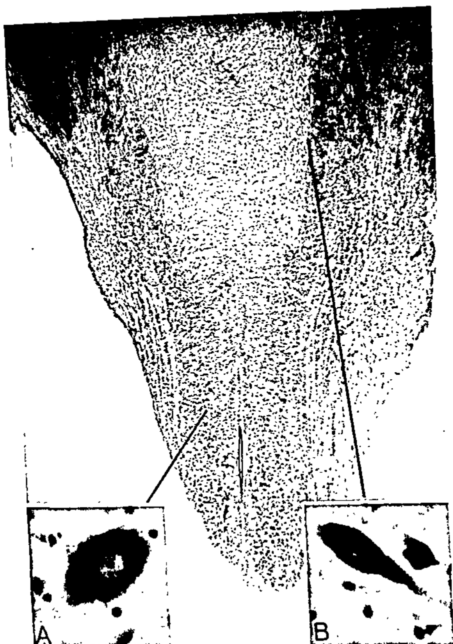


Fig 1 Horizontal brain stem section of rhesus C-972 demonstrating continuity of the intermediate and central cervical nuclei. This section corresponds to the most ventral section of the serial drawings of figure 2 Nissl × 14 Insets A and B are photomicrographs of single cells in different parts of the cell column Nissl × 500

nucleus was located closer to the central canal than in the cat and frequently extended into the rostral part of the C5 segment. Even though the borders of this nucleus were not sharply delimited, its large, star-shaped cells were distinctive (fig. 1). This nucleus received dorsal root fibers from certain cervical spinal roots and appeared to be directly continuous with the intermediate nucleus of the medulla.

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Ascending root fibers passing to the accessory cuneate nucleus were more numerous than those terminating in the

cuneate nucleus. Caudally, degenerated fibers, which entered the dorsolateral border of the cuneate nucleus and swept ventromedially, appeared to be primarily fibers of passage. Degeneration, entering the lateral part of the nucleus in its middle and rostral thirds, terminated in a small region along the ventrolateral border of the nucleus (fig. 5). Additional small islands of degeneration were present in ventromedial and ventrocentral portions of the nucleus. The extreme oral pole of the cuneate nucleus was free of degeneration. Dorsal root fibers entering the medial part of the accessory cuneate nucleus in its middle and rostral thirds were distributed in a discrete wedge-shaped area in the most ventromedial portion of the nucleus (fig. 25). Degeneration in this wedge-shaped area diminished rostrally, but extended to the oral pole of the accessory cuneate nucleus.

Second cervical root. The second cervical dorsal root was sectioned in five monkeys (C-913, C-970, C-981, C-1006 and C-1024). Observations in these animals were similar, except that degeneration in rhesus C-1024 was more profuse, distributed more extensively, and extended farther caudally than in the other animals. At the level of the rhizotomy degenerated fibers entered the lateral part of the fasciculus cuneatus from which site part of the fibers traversed lamina II and terminated primarily in lateral regions of laminae III and IV (fig. 13). A bundle of degenerated fibers traversed central parts of laminae V and VI to arborize profusely about large cells of the central cervical nucleus (figs. 19, 20). In rhesus C-1024 a relatively large field of terminal degeneration was observed in the central portion of lamina VI. A contingent of degeneration passed to the intermediomedial nucleus. Groups of fibers (scattered and small in four monkeys and numerous in C-1024) coursed ventrally, both traversing and terminating, in medial and central parts of laminae VII and VIII. Relatively modest degeneration was seen in the ventromedial nucleus of lamina IX in rhesus C-913 and C-981, while in rhesus C-970 and C-1006 modest degeneration was seen mainly in the spinal accessory cell group. Approximately equal numbers of degenerated fibers were distributed to these cell groups of lamina IX in rhesus C-1024.

Descending C2 root fibers in the lateral part of the fasciculus cuneatus could be found at C3 (C-970 and C-981), and C4 (C-913, C-1006 and C-1024) spinal segments, but not at more caudal levels. Root fibers arborized extensively about cells of the central cervical nucleus at C3 and C4 levels in all animals and at the C5 level in one animal (C-1024). Less intense degeneration was present in laminae III, IV and IX at C3 and C4 segments in the same pattern as at C2. Degeneration extended caudally one segment in the intermediomedial nucleus and two to four segments in medial and central parts of lamina VII.

In the C1 segment the distribution and amount of ascending C2 root fibers in all portions of the spinal gray, except in laminae III and IV, was similar to that observed in the segment of root entry. Degenerated fibers in laminae III and IV were absent (three animals), scant and localized ventrolaterally (C-1006), and numerous and localized laterally (C-1024).

Ascending degeneration passing to medullary relay nuclei was studied in horizontal (C-913 and C-970) and transverse (C-1006 and C-1024) sections. Degeneration in the accessory cuneate nucleus was profuse and well localized throughout its rostrocaudal extent. Degenerated fascicles from the fasciculus cuneatus passed directly into the ventralmost part of the caudal pole of the accessory cuneate nucleus (figs. 26, 31). Degeneration in middle and rostral portions of the nucleus occupied a prominent wedge-shaped area in its ventral third. A narrow area in the ventralmost part of the nucleus was relatively free of degeneration. In the ventral third of the rostral pole of the nucleus, degeneration terminated in a circular area slightly removed from the most ventral and lateral borders of the nucleus.

Degenerated fibers from the fasciculus cuneatus entered the dorsolateral part of the cuneate nucleus and swept ventromedially, forming an oblique band along virtually its entire length (fig. 26). This degeneration was slightly removed from the most ventrolateral portion of the nucleus. Caudally, the majority of these

Abbreviations

ACN, accessory cuneate nucleus
Ce, central cervical nucleus
CST, corticospinal tract
CST dec., corticospinal decussation
FC, fasciculus cuneatus
FG, fasciculus gracilis
ICP, inferior cerebellar peduncle
IO, inferior olivary complex
LCN, lateral cervical nucleus
LRN, lateral reticular nucleus
ML, medial lemniscus
MLF, medial longitudinal fasciculus
NC, nucleus cuneatus
NG, nucleus gracilis
NI, nucleus intermedius
Nuc. V, spinal trigeminal nucleus
PH, posterior horn
PRN, paramedian reticular nucleus
RF, reticular formation
SF, solitary fasciculus
SN, solitary nucleus
Tr. V, spinal trigeminal tract
X, dorsal motor nucleus of the vagus
XII, hypoglossal nucleus

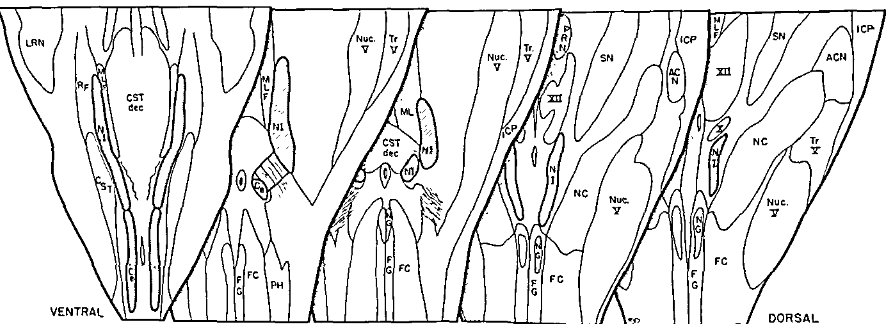


Fig 2 Rhesus C-972. Selected drawings from projected serial horizontal sections demonstrating the continuity and extent of the intermediate and central cervical nuclei. Abbreviations used here and in other figures are listed separately.

terminations were seen in all portions of the nucleus, but were most concentrated dorsomedially. In rhesus C-1024 degeneration in the supraspinal nucleus was more profuse than in other animals.

Two animals (C-1006 and C-1024) exhibited small, but definite, projections to certain other medullary nuclei. A few fibers from the ventrolateral part of the fasciculus cuneatus coursed through medial portions of the spinal trigeminal nucleus to terminate in ventrolateral parts of the magnocellular division of the nucleus. Degenerated fibers also were traced to the nucleus of Roller, the parvocellular reticular formation medial to the cuneate nucleus, the nucleus intercalatus and the peripheral ventral parts of the solitary nucleus. Degeneration seen in ventrolateral parts of the inferior vestibular nucleus and in cell group x in one animal (C-1024) appeared related to ascending degeneration in the posterior spinocerebellar tract.

Third cervical root. Since section of the third cervical dorsal root in two animals (C-912 and C-978) produced essentially the same pattern and amount of degeneration, a single description will be given. At the level of rhizotomy degeneration was similar to that described for the C2 dorsal root. Root fibers entered the most lateral part of the fasciculus cuneatus, traversed lamina II in small fascicles, and were distributed to cells throughout laminae III and IV (fig. 8). Fibers coursing through central parts of laminae V and VI projected profusely to the central cervical nucleus. From this region some fibers passed to the intermediomedial nucleus while other fibers projected into parts of laminae VII and VIII. Degeneration seen among cells of lamina IX was particularly prominent in the spinal accessory cell group (fig. 22), though not limited to this group.

At the C4 level degeneration descending in the lateral part of the fasciculus cuneatus terminated in lateral parts of laminae III and IV, while other fibers projected to the central cervical nucleus, as well as, the spinal accessory cell group of lamina IX. The amount of degeneration in these locations was less than at C3. No degeneration was detectable in the fasciculus cuneatus at the C5 level (C-978), but sparse degeneration was seen in laminae III, IV and VII. Modest degeneration was seen in the intermediomedial nucleus and in what appeared to be the caudal part of the central cervical nucleus. No degeneration was seen in more caudal spinal segments.

At the second cervical segment spinal degeneration was similar to that described at C3. Degeneration in the central cervical nucleus was profuse while fibers passing to lamina IX surrounded primarily spinal accessory neurons. Ascending fibers in the fasciculus cuneatus formed a narrow band dorsomedial to the posterior horn. No sections of C1 were available for these animals.

Sections of the medulla were cut transversely (C-912) and horizontally (C-978). Degenerated fibers traversed and terminated in caudal portions of the cuneate nucleus. Fibers entering the lateral part of the cuneate nucleus in its middle third swept ventromedially in a gentle arc and terminated in a crescent-shaped area along the ventral and medial borders of the nucleus. Terminal degeneration was maximal ventrally. More moderate degeneration in the oral third of the nucleus was seen primarily in medial regions. The area of termination of C3 dorsal root fibers in the cuneate nucleus was slightly dorsal to that receiving C2 dorsal root fibers. Root fibers entered the caudal pole of the accessory cuneate nucleus and terminated in ventromedial portions of its caudal third. In ventral parts of the nucleus a bundle of degenerated fibers, coursing in the lateral half of the nucleus, projected to rostral parts of the nucleus, excluding the oral pole. Only modest degeneration was seen caudally in dorsolateral parts of the nucleus.

Collateral fibers from the fasciculus cuneatus projected ventromedially beyond the cuneate nucleus to the intermediate nucleus of the medulla. Although a considerable number of fibers surrounded elements of this long cell column, degeneration was less profuse and extensive than that associated with the C2 dorsal root. Degenerated fibers in the intermediate nucleus extended rostrally almost to the hypoglossal nucleus and caudally as far as the lower border of the corticospinal decus-

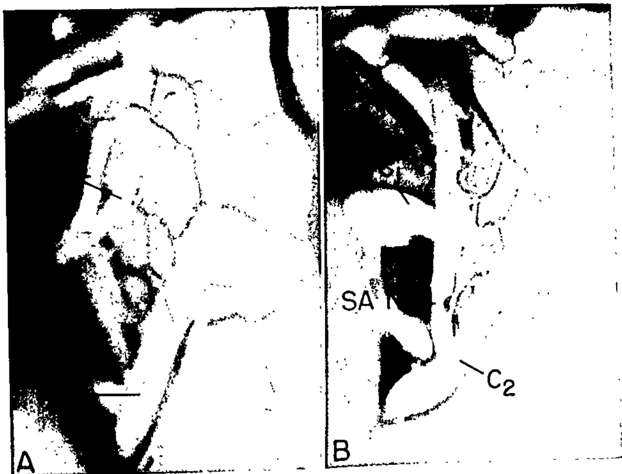


Fig. 3 Rhesus C-1008. Photographs of the first and second cervical dorsal roots taken through the operating microscope. In A, intact rootlets of the left first cervical root are seen. In B, the rootlets of the first cervical dorsal root have been sectioned. The spinal part of the spinal accessory nerve (SAN) can be seen coursing rostrally dorsal to the denticulate ligaments.

fibers appeared to merely traverse the nucleus, though scant terminations were sometimes observed dorsolaterally. In the middle third of the cuneate nucleus, terminal degeneration was chiefly in ventrolateral and ventromedial portions. Rostrally, terminations were localized primarily in the ventromedial part of the nucleus. Collateral fibers, traversing the cuneate nucleus in an oblique ventromedial direction, projected profusely to cells of the intermediate cell column (figs. 33, 34). Impressive pericellular arborizations were seen throughout this cell column (fig. 35). Degeneration about cells of this nucleus extended rostrally beyond the caudal poles of the hypoglossal and dorsal motor vagal nuclei; rostrally terminal degeneration was found about cells of the intermediate nucleus lateral to the dorsal motor nucleus of the vagus. Caudally degenerated fibers were seen about cell groups of the nucleus lying among decussating corticospinal fibers; caudal to the corticospinal decussation virtually identical degeneration arborized about cells of the central cervical nucleus at the C1 level. Observations in these four animals demonstrated that collaterals from the C2 dorsal root project profusely to cells in all parts of the intermediate nucleus of the medulla and to the central cervical nucleus in the upper four cervical spinal segments. The amount and the extent of degeneration seen in these nuclei following C2 dorsal root section was more profuse than that associated with any other cervical dorsal root.

Fascicles of degeneration coursed from ventromedial portions of the cuneate nucleus and ventrolateral portions of the intermediate nucleus to enter the supraspinal nucleus throughout its length. Profuse

upraspinal nucleus. Terminations in the upraspinal nucleus were greatest dorsomedially.

Fifth cervical root. In four animals (C-879, C-914, C-974 and C-1014) the fifth cervical dorsal root was sectioned. Resulting degeneration was similar in these animals, but not identical. At the level of the rhizotomy most of the root fibers coursed medial to the posterior horn to enter the lateral part of the fasciculus cuneatus (figs. 4, 9). From the above location fascicles of root fibers traversed lamina II to arborize about cells in laminae III and IV. Degenerated fibers passed ventromedially through central parts of laminae V and VI and broke up into smaller fascicles in lamina VII. Some fibers appeared to terminate about cells in the central part of lamina VI. A small number of fibers in the medial part of lamina VII arborized about cells of the intermediomedial nucleus (fig. 9). In rhesus C-974 some cells in this part of lamina VII resembled those of the central cervical nucleus. A large number of fibers in central parts of lamina VII projected to ventral and lateral cell groups of lamina IX. Some of these fibers were distributed to the spinal accessory nucleus. A relatively small bundle of fibers entered the most medial part of lamina VIII.

At the C6 level (fig. 4) degeneration was distributed similarly to that at C5, except in laminae III and IV where terminal degeneration was confined to a small ventrolateral region. Descending fibers in the fasciculus cuneatus moved medially away from the posterior horn to a position near the posterior intermediate septum. In diminishing quantities these fibers could be followed caudally in the fasciculus cuneatus to the T4 level (C-879, C-914 and C-1014), but only as far as T1 in rhesus C-974. In lower cervical segments degenerated fibers from the fasciculus cuneatus entered medial parts of lamina VI and central regions of lamina VII. In the upper four thoracic segments modest degeneration passed directly into the dorsal nucleus (fig. 4).

At the C4 level degenerated root fibers were distributed similar to that at C5 with two exceptions: (1) modest degeneration surrounded cells of the central cervical nucleus in all animals, and (2) virtually no degeneration was seen in laminae VIII or IX in two animals (C-879 and C-914). In rostral cervical spinal segments much less degeneration was seen in cell groups of the posterior horn. Some fibers from the fasciculus cuneatus traversed medial parts of laminae V and VI to enter the central cervical nucleus, but the number of degenerated fibers in this nucleus was less than at the C4 level. At C3 and C2 spinal levels a moderate number of degenerated fibers were seen about cells of the spinal accessory nucleus. In all spinal segments rostral to the rhizotomy, fascicles of degenerated fibers streamed ventrally from the fasciculus cuneatus into lateral portions of medial lamina VI.

In the lower medulla ascending fibers from the C5 dorsal root formed a discrete band slightly lateral to the central part of the fasciculus cuneatus (fig. 11). Fibers given off from this bundle entered dorsolateral parts of the cuneate nucleus, swept ventromedially in a curved fashion, and formed a crescent-shaped area just ventral to the central region of the nucleus (figs. 4, 27). Degeneration in this position was seen throughout the cuneate nucleus, except for its oral pole. Fibers tended to be more ventrolateral in rostral areas. Throughout most of the nucleus a small group of fibers remained in the dorsolateral corner of the cuneate nucleus adjacent to the spinal trigeminal nucleus. In regions of the medulla caudal to the hypoglossal nucleus a moderate number of fibers passed ventromedially beyond the cuneate nucleus to enter the intermediate nucleus. Fibers arborizing about cells of this nucleus were most numerous in caudal regions. Some fibers projected beyond the intermediate nucleus to terminate about dorsomedial cells of the supraspinal nucleus.

A moderate number of degenerated fibers entered ventral parts of the accessory cuneate nucleus caudally, but larger numbers of fibers entered medial parts of the nucleus throughout most of its extent. These fibers arborized about cells in a wedge-shaped area just ventral to the central part of the nucleus (figs. 4, 27). In the rostral third of the nucleus a group of fibers from the main wedge-shaped area curved dorsally in a wing-like extension, slightly removed from the lateral border of the nucleus. Although terminal degeneration in this wedge-shaped pattern extended

sation. A small number of collateral fibers passed ventrally through and around cells of the intermediate nucleus to dorsomedial parts of the supraspinal nucleus.

Fourth cervical root. The fourth cervical dorsal root was sectioned in two animals (C-981 and C-1006). Only the spinal degeneration will be described in rhesus C-981 (since the medulla was not available for study) and only the medullary degeneration will be described in rhesus C-1006 (since the spinal cord exhibited some trauma).

Degeneration produced by this root section entered the lateral part of the fasciculus cuneatus and projected in small fascicles through lamina II to arborize about cells throughout laminae III and IV. Root fibers passing ventromedially through central portions of laminae V and VI projected: (1) profusely to cells of the central cervical nucleus, (2) to cells of the intermediomedial nucleus, and (3) to ventral cell groups in lamina IX. Ventromedial cell groups in lamina IX received most of these fibers, although a few fibers passed to cells of the spinal accessory cell group. At C5 descending branches of the C4 root were distributed in the same way as at C4, except that terminal degeneration in laminae III and IV was confined to lateral regions (fig. 14). Root fibers arborized about a discrete collection of large cells in the medial part of lamina VII that resembled the central cervical nucleus. At the C6 level descending degeneration in the fasciculus cuneatus was more medial while degeneration in the gray was present mainly in medial parts of lamina VII. In C7 and C8 spinal segments descending C4 dorsal root fibers were diminished and located medially near the posterior intermediate septum. A few fibers were present about large cells in the medial part of lamina VII at C8 and T1. No descending degeneration was evident in more caudal spinal segments.

In sections of the upper cervical cord ascending degeneration in the fasciculus cuneatus shifted dorsomedially. The distribution of C4 root fibers at C3 was similar to that described at the level of the dorsal rhizotomy, except that a larger number of degenerated fibers were present about cells of the spinal accessory nuclear group. Degeneration surrounding the central cervical nucleus at these levels was impressive. At C2 degeneration was most in laminae IV and VII, moderate in the central cervical nucleus, and scant in the anterior horn.

The medulla of C-1006 was studied in transverse sections. Caudally, degeneration was disposed in a lamina in the dorsal part of the lateral third of the fasciculus cuneatus. A few fibers coursed dorsolaterally for a short distance along the periphery of the fasciculus. Fibers from this bundle entered the lateral part of the cuneate nucleus and swept ventromedially forming a crescent shaped area of degeneration in the dorsal part of the ventral third of the nucleus. Although a large number of these fibers merely traversed the nucleus, a locus of terminal degeneration was seen ventrolaterally. In middle portions of the cuneate nucleus degenerated fibers formed a discrete wedge-shaped area with the base lateral and the tapering apex curving ventromedially. This wedge-shaped area occupied the dorsal part of the ventral third of the nucleus. Rostrally, some fibers from the base of the wedge extended dorsolaterally for a short distance. Degeneration in the rostral pole of the nucleus was diminished compared with mid-cuneate levels.

A moderate amount of degeneration entered ventral portions of the accessory cuneate nucleus caudally, but a larger number of fibers entered medial parts of the nucleus throughout most of its extent. In middle portions of the nucleus these fibers arborized about cells in a wedge-shaped area (base directed laterally) in the dorsal part of its ventral third. Rostrally, fibers from the base of this wedge of degeneration moved medially, leaving a clear area laterally near the border of the accessory cuneate nucleus. This wedge-shaped configuration of degeneration gradually diminished rostrally but extended to the rostral pole of the nucleus.

In caudal parts of the medulla a moderate number of degenerated fibers coursed ventromedially beyond the cuneate nucleus to enter the intermediate nucleus. Terminations, gradually diminishing rostrally, were present in all, except the most rostral portions, of the intermediate nucleus. Some degenerated fibers traversed the intermediate nucleus, and fibers of the pyramidal decussation to terminate in the

ɔraspinal nucleus. Terminations in the ɔraspinal nucleus were greatest dor-nedially.

Fifth cervical root. In four animals (C-9, C-914, C-974 and C-1014) the fifth rvical dorsal root was sectioned. Resulting degeneration was similar in these anials, but not identical. At the level of the izotomy most of the root fibers coursed edial to the posterior horn to enter the ost lateral part of the fasciculus cuneus (figs. 4, 9). From the above location scicles of root fibers traversed lamina II arborize about cells in laminae III and V. Degenerated fibers passed ventromedily through central parts of laminae V and I and broke up into smaller fascicles in mina VII. Some fibers appeared to termiate about cells in the central part of mina VI. A small number of fibers in the edial part of lamina VII arborized about ells of the intermediomedial nucleus (fig.). In rhesus C-974 some cells in this part of amina VII resembled those of the central ervical nucleus. A large number of fibers n central parts of lamina VII projected to entral and lateral cell groups of lamina X. Some of these fibers were distributed to he spinal accessory nucleus. A relatively mall bundle of fibers entered the most nedial part of lamina VIII.

At the C6 level (fig. 4) degeneration was listributed similarly to that at C5, except n laminae III and IV where terminal degeneration was confined to a small ventrolateral region. Descending fibers in the fasciculus cuneatus moved medially away from the posterior horn to a position near the posterior intermediate septum. In diminishing quantities these fibers could be followed caudally in the fasciculus cuneatus to the T4 level (C-879, C-914 and C-1014), but only as far as T1 in rhesus C-974. In lower cervical segments degenerated fibers from the fasciculus cuneatus entered medial parts of lamina VI and central regions of lamina VII. In the upper four thoracic segments modest degeneration passed directly into the dorsal nucleus (fig. 4).

At the C4 level degenerated root fibers were distributed similar to that at C5 with two exceptions: (1) modest degeneration surrounded cells of the central cervical nucleus in all animals, and (2) virtually no degeneration was seen in laminae VIII or IX in two animals (C-879 and C-914). In rostral cervical spinal segments much less degeneration was seen in cell groups of the posterior horn. Some fibers from the fasciculus cuneatus traversed medial parts of laminae V and VI to enter the central cervical nucleus, but the number of degenerated fibers in this nucleus was less than at the C4 level. At C3 and C2 spinal levels a moderate number of degenerated fibers were seen about cells of the spinal accessory nucleus. In all spinal segments rostral to the rhizotomy, fascicles of degenerated fibers streamed ventrally from the fasciculus cuneatus into lateral portions of medial lamina VI.

In the lower medulla ascending fibers from the C5 dorsal root formed a discrete band slightly lateral to the central part of the fasciculus cuneatus (fig. 11). Fibers given off from this bundle entered dorsolateral parts of the cuneate nucleus, swept ventromedially in a curved fashion, and formed a crescent-shaped area just ventral to the central region of the nucleus (figs. 4, 27). Degeneration in this position was seen throughout the cuneate nucleus, except for its oral pole. Fibers tended to be more ventrolateral in rostral areas. Throughout most of the nucleus a small group of fibers remained in the dorsolateral corner of the cuneate nucleus adjacent to the spinal trigeminal nucleus. In regions of the medulla caudal to the hypoglossal nucleus a moderate number of fibers passed ventromedially beyond the cuneate nucleus to enter the intermediate nucleus. Fibers arborizing about cells of this nucleus were most numerous in caudal regions. Some fibers projected beyond the intermediate nucleus to terminate about dorsomedial cells of the supraspinal nucleus.

A moderate number of degenerated fibers entered ventral parts of the accessory cuneate nucleus caudally, but larger numbers of fibers entered medial parts of the nucleus throughout most of its extent. These fibers arborized about cells in a wedge-shaped area just ventral to the central part of the nucleus (figs. 4, 27). In the rostral third of the nucleus a group of fibers from the main wedge-shaped area curved dorsally in a wing-like extension, slightly removed from the lateral border of the nucleus. Although terminal degeneration in this wedge-shaped pattern extended

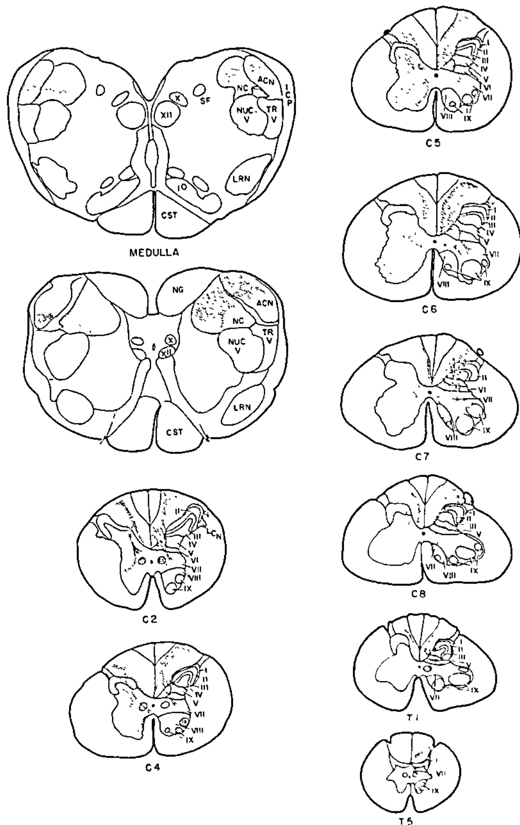


Fig. 4 Rhesus C-1014. Projected drawings showing the distribution of spinal and medullary degeneration resulting from section of dorsal roots C5 (left) and C8 (right). Roman numerals at spinal levels indicate laminae of Rexed.

ughout most of the nucleus, only scant ;neration was seen in its oral pole. somedial regions of the nucleus were of degeneration at all levels.

ixth cervical root. The sixth cervical sal root was sectioned in two animals 967 and C-1012). At the level of the root tion the majority of degenerated fibers rsed medial to the posterior horn and ered the lateral part of the fasciculus ieatus. From this site fibers passed in all fascicles into laminae III and IV. ntrally coursing fibers both traversed d arborized about cells in the medial rts of laminae V and VI. Fibers passing rough these laminae entered lamina VII d projected almost exclusively to ventro- eral cell groups of lamina IX. A small imber of fibers projected to the interme- omedial nucleus and some of these fibers ntinued into the dorsomedial part of mina VIII. Similar degeneration was en in adjacent spinal segments (C7 and 5).

Descending degeneration, migrating into edial parts of the fasciculus cuneatus ear the posterior intermediate septum at 8, continued caudally as far as T3. At C8 me degenerated fibers were seen in iedial parts of laminae V, VI and VII. In ie upper three thoracic segments degen- ration was present in the dorsal nucleus nd medial parts of lamina VII. Only ques- onable degeneration was seen in the dor- al nucleus at T4.

The number and distribution of ascend- ig degenerated fibers in the spinal gray as more extensive in C-1012 than in C- 67. Degeneration in the C4 and C3 seg- nents of C-1012 was distributed similarly o that observed at the level of root section, ut was less profuse. At C3 (C-967) and C2 C-1012) levels a few fibers were scattered n parts of lamina VII and in parts of the entral cervical nucleus. In all spinal seg- nents rostral to the root section, fascicles of degenerated fibers streamed ventrally from the fasciculus cuneatus into lateral portions of medial lamina VI.

In medullary transverse sections degenerated fibers entering the dorsolateral part of the cuneate nucleus swept ventromedially in a crescent configuration (fig. 28). The most ventrolateral part of the nucleus and a large dorsomedial region was free of degeneration. Fibers in the ventral part of the nucleus passed into a ventromedial area along the border of the nucleus. Thus degenerated fibers in the caudal half of the nucleus formed a half circle about a clear dorsomedial region of cuneate neurons. In rostral parts of the cuneate nucleus degenerated fibers appeared to end about cells in an oblique band extending from dorsolateral to ventromedial borders of the nucleus. Near the oral pole of the cuneate nucleus degeneration was seen only in ventromedial regions.

Posterior column fibers entered the accessory cuneate nucleus from its medial border and arborized about cells in a wedge-shaped area slightly ventral to the central part of the nucleus (fig. 28). These fibers occupied a larger area laterally but did not extend to the lateral margin of the nucleus. The position of these fibers was similar to that described for root fibers of C5 except that the wedge-shaped area was slightly more dorsal. In rostral parts of the nucleus the area of maximal degeneration was smaller and more dorsal. These findings suggested a partial overlap in the areas of termination of C5 and C6 dorsal root fibers in the accessory cuneate nucleus.

A few degenerated fibers from the C6 dorsal root projected to dorsomedial regions of the supraspinal nucleus (C-967) and caudal parts of the intermediate nucleus in both animals.

Seventh cervical root. The seventh cervical dorsal root was sectioned in rhesus C-915. At the level of rhizotomy root fibers projected medial to the posterior horn and entered the lateral part of the fasciculus cuneatus. Fascicles of degenerated fibers coursed through medial parts of lamina II to terminate about cells in all parts of laminae III and IV. Small fascicles of fibers from this region passed ventrally, both traversing and arborizing about cells in central portions of laminae V and VI. Fibers entering dorsocentral parts of lamina VII followed two principal projection routes to cells of the anterior horn. A small, distinct bundle of fibers passing ventromedially entered the intermediomedial nucleus and the area immediately ventral to it. Part of the fibers, bypassing these nuclei, projected into medial parts of

lamina VIII and reached the ventromedial cell group of lamina IX. A larger number of fibers in dorsocentral parts of lamina VII streamed ventrally through central parts of this lamina to enter lateral and ventrolateral cell groups of lamina IX. Most of these fibers appeared to end within the dendritic fields of large motor neurons. The pattern of distribution of root fibers in adjacent spinal segments (C8 and C6) was similar but degeneration was less profuse.

Descending degeneration in medial parts of the fasciculus cuneatus was followed caudally as far as T2; degeneration in this location probably continued beyond this level but was not detectable at T5. Most of these fibers terminated about cells in the dorsolateral part of the dorsal nucleus, though some fibers were scattered in adjacent parts of lamina VII.

Ascending degeneration in the fasciculus cuneatus occupied a broad band in the central part of the fasciculus. At the C3 level fibers projected from the fasciculus cuneatus into the lateral part of the medial lamina VI. A tiny amount of degeneration was seen about cells of the central cervical nucleus. Sections of more rostral cervical segments were not available for study.

Sections of the lower brain stem were cut horizontally. Fibers from the fasciculus cuneatus entered the dorsolateral surface of the cuneate nucleus in its caudal half. In successively more ventral sections these fibers occupied more medial areas and extended rostrally in the nucleus. Near central regions of the nucleus a large core area of terminal degeneration was oriented obliquely to the long axis of the nucleus (fig. 32). Degeneration in rostral regions extended nearly to the medial border of the nucleus, while caudally degeneration touched the lateral border of the nucleus. At these levels extreme rostral and caudal poles of the nucleus were free of degeneration, as were the most medial and lateral regions of the nucleus. In the ventral half of the nucleus the central core of terminal degeneration broke up into two linear bands partially connected by fascicles of degenerated fibers. One of these linear bands was along the medial margin of the rostral half of the nucleus and the other, extending the length of the nucleus, was slightly lateral to the central longitudinal axis of the nucleus. In the most ventral regions of the cuneate nucleus termin degeneration was present in both and caudal poles of the nucleus.

Fibers from the fasciculus directly entered the medial and caud parts of the accessory cuneate where the majority of fibers terminated a wedge-shaped central region (fig. Centrally areas of terminal degenerati were found along the medial border of t nucleus. In the most ventral sections small number of fibers from the oral pole the cuneate nucleus passed into n regions of the accessory cuneate nucleu

A very small number of degenerate fibers from the fasciculus cuneatus passe ventromedially beyond the cuneate nu cleus to cells of the intermediate nucleu near the decussation of the corticospin tract. These fibers were localized to th small region and did not arborize exte sively about the individual cells. Rostr portions of the intermediate nucleus the medulla were free of degeneration. N fibers from the C7 dorsal root we observed to enter the supraspinal nucleu

Eighth cervical root. The eighth cerv cal dorsal root was sectioned in two an mals (C-914 and C-1014).

At the level of the rhizotomy the bulk degenerated root fibers arched medial the posterior horn to enter the lateral pa of the fasciculus cuneatus (fig. 4). Fro this location fibers traversed the su stantia gelatinosa to enter laminae I and IV. Ventrally coursing degeneratic traversed central parts of laminae V an VI to enter lamina VII. Numerous sma fascicles of degenerated fibers in lamir VII projected ventrolaterally to lateral ce groups of lamina IX. A distinct group degenerated fibers passed into medi parts of lamina VII where they closely su rounded cells of the intermediomedi nucleus (fig. 17). Part of these medial coursing fibers descended ventrally in tl medial part of lamina VIII to reach tl ventromedial cell group of lamina I Fiber degeneration in the adjacent se ments (fig. 23) was similar to that de scribed for C8 except that degeneration i lamina IX was relatively modest an limited to the lateral cell group at T1.

Descending collaterals of C8 dorsal ro fibers, shifting into more medial parts c the fasciculus cuneatus at the T2 leve

re followed caudally to the T6 level in C-4 and through the T8 level in C-1014. In e upper six (C-914) or nine (C-1014) oracic segments fibers entering the inal gray were distributed to dorsal parts the dorsal nucleus (fig. 4). In the upper-ost thoracic segments degeneration also as present in medial parts of lamina V, edial and central parts of lamina VI, and ljacent regions of lamina VII.

The number of degenerated ascending bers from the C8 dorsal root was greater ian that associated with any other cervi-al dorsal root and these fibers occupied a irger area in the fasciculus cuneatus figs. 4, 11). In the C6 and C5 segments (C-14) degeneration was distributed in a pat-ern similar to that observed at the level of oot entry but fewer fibers were present, specially in lamina IX. At C4 and all more ostral cervical segments, degenerated ibers, entering the spinal gray from the asciculus cuneatus, were limited virtually o medial lamina VI (fig. 21).

In transverse sections of the caudal cuneate nucleus degenerated fibers termi-nated in an oval or circular area in the dorsocentral region of the nucleus immedi-ately ventral to the fasciculus cuneatus. Terminal degeneration in this region of the nucleus persisted throughout the caudal half of the nucleus with only a few fibers passing into the dorsomedial regions. In the middle third of the nucleus the main area of terminal degeneration shifted slightly ventrally, leaving dorsomedial, ventromedial and lateral areas free of terminal degeneration. Rostrally terminal degeneration occupied an oval area in the dorsomedial part of the nucleus (figs. 4, 29), but removed from the borders of the nucleus. Regions of terminal degeneration in the cuneate nucleus associated with the C8 dorsal root were more extensive than those seen with any other dorsal root.

Dorsal root fibers entering the accessory cuneate nucleus projected laterally from the fasciculus cuneatus. These fibers entered dorsomedial parts of the nucleus throughout most of its length and occupied approximately the dorsal third of the nucleus (figs. 4, 29), except rostrally and caudally where degeneration was dimin-ished. Throughout levels of maximal degeneration in the middle third of the accessory cuneate nucleus degeneration was always minimal or absent along the dorsolateral border of the nucleus (fig. 6). Near the oral pole of the nucleus terminal degeneration was found mainly in dorso-central regions. No C8 root fibers projected to the intermediate or supraspinal nuclei.

Thoracic dorsal roots

Each of the upper seven thoracic dorsal roots was sectioned individually in differ-ent monkeys. The first, second and seventh thoracic dorsal roots were sectioned in two different animals.

First thoracic root. In two animals (C-912 and C-1025) the first thoracic dorsal root was sectioned. The distribution of degenerated fibers in these monkeys dif-fered at certain levels, while the amount of degeneration differed at all levels, being much more profuse in rhesus C-1025. Attention will be directed to notable indi-vidual variations in the distribution. At the level of rhizotomy, degenerated fibers from the root entry zone mainly passed into the lateral part of the fasciculus cuneatus, from which location part of the fibers traversed medial portions of the substantia gelatinosa to enter the medial two-thirds of laminae III and IV. Fewer fibers coursed from the zone of Lissauer along the lateral border of the substantia gelatinosa to terminate in a small ventrolateral part of lamina III and to a lesser extent in lamina IV. Degenerated fibers traversed medial and central parts of lamina V to enter cen-tral portions of laminae VI and VII. Modest degeneration was seen about cells of the dorsal nucleus in rhesus C-912, but this nucleus was not apparent in our sections in rhesus C-1025. Degenerated fibers from this locus in central parts of laminae VI and VII followed two trajectories. A smaller number of fibers projected ventromedi-ally to the intermediomedial nucleus. Some fibers traversing this nucleus streamed ventrally along the medial border of lamina VIII to enter the ventromedial cell group of lamina IX. Numerous fibers, forming the other trajectory, both termi-nated in, and coursed ventrally through, medial and central parts of lamina VII to terminate in portions of all lateral cell groups of lamina IX.

In the adjacent rostral segment (C8) the amount and distribution of degeneration was similar to that at TI, except that

degeneration was present throughout laminae III and IV and present in the ventrolateral part of lamina II in both animals. However, fewer fibers terminated in the medial third of laminae III and IV in rhesus C-1025. In the adjacent caudal segment (T2) the amount of degeneration in the spinal gray was decreased, especially in laminae VII and IX. The distribution of fibers was similar to that at T1, with the following exception: degeneration was localized sharply to the medial half of laminae III and IV, although a small island of degeneration persisted in ventrolateral parts of these laminae.

Descending degeneration in the lateral part of the fasciculus cuneatus shifted to more medial parts of the fasciculus at caudal levels; in diminishing numbers these fibers descended to the T9 level. Fibers given off from this bundle projected without localization to portions of the dorsal nucleus as far caudally as T9, and at some levels scattered degeneration also was seen in parts of lamina VII. In one animal (C-1025) diminished and scattered degeneration was present in laminae III, IV and medial and central parts of V through the T4 segment.

Ascending fibers from the T1 dorsal root shifted to medial parts of the fasciculus cuneatus adjacent to the posterior intermediate and posterior median septa. Some of the dorsal fibers extended laterally along the periphery of the fasciculus cuneatus for a short distance. This lamina of ascending T1 dorsal root fibers was extensive though narrower than that formed by fibers of the C8 dorsal root. No sections of the C7 through C5 segments were available for rhesus C-912. In the C7 and C6 segments in rhesus C-1025, degeneration terminating in the spinal gray was considerably decreased but distributed in a pattern similar to that described for the C8 segment. In the C5 segment degenerated fibers projected to: (1) the medial half of laminae III and IV, (2) medial parts of lamina V, (3) medial and central parts of lamina VI, and (4) dorsocentral parts of lamina VII. We were unable to explain this extensive distribution of T1 fibers in laminae III and IV at this level. At C4 and more rostral cervical segments, some fibers were given off from ventromedial parts of the fasciculus cuneatus to the medial subdivision of lamina VI, but these fibers were not as numerous as those associated with the C8 dorsal root.

In transverse medullary sections, degenerated fibers from the medial part of the fasciculus cuneatus entered dorsomedial parts of the cuneate nucleus caudally. At caudal levels terminal degeneration was seen in a crescent-shaped area in the dorsomedial part of the cuneate nucleus partially surrounding a large circular cell mass. Some of the degenerated fibers extended into ventromedial parts of the nucleus, leaving the dorsal and medial borders of the nucleus free of degeneration. In central parts of the nucleus (fig. 5) terminal degeneration was seen in an inverted "U" shaped formation in the dorsomedial part of the nucleus. The open part of the "U" was directed ventrolaterally and a clear zone separated the degeneration from the borders of the nucleus, except dorsolaterally where fibers were entering from the fasciculus cuneatus. A small number of fibers from the medial arm of the "U" projected dorsolaterally to virtually encircle the central cell mass. The latter fibers were especially prominent in rhesus C-1025. The above pattern persisted throughout most of the nucleus but near the oral pole terminal degeneration was concentrated in a smaller dorsomedial region slightly removed from the borders of the nucleus.

Fibers entering the accessory cuneate nucleus arched laterally into dorsomedial parts of the nucleus. While terminal degeneration was greatest dorsomedially, it also was seen about some cells in lateral parts of the nucleus rostrally. The distribution of terminal degeneration was comma-shaped with the tail directed laterally and ventrally (fig. 6). The most dorsal margin of the nucleus was free of degenerated fibers as was most of the ventral two-thirds of the nucleus. In extreme rostral parts of the nucleus reduced amounts of degeneration were seen in dorsocentral regions in C-912 and in dorsal and lateral regions in C-1025. Although these fibers were distributed in a pattern similar to that of the C8 dorsal root they were less numerous and extended further ventrally in lateral parts of the nucleus. No T1 dorsal root fibers projected to other medullary relay nuclei.

Second thoracic root. The second thoracic dorsal root was sectioned in two animals (C-913 and C-1008). In sections of the T2 segment degeneration was present in the lateral part of the fasciculus cuneatus; part of these fibers traversed, or curved around the medial border of, the substantia gelatinosa to enter laminae III and IV (fig. 10). A smaller number of fibers from the lateral part of the zone of Lissauer coursed through lateral parts of the posterior horn to terminate in a small island of cells in the ventrolateral parts of laminae III and IV (fig. 15). Degenerated fibers destined for the ventromedial cell group of lamina IX followed two main routes: some ventrally coursing fibers traversed central parts of laminae V, VI, VII and VIII, while a greater number of fibers passed through the dorsal nucleus and coursed ventrally along the medial border of lamina VIII. In the dorsal nucleus degeneration was profuse, but not all cells were surrounded by root fibers. A modicum of degeneration was present in the intermediomedial nucleus.

Degeneration in sections of the T1 spinal segment was distributed similar to that of the T2 segment, except that degeneration in the dorsal nucleus (C-913; fig. 16) and lamina IX (C-1008) was more profuse. Similarly distributed, but more modest degeneration, at T3 was seen in laminae III and IV (C-1008) and in laminae VII, VIII and IX. The extent of the descending degeneration from the T2 dorsal root could not be accurately determined in rhesus C-913. In rhesus C-1008 descending degeneration from the T2 dorsal root was present in the lateral part of the fasciculus cuneatus through T6. The dorsal nucleus, and all laminae which received degeneration at the level of root section, except lamina IX, exhibited some degeneration at T4 and T5 levels; the amount of degeneration was reduced at these levels. At the T6 level, degeneration was limited to dorsal portions of the dorsal nucleus.

Fibers from the T2 dorsal root ascended in the dorsomedial portion of the fasciculus cuneatus. The extent and distribution of T2 dorsal root degeneration in the spinal gray of the lower cervical cord could not be determined in rhesus C-1008 because of trauma to the lateral funiculus. At C8 in rhesus C-913 scant degeneration was present in lateral portions of laminae III and IV and in medial and central portions of laminae V, VI and VII. No sections of the C7 through C5 spinal segments were available; degeneration at C4 and in more rostral cervical segments was confined to the fasciculus cuneatus.

Sections of the lower brain stem were cut horizontally (C-913) and transversely (C-1008). Observations in these two animals were similar in that relatively modest degeneration was found mainly in dorsomedial parts of the cuneate nucleus. In rhesus C-913 the bulk of this degeneration was in the caudal two-thirds of the nucleus, while in rhesus C-1008 degeneration in this location was found predominantly in the rostral two-thirds of the nucleus. In both animals degenerated fibers extended ventrally along the medial border of the cuneate nucleus, and a few fibers arched laterally along the dorsal border of the nucleus. Degenerated fibers in the accessory cuneate nucleus were distributed along the lateral border of the nucleus (fig. 30). Rostrally degeneration was most abundant dorsally; smaller amounts of degeneration coursed ventrally along the lateral border of the nucleus.

Third thoracic root. After section of the T3 dorsal root in rhesus C-967 the majority of degenerated fibers at this level passed into lateral parts of the fasciculus cuneatus. Degeneration entered the spinal gray by traversing the substantia gelatinosa and by curving along its medial border. Degeneration was seen in laminae III and IV, central portions of lamina V, and in the dorsal and intermediomedial nuclei. Degeneration in the dorsal nucleus was scattered. From the area of the dorsal nucleus degenerated fibers coursed ventrally in medial portions of laminae VII and VIII to the ventromedial cell groups in lamina IX. Many of these medial fibers curved along the border of the anterior horn to supply cells in medial, central and lateral portions of lamina IX. Degeneration in adjacent thoracic segments, T2 and T4, was essentially the same as at the T3 level though less abundant.

Descending degeneration consisted of a small bundle of fibers in lateral parts of the fasciculus cuneatus that could be traced as far as T5; degeneration in the dorsal

degeneration was present throughout laminae III and IV and present in the ventrolateral part of lamina II in both animals. However, fewer fibers terminated in the medial third of laminae III and IV in rhesus C-1025. In the adjacent caudal segment (T2) the amount of degeneration in the spinal gray was decreased, especially in laminae VII and IX. The distribution of fibers was similar to that at T1, with the following exception: degeneration was localized sharply to the medial half of laminae III and IV, although a small island of degeneration persisted in ventrolateral parts of these laminae.

Descending degeneration in the lateral part of the fasciculus cuneatus shifted to more medial parts of the fasciculus at caudal levels; in diminishing numbers these fibers descended to the T9 level. Fibers given off from this bundle projected without localization to portions of the dorsal nucleus as far caudally as T9, and at some levels scattered degeneration also was seen in parts of lamina VII. In one animal (C-1025) diminished and scattered degeneration was present in laminae III, IV and medial and central parts of V through the T4 segment.

Ascending fibers from the T1 dorsal root shifted to medial parts of the fasciculus cuneatus adjacent to the posterior intermediate and posterior median septa. Some of the dorsal fibers extended laterally along the periphery of the fasciculus cuneatus for a short distance. This lamina of ascending T1 dorsal root fibers was extensive though narrower than that formed by fibers of the C8 dorsal root. No sections of the C7 through C5 segments were available for rhesus C-912. In the C7 and C6 segments in rhesus C-1025, degeneration terminating in the spinal gray was considerably decreased but distributed in a pattern similar to that described for the C8 segment. In the C5 segment degenerated fibers projected to: (1) the medial half of laminae III and IV, (2) medial parts of lamina V, (3) medial and central parts of lamina VI, and (4) dorsocentral parts of lamina VII. We were unable to explain this extensive distribution of T1 fibers in laminae III and IV at this level. At C4 and more rostral cervical segments, some fibers were given off from ventromedial parts of the fasciculus cuneatus to the medial subdivision of lamina VI, but these fibers were not as numerous as those associated with the C8 dorsal root.

In transverse medullary sections, degenerated fibers from the medial part of the fasciculus cuneatus entered dorsomedial parts of the cuneate nucleus caudally. At caudal levels terminal degeneration was seen in a crescent-shaped area in the dorsomedial part of the cuneate nucleus, partially surrounding a large circular cell mass. Some of the degenerated fibers extended into ventromedial parts of the nucleus, leaving the dorsal and medial borders of the nucleus free of degeneration. In central parts of the nucleus (fig. 5) terminal degeneration was seen in an inverted "U" shaped formation in the dorsomedial part of the nucleus. The open part of the "U" was directed ventrolaterally and a clear zone separated the degeneration from the borders of the nucleus, except dorsolaterally where fibers were entering from the fasciculus cuneatus. A small number of fibers from the medial arm of the "U" projected dorsolaterally to virtually encircle the central cell mass. The latter fibers were especially prominent in rhesus C-1025. The above pattern persisted throughout most of the nucleus but near the oral pole terminal degeneration was concentrated in a smaller dorsomedial region slightly removed from the borders of the nucleus.

Fibers entering the accessory cuneate nucleus arched laterally into dorsomedial parts of the nucleus. While terminal degeneration was greatest dorsomedially, it also was seen about some cells in lateral parts of the nucleus rostrally. The distribution of terminal degeneration was comma-shaped with the tail directed laterally and ventrally (fig. 6). The most dorsal margin of the nucleus was free of degenerated fibers, as was most of the ventral two-thirds of the nucleus. In extreme rostral parts of the nucleus reduced amounts of degeneration were seen in dorsocentral regions in C-912 and in dorsal and lateral regions in C-1025. Although these fibers were distributed in a pattern similar to that of the C8 dorsal root, they were less numerous and extended further ventrally in lateral parts of the nucleus. No T1 dorsal root fibers projected to other medullary relay nuclei.

Descending degeneration exhibited a notable attenuation between the T6 and T7 segments but was traced through the T11 segment. This degeneration consisted of a narrow, progressively diminishing, lamina of fibers in the lateral part of the fasciculus cuneatus, and a modicum of fibers in the dorsal nucleus. Degeneration in caudal portions of the dorsal nucleus was scant.

Ascending degeneration in the fasciculus cuneatus moved progressively dorsomedially in upper thoracic segments. At T1 and C8 levels, these fibers occupied portions of the fasciculus cuneatus adjacent to the posterior intermediate and posterior median septa. Some of the most dorsal fibers extended laterally along the periphery of the fasciculus cuneatus. In the C7 and all more rostral cervical segments, ascending degeneration in the fasciculus cuneatus formed a discrete, narrow lamina along the entire length of the posterior intermediate septum and extended laterally for a short distance along the dorsal border of the fasciculus. In the T3 segment, scant degeneration was present in laminae III and IV, and medial parts of laminae VII and VIII. The dorsal nucleus exhibited degeneration through the T2 segment.

In transverse sections of the medulla degenerated fibers formed a very narrow, discrete lamina along the entire medial part of the fasciculus cuneatus. Some fibers coursed ventrolaterally along the periphery of the fasciculus. Caudally, a few fibers terminated in medial parts of the cuneate nucleus. At mid-cuneate levels, degeneration extended along the entire medial portion of the nucleus. Degeneration filled the apex of the nucleus dorsally but was slightly removed from the medial border of the nucleus. Ventrally a few fibers appeared to terminate within the dorsolateral border of the nucleus gracilis. In its rostral third, degeneration terminated along the medial border of the cuneate nucleus with the greatest concentration of fibers dorsomedial. No degeneration was apparent in the rostral pole of the cuneate nucleus.

In caudal portions of the accessory cuneate nucleus, degeneration terminated dorsolaterally. Proceeding rostrally, some fibers from this dorsolateral area extended ventrally so that two areas of termination could be distinguished in middle portions of the nucleus: (1) a larger dorsolateral area, and (2) a smaller ventrolateral area. Rostrally, degeneration was limited to a small ventrolateral area in the accessory cuneate nucleus.

Sixth thoracic root. Section of the sixth thoracic dorsal root in rhesus C-1017 produced degeneration in the lateral part of the fasciculus cuneatus adjacent to the posterior horn, and in the zone of Lissauer. Fascicles of degenerated fibers from the fasciculus cuneatus entered the spinal gray by coursing through, or curving around, medial portions of the substantia gelatinosa. Fibers terminated throughout laminae III and IV, but were most concentrated medially. Other fascicles of degeneration traversed medial parts of laminae III and IV and central parts of lamina V to enter the dorsal nucleus. Profuse pericellular arborizations were observed throughout the dorsal nucleus, but were most abundant medially. Some fibers merely traversing the dorsal nucleus or adjacent portions of lamina VII, were distributed in three main directions. Numerous fibers emerged from the ventromedial border of the dorsal nucleus and after traversing and terminating in the intermediomedial nucleus and adjacent parts of lamina VII, coursed ventrally along the medial borders of laminae VII and VIII to enter lamina IX. Some of these fibers following the curvature of the anterior horn were distributed throughout the ventromedial cell group and to portions of the lateromedial cell group. Other fibers issuing from the dorsal nucleus coursed ventrolaterally to terminate along the lateral border of lamina VIII; a few of these fibers reached the lateromedial cell group of lamina IX. Few fibers passed ventrally through central portions of laminae VII and VIII.

In the T7 segment degenerated fibers were markedly decreased in number, but their distribution in the spinal gray was similar to that at T6, except that degeneration in the dorsal nucleus was primarily dorsolateral. Descending degeneration in the lateral part of the posterior funiculus formed a narrow, progressively diminishing, lamina which extended caudally through the T12 segment. These fibers primarily projected to dorsolateral parts of the dorsal nucleus through the L2 level.

nucleus was modest at T5 and minute at T7.

Ascending degeneration shifted to dorsomedial parts of the fasciculus cuneatus near the posterior intermediate septum. Spinal terminations observed rostral to the T2 segment were not reliable due to cord trauma.

In transverse sections of the caudal medulla degenerated fibers entered dorsomedial parts of the cuneate nucleus; at mid-cuneate levels, this dorsomedial degeneration was greater and some fibers coursed toward ventromedial regions of the nucleus. Degeneration in the cuneate nucleus was most abundant in dorsal regions rostrally. A small contingent of degenerated fibers passed along the lateral border of the cuneate nucleus throughout its rostrocaudal extent and terminated in a small ventrolateral area. Fibers from the fasciculus cuneatus entered dorsolateral parts of the accessory cuneate nucleus where they terminated in a crescent-shaped area. No degeneration was observed in other medullary nuclei.

Fourth thoracic root. In rhesus C-970 the fourth thoracic dorsal root was sectioned. At the level of the root section the distribution and amount of degeneration in the fasciculus cuneatus, the posterior horn and the anterior horn (fig. 24) was similar to that described after section of the T3 dorsal root. Degeneration in the T3 segment was like that at T4, but much more modest degeneration was seen in the T5 segment. Caudal to T5 degenerated dorsal root fibers were present in the posterior funiculus and dorsal nucleus through the T9 segment. The amount of degeneration in the dorsal nucleus was small and never distributed throughout the nucleus.

Fibers from the T4 dorsal root ascended in the dorsomedial part of the fasciculus cuneatus near the posterior intermediate septum. At T2 diminished degeneration was present primarily in the dorsal nucleus and adjacent parts of lamina VII. Degeneration at C8 was confined to the fasciculus cuneatus.

In horizontal sections of the medulla ascending fibers entering the dorsomedial aspect of the cuneate nucleus extended nearly the entire length of the nucleus; terminations were most numerous in the caudal two-thirds of the nucleus. More ventral portions of the cuneate nucleus contained small amounts of degeneration in medial and caudal regions. Small patches of degeneration occupied lateral portions of the cuneate nucleus, especially in its middle third.

Dorsal root fibers primarily reached the accessory cuneate nucleus by passing rostral to the cuneate nucleus. Terminal degeneration was localized to a small lateral area of the nucleus. No degeneration was present in other medullary nuclei.

Fifth thoracic root. Section of the fifth thoracic dorsal root in rhesus C-1024 produced degeneration in the root entry zone, the lateral part of the fasciculus cuneatus and lateral portions of the zone of Lissauer. Degenerated fibers from the fasciculus cuneatus entered the spinal gray in large fascicles by coursing through, or curving around, the ventromedial portion of lamina II. A small number of fine fibers passed from the zone of Lissauer through dorsal parts of lamina II into laminae III and IV. Terminations were noted throughout laminae III and IV, but were scant ventrolaterally. Degenerated fibers traversed the central part of lamina V to reach the intermediomedial and dorsal nuclei (fig. 18), where profuse pericellular arborizations were observed. Other fibers passing through the dorsal nucleus were distributed in three trajectories. Numerous fibers, emerging from the ventral surface of the dorsal nucleus, coursed ventrally along the medial borders of laminae VII and VIII and entered lamina IX. Some of these fibers following the curvature of the anterior horn were distributed to all portions of the ventromedial cell group. A scant number of fibers coursed through central parts of laminae VII and VIII to reach the lateromedial cell group of lamina IX. Finally, a few fibers passed from the lateral part of the dorsal nucleus ventrally along the lateral border of the anterior horn.

The distribution and amount of degeneration in the T4 spinal segment was similar to that in the T5 segment, except that fewer fibers were seen in medial parts of laminae VII and VIII, and in lamina IX. Degeneration in the adjacent caudal segment (T6) was similar to that described for the segment of root section, but the number of degenerated fibers was reduced.

number of cell groups within the medial border of the nucleus cuneatus and within the lateral border of the nucleus gracilis. The bulk of degeneration in the posterior column nuclei was seen dorsally in a junctional zone between the caudal two-thirds of the nucleus cuneatus and the middle third of the nucleus gracilis. Dorsally, degenerated fibers projected from the common border between the nuclei gracilis and cuneatus into the lateral parts of the accessory cuneate nucleus.

DISCUSSION

This study provides data for the rhesus nonkey concerning the distribution and ermination of cervical and upper thoracic lorsal root fibers within the spinal cord and certain medullary nuclei. Observations consider the distribution of dorsal root fibers within the spinal gray, ascending and descending dorsal root projections in the posterior funiculus, and projections of dorsal root fibers to the cuneate, the accessory cuneate, the intermediate and the supraspinal nuclei of the medulla. Comparisons of these observations with similar data for the lower thoracic, lumbosacral and coccygeal dorsal roots are made in Part II of this study (Carpenter, Stein and Shriver, '68).

Spinal projections

Fiber projections to the spinal gray. The local distribution of degenerated fibers from all cervical and the upper seven thoracic dorsal roots in the monkey appears similar to that described in the cat following section of selected individual dorsal roots (Sprague and Ha, '64; Sterling and Kuypers, '67). At the level of rhizotomy degenerated fibers occupy the root entry zone and the most lateral part of the posterior funiculus. From this site dorsal root fibers enter the posterior horn by traversing the medial portions of, or by coursing medially around, laminae II through IV. While numerous degenerated fibers terminate in laminae III and IV, relatively few appear to end in lamina II (Anderson, '60; Ralston, '65; Sterling and Kuypers, '67). Other authors (Szentágothai, '64; Sprague and Ha, '64) described dorsal root fibers that project to lamina IV and then recurve dorsolaterally to enter in radial fashion the entire depth of laminae II and III. Many degenerated fibers traverse lamina IV to course ventrally into laminae V and VI, where they terminate, especially in central and medial regions. A paucity of degeneration in lateral portions of laminae V and VI is notable both in the cat and monkey. From central portions of lamina VI, degenerated fibers stream ventrally through portions of laminae VII and VIII to reach cell groups in lamina IX. A smaller number of fibers consistently project ventromedially to terminate in a nuclear group that seems to correspond to the intermediomedial nucleus of Rexed ('52, '54).

The density of degenerated dorsal root fibers in the spinal cord is greatest at the level of the cut root (fig. 4), an observation in accord with virtually all authors. With certain exceptions, degeneration in spinal segments above and below the rhizotomy is distributed similarly in the spinal gray, but in reduced amounts.

The distribution of dorsal root afferents to the spinal gray at the level of the rhizotomy and in adjacent spinal segments can be discussed systematically with respect to the laminae of Rexed. While a moderate number of dorsal root afferents project to lamina I, especially its medial part, projections to other laminae are more impressive.

Laminae II through IV. As mentioned previously recent observations concerning dorsal root projections to lamina II are not consistent. The extent to which this may be due to differences in terminology is difficult to resolve, but based on the criteria of Rexed ('52, '54, '64), very few dorsal root fibers appear to terminate in this lamina (Anderson, '60; Ralston, '65; Sterling and Kuypers, '67). There is almost universal agreement that dorsal root fibers project profusely to laminae III and IV (i.e., nucleus proprius).

Several investigators have reported a localization of dorsal root terminations within different parts of the laminae of the posterior horn. Szentágothai and Kiss ('49) and Szentágothai ('64) reported that collaterals of large caliber dorsal root afferents exhibit a marked somatotopic projection of dorsal cutaneous regions upon the lateral, and of ventral cutaneous regions upon the

Degeneration one and two segments above the root section was slightly decreased and considerably decreased, respectively, but distributed similar to that in the T6 segment. In the dorsal nucleus, progressively diminishing amounts of degeneration were distributed, without precise localization, through the T2 level. The disposition of the narrow, discrete lamina of ascending degenerated fibers in the fasciculus cuneatus was very similar to that described for the T5 dorsal root.

In transverse sections of the caudal medulla, the majority of degenerated fibers formed a narrow lamina along the medial border of the fasciculus cuneatus; a few fibers extended along the lateral border of the fasciculus gracilis. Caudally, scant degeneration was present in the medial border of the cuneate nucleus and in the ventrolateral border of the nucleus gracilis. At mid-cuneate levels, terminal degeneration was maximal in the medial border of the dorsal two-thirds of the cuneate nucleus; some fibers terminated in adjacent lateral portions of the nucleus gracilis. Fewer fibers extended into ventral parts of these nuclei. Rostrally, degeneration in the medial border of the cuneate nucleus was most abundant dorsally. No degenerated fibers were apparent in the rostral pole of the cuneate nucleus. Throughout the extent of the accessory cuneate nucleus, a modicum of degeneration terminated in a small cluster of neurons in its lateral border. This area of degeneration was dorsal in caudal parts of the nucleus, but shifted ventrally farther rostrally.

Seventh thoracic root. The seventh thoracic dorsal root was sectioned in two monkeys (C-915 and C-1025). At the level of root section degenerated fibers were located in the lateral part of the posterior funiculus and portions of the zone of Lissauer (fig. 12). Degenerated fibers entered the spinal gray by traversing medial portions of lamina II and terminated primarily in medial and central portions of laminae III and IV; sparse degeneration was seen ventrolaterally in these laminae. Degenerated fibers traversed the central part of lamina V to enter the dorsal nucleus where degeneration was noted throughout. Some fibers merely traversed the dorsal nucleus and coursed in three main directions. Numerous fibers emerged from the ventromedial border of the dorsal nucleus and, after sending a small projection to the intermediomedial nucleus, coursed ventrally along the medial borders of laminae VII and VIII to enter lamina IX. These fibers followed the curvature of the anterior horn and were distributed to the ventromedial cell group. Fewer fibers issuing from the ventrolateral surface of the dorsal nucleus coursed: (1) ventrally through central portions of laminae VII and VIII to reach the lateromedial cell group of lamina IX, or (2) ventrolaterally to terminate along the lateral border of lamina VIII.

In the T8 segment degeneration was considerably decreased, but fibers were distributed in laminae III, IV and V as at T7. Degeneration in the dorsal nucleus was concentrated in dorsal regions. At this level fibers coursed through central parts of laminae VII and VIII, but few reached cell groups of lamina IX. Degenerated descending fibers in the lateral part of the posterior funiculus passed primarily to the dorsal nucleus at levels as far caudally as L1. These fibers were distributed mainly to ventral and lateral portions of the nucleus.

In the segment above the root section (i.e., T6) degeneration was reduced but distributed similar to that in the T7 segment. In segments T5 to T2 ascending fibers were distributed to the dorsal nucleus without precise localization.

Ascending degeneration in the fasciculus cuneatus moved progressively dorsomedially in upper thoracic segments, forming a narrow lamina adjacent to the posterior intermediate and posterior median septa. Dorsolaterally some fibers extended along the margin of the fasciculus cuneatus for a short distance. In C7 and all more rostral cervical segments, degenerated fibers formed a narrow, discrete lamina along the entire length of the posterior intermediate septum. Some fibers were located in the most medial portion of the fasciculus cuneatus while others were in the most lateral part of the fasciculus gracilis.

In horizontal (C-915) and transverse (C-1025) sections of the medulla ascending T7 dorsal root fibers supplied a modest

Studies of Escolar ('48) in the cat indicated that the C1 dorsal root projects no fibers to the substantia gelatinosa at that level. Fibers from C2 and C3 dorsal roots were reported to enter the substantia gelatinosa via Lissauer's tract in these segments. Torvik ('56) found that the C2 and C3 dorsal roots projected fibers into the substantia gelatinosa and the nucleus proprius at these levels, but none entered the posterior horn at C1. On the other hand, descending trigeminal fibers projected into the posterior horn at C1 while a few fibers, localized to dorsomedial parts of the posterior horn, were found at C2 levels. More recent studies in the cat (Kerr, '61) indicated that trigeminal fibers form a major afferent component of the posterior horn at C1 and fibers from the first three cervical dorsal roots also end there. Trigeminal fibers were dorsomedial while dorsal root fibers were found ventrolateral.

Although the C1 dorsal root is frequently absent in the monkey (Sherrington, 1898; Howell and Straus, '33; Corbin, Lhamon and Petit, '37; Thomas and Combs, '65), no fibers from this root were seen to enter the head of the posterior horn in the present study. Fibers from the C2 dorsal root terminated primarily in lateral portions of laminae III and IV, while C3 fibers terminated in all parts of these laminae at the C2 level. In two animals a small number of C2 root fibers were seen in ventrolateral (C-1006) or lateral (C-1024) parts of laminae III and IV at the C1 level. In horizontal sections of the C1 segment in two other animals with C2 dorsal rhizotomies only scant degeneration was seen in the head of the posterior horn. These observations suggest that very few dorsal root fibers from the upper cervical segments enter the posterior horn at C1.

Lamina V and VI. The pattern of dorsal root degeneration in the neck (lamina V) and base (lamina VI) of the posterior horn as observed in the cat (Sprague and Ha, '64; Sterling and Kuypers, '67) is strikingly similar to that noted in the monkey. Fascicles of degenerated dorsal root fibers course ventrally through lamina IV into laminae V and VI where they form a large terminal field, primarily in central and medial regions. In all segments, lateral parts of laminae V and VI are nearly free of terminal degeneration.

According to Sterling and Kuypers ('67), dorsal root fibers terminate profusely in laminae V and VI, but, unlike those in laminae III and IV, they do not terminate somatotopically. Observations in the present study indicate that fibers from C5 to T1 dorsal roots terminate in central and medial portions of lamina VI throughout the entire cervical spinal cord. Fibers from the more caudal of these roots project predominantly to more medial areas, while those from the more rostral roots project to more lateral areas. Fibers from C8 dorsal root are the most numerous and fill nearly the entire medial part of lamina VI at all cervical levels (figs. 4, 21). The cells in the medial part of lamina VI at high cervical levels seem to merge almost imperceptibly with cells in the caudal part of the cuneate nucleus. This finding, not noted by previous authors, suggests a unique relationship between dorsal root fibers of the brachial enlargement and cells of the medial part of lamina VI.

Laminae VII and VIII. Degenerated upper cervical dorsal root fibers course ventrally through medial and central portions of laminae VII and VIII to reach motor neurons of lamina IX. The number of terminal ramifications in laminae VII and VIII appeared to be considerably less than in the posterior horn. The pattern of distribution of dorsal root degeneration in laminae VII and VIII in the enlargment confirms the findings of Sprague and Ha ('64) and Sterling and Kuypers ('67). The C5 to T1 dorsal root projections to lamina VIII in segments through the brachial enlargement, where this lamina is restricted to the medial portion of the anterior horn, are chiefly to its dorsal part. Few fibers coursing ventrally in the medial part of lamina VIII reach the medial cell groups of lamina IX. At these levels a larger number of fibers coursing ventrally through medial and central portions of lamina VII reach the more laterally situated cell groups of lamina IX. The majority of degenerated T2 to T7 dorsal root fibers coursed ventrally along the medial borders of laminae VII and VIII to enter lamina IX (fig. 24). Few of these fibers coursed ventrally in central portions or along the lateral borders of these laminae to enter lamina IX.

Four nuclei can be distinguished within lamina VII at certain spinal levels: (1) the

medial halves of laminae II and III. After single rhizotomies of selected lumbosacral dorsal roots, Sprague and Ha ('64) noted the "synaptic compartments of arborization" described by the above authors, as well as a similar compartmentalization of the plexus in lamina IV, not previously described. Comparisons of the distribution of degenerated root fibers at the level of section with segments above and below revealed different parts of laminae II, III and IV to be free of degeneration. In spite of some variation in the terminal fields associated with different lumbosacral roots, in all cases ascending and descending dorsal root fibers showed a marked tendency to send collaterals into medial, or more frequently, lateral parts of laminae II, III and IV of segments adjacent to the cut root. Sterling and Kuypers ('67) reported a somatotopically organized distribution of C5 to T1 dorsal root fibers in laminae III and IV of the brachial enlargement in the cat. They found that caudal roots, supplying primarily distal and postaxial portions of the limb (Hekmatpanah, '61), terminated medially, and more rostral roots, supplying primarily proximal and preaxial portions of the limb, terminated laterally. These observations regarding a somatotopic organization within laminae of the posterior horn are supported by physiological data. Micro-electrode studies of the lumbosacral enlargement in the cat (Wall, '60) suggest that medial portions of the laminae are related to peripheral parts of the leg and lateral portions are related to more proximal parts. Portions of the posterior horn ventral to the substantia gelatinosa could be divided into a dorsal lamina containing cells responding to light touch and a ventral lamina of cells monopolized by proprioceptive afferents.

According to certain authors (Szentágothai and Kiss, '49; Szentágothai, '64; Sprague and Ha, '64), arborizations of collaterals of dorsal root fibers divide laminae II, III and IV into a series of radially oriented "synaptic compartments" or "lobuli." On the basis of studies of brachial spinal cord segments in all three planes with a variety of staining methods, Sterling and Kuypers ('67) have concluded that: (1) the dendritic orientation in laminae I-III is predominantly longitudinal, while that in lamina IV is radial, (2) some dorsal root fibers from Lissauer's tract pass radially into laminae I-III, or recurve from below into laminae III-IV, and (3) the majority of dorsal root afferents to laminae I, III and IV course longitudinally in these laminae after separating from the bundles penetrating the posterior horn. These authors suggested that the posterior horn of the brachial spinal cord consists of longitudinal "slabs" of cells extending throughout this portion of the cord in which progressively more medial portions of the laminae are related to progressively more distal and postaxial portions of the limb.

In the present study dorsal root fibers from C2 to C4 terminated in parts of laminae III and IV one segment above and below the rhizotomy. The dispersion of dorsal root fibers to parts of these laminae was greater for lower cervical dorsal roots (i.e., C5 to C8) in that it extended two or three segments above and below the rhizotomy. Dorsal root fibers from C2 were consistently localized in their terminations to the lateral parts of laminae III and IV at the root entry level and in the adjacent rostral and caudal segments (fig. 13). In other upper cervical segments (i.e., C3, C4 and C5) dorsal root fibers in the segment below their entrance occupied lateral portions of these laminae (fig. 14), but in immediately rostral segments fibers usually were distributed throughout. In all remaining cervical and thoracic segments, dorsal root fibers at their level of entrance and in adjacent segments were distributed to all parts of laminae III and IV, though thoracic dorsal root degeneration frequently was most abundant medially.

The posterior horn in the upper cervical region (i.e., C1 to C3) merits special consideration, because portions of it constitute the nucleus caudalis of the spinal trigeminal nucleus. According to Olszewski ('50) and Torvik '('56), the nucleus caudalis has the same fundamental cellular arrangement as the head of the posterior horn. Following section of the upper three cervical dorsal roots in the cat (Ranson, Davenport and Doles, '32), degenerated fibers were scattered in the posterior horn at C2 and C3, but little degeneration was seen at C1. Very few fibers entered the substantia gelatinosa in any of these spinal segments.

e VII and VIII. Based upon these ithors' diagramatic representation of this ier distribution, this area of termination ipears to correspond to the intermedi- nedial nucleus and the area immediately entral to it – the same areas which receive orsal root afferents in the monkey. Fibers rojecting to the intermediomedial nucleus ere seen at the level of the rhizotomy and adjacent spinal segment (figs. 4, 17).

Lamina IX. In the present study all dorsal roots, except the first cervical, projected bers to motor neurons of the anterior horn t the level of root entry and in adjacent pinal segments. The projection from C1 is nconsistent, very sparse, and only at the evel of entrance. These findings are similar to those of Yee and Corbin ('39), Liu '56), Kerr ('61) and Dunn and Matzke ('67) with respect to the C1 dorsal root. In all animals the amount of degeneration in lamina IX was greatest in the segment of dorsal rhizotomy. Previous observations (Sprague and Ha, '64; Sterling and Kuypers, '67) indicating that the number of degenerated fibers in cell groups of lamina IX diminish sharply immediately rostral and caudal to the cut root are corroborated. The finding of Sterling and Kuypers ('67) that the C7 and C8 dorsal roots seem to be distributed more profusely to motor neurons in segments below these roots than to those above was not observed in the monkey.

Dorsal roots of C2 through C5 projected fibers to the spinal accessory nucleus in the upper cervical cord. These findings are similar to those of Escolar ('48) and Liu ('56) indicating that C2 and C3 dorsal roots project to the spinal accessory nucleus, but differ from those of Corbin, Lhamon and Petit ('37) who found no projection from C2 to the spinal accessory nucleus. In the monkey the C3 dorsal root contributed the greatest number of fibers to the spinal accessory nucleus (fig. 22). Quantitative estimates of individual dorsal root projections to this nucleus were not reported to the other investigators. A dorsal root projection from C1 to the spinal accessory nucleus, as reported by Escolar ('48) and Liu ('56) was not consistently observed in the monkey.

Projections in the posterior funiculus. The cross-sectional configuration of the posterior columns varies at different levels of the spinal cord and the precise limits of the fasciculi gracilis and cuneatus are difficult to determine especially in the mid-thoracic region. In the strict sense the fasciculus cuneatus should be defined as consisting of posterior column fibers terminating in the nucleus cuneatus, but it should be noted that all dorsal roots projecting fibers to the cuneate nucleus also project to the accessory cuneate nucleus. The principal spinal projections to these nuclei come from the cervical and upper seven thoracic dorsal roots.

Dorsal root fibers enter the lateral part of the fasciculus cuneatus where they divide into long ascending and short descending branches. At successively higher levels ascending fibers shift to more medial parts of the fasciculus cuneatus as in classical descriptions (figs. 4, 11). Ascending dorsal root fibers from the upper thoracic segments form relatively narrow bands of degeneration in medial portions of the fasciculus cuneatus. The dorsal part of this band of degeneration extends laterally at the surface of the spinal cord for approximately half the width of the fasciculus cuneatus. Ascending fibers from the T2 through T7 dorsal roots form discrete, very narrow laminae along the entire length of the posterior intermediate septum. The T7 fibers are located in the most medial portion of the fasciculus cuneatus and in the most lateral part of the fasciculus gracilis, while the majority of T2 through T6 fibers are in medial portions of the fasciculus cuneatus, especially dorsomedially.

Ascending fibers from brachial dorsal roots (i.e., C5 through T1) occupy the largest portions of the fasciculus cuneatus. These dorsal roots contribute the greatest number of ascending fibers but also demonstrate considerable overlap. The T1 dorsal root fibers occupy a large medial part of the fasciculus cuneatus, overlapping the area containing fibers from all more caudal thoracic dorsal roots. The number of ascending fibers from the C8 dorsal root is greater than that of any other cervical root. Ascending degeneration in the fasciculus cuneatus from the upper four cervical dorsal roots is relatively small, occupies lateral parts of the fasciculus and exhibits overlapping features de-

central cervical nucleus at C1 through C4, (2) the dorsal nucleus of Clarke and the intermediolateral nucleus at T1 through L3, and (3) the intermediomedial nucleus at all spinal levels. All of these nuclei, except the intermediolateral nucleus, receive afferents from specific dorsal roots. Afferent fibers to the central cervical nucleus are derived chiefly from the upper five cervical dorsal roots. Because of the relationship of this nucleus to the intermediate nucleus of the medulla, it will be discussed with the medullary relay nuclei.

Pass ('33), Liu ('56) and Grant and Rexed ('58) have published detailed accounts of degeneration in the dorsal nucleus following dorsal rhizotomies in the cat. Present findings corroborate those of Liu ('56) indicating afferent projections to the dorsal nucleus from all, except the upper four cervical dorsal roots, and differ from those of Pass ('33) and Grant and Rexed ('58) which indicated that the cervical and first thoracic dorsal roots project no fibers to the dorsal nucleus. In accordance with earlier observations in the cat (Pass, '33; Schimert, '39; Liu, '56; Grant and Rexed, '58) and dog (Szentágothai and Albert, '55), the present experiments indicate that dorsal root collaterals project only to the ipsilateral dorsal nucleus. In the cat (Liu, '56; Grant and Rexed, '58) and monkey, dorsal root collaterals from the posterior funiculus penetrate the medial aspect of the posterior horn and enter the dorsal nucleus directly, without passing through other parts of the posterior horn.

At the level of root section degeneration in the dorsal nucleus is profuse, but not all cells are surrounded by root fibers. In segments rostral and caudal to the cut root, degeneration in the dorsal nucleus is diminished and distributed to only part of the nucleus. Although the dorsal root collaterals exhibit an asymmetrical and differential distribution within the dorsal nucleus, no unequivocal and consistent pattern of localized terminations for different dorsal roots is apparent. These findings differ from those of Liu ('56) indicating that C5 to T5 dorsal root degeneration is distributed throughout the dorsal nucleus, and from those of Grant and Rexed ('58) who indicated that ascending degenerated fibers from T2 to T8 dorsal roots project throughout the cross-section of the nucleus in the more rostral segments. In general, descending collaterals of dorsal roots are most often localized to lateral parts of the dorsal nucleus.

The C5 through T7 dorsal roots have an extensive projection to the dorsal nucleus in the monkey, with a single dorsal root distributing fibers in four to ten segments. The projections of the lower four cervical dorsal roots to the dorsal nucleus in the monkey differ from those of the upper seven thoracic dorsal roots in that there are: (1) considerably fewer fibers, and (2) pericellular arborizations are infrequent or absent. Previous investigators who have reported relatively extensive projections from individual dorsal roots to the dorsal nucleus in the cat include Liu ('56) who indicated a two to nine segment distribution of fibers from C5 to T5 dorsal roots, and Grant and Rexed ('58) who indicated a two to six segment distribution of fibers from T2 to T7 dorsal roots. Considerable overlap in the central distribution of different dorsal root fibers to the dorsal nucleus was observed in the monkey, in agreement with findings in the cat (Liu, '56; Grant and Rexed, '58).

After all dorsal rhizotomies in this series, a contingent of degeneration terminates in a small group of cells in the medial part of lamina VII ventrolateral to the central canal (figs. 4, 9, 17). This nucleus appears to correspond with the intermediomedial nucleus of Rexed ('52, '54). Some fibers coursed through, or adjacent to, this nucleus to terminate in areas immediately ventral to it. Fibers projecting to the intermediomedial nucleus course ventromedially from an area of concentrated degeneration which is the central cervical nucleus in the upper four cervical segments, the central portion of lamina VI in the brachial enlargement and the dorsal nucleus in the thoracic segments. In accordance with these observations Schimert ('39) and Sprague and Ha ('64) traced collaterals of dorsal root fibers to the intermediomedial nucleus in the cat. After C5 to T1 dorsal rhizotomies in the cat, Sterling and Kuypers ('67) reported irregular "bursts" of terminal degeneration distributed to a group of cells occupying the medial parts of the junction between lami-

ne VII and VIII. Based upon these authors' diagramatic representation of this fiber distribution, this area of termination appears to correspond to the *intermediomedial* nucleus and the area immediately ventral to it – the same areas which receive dorsal root afferents in the monkey. Fibers projecting to the intermediomedial nucleus were seen at the level of the *rhizotomy and* an adjacent spinal segment (figs. 4, 17).

Lamina IX. In the present study all dorsal roots, except the first cervical, projected fibers to motor neurons of the anterior horn at the level of root entry and *in adjacent* spinal segments. The projection from C1 is inconsistent, very sparse, and only at the level of entrance. These findings are similar to those of Yee and Corbin ('39), Liu ('56), Kerr ('61) and *Dunn and* Matzke ('67) with respect to the C1 dorsal root. In all animals the amount of degeneration in lamina IX was greatest in the segment of dorsal rhizotomy. Previous observations (*Sprague and Ha*, '64; *Sterling and Kuypers*, '67) indicating that the number of degenerated fibers in cell groups of lamina IX diminish sharply immediately rostral and caudal to the cut root are corroborated. The finding of Sterling and Kuypers ('67) that the C7 and C8 dorsal roots seem to be distributed more profusely to motor neurons in segments below these roots than to those above was not observed in the monkey.

Dorsal roots of C2 through C5 projected fibers to the spinal accessory nucleus in the upper cervical cord. These findings are similar to those of Escolar ('48) and Liu ('56) indicating that C2 and C3 dorsal roots project to the spinal accessory nucleus, but differ from those of Corbin, Lhamon and Petit ('37) *who found no projection* from C2 to the spinal accessory nucleus. In the monkey the C3 dorsal *root contributed the* greatest number of fibers to the spinal accessory nucleus (fig. 22). Quantitative estimates of individual *dorsal root projections* to this nucleus were not reported to the other investigators. A dorsal root projection from C1 to the spinal accessory nucleus, as reported by Escolar ('48) and Liu ('56) was not consistently observed in the monkey.

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Following a multiple section of the upper three cervical dorsal roots in the rat, Torvik ('56) observed a small, sharply circumscribed area of degeneration in the lateral part of the cuneate nucleus. Escolar ('48) individually sectioned the upper three cervical dorsal roots in the cat and reported a somatotopic arrangement of degeneration in the cuneate nucleus, in which C1 fibers were located most superiorly and laterally while C2 and C3 fibers were located laterally at all levels. Fibers from C3 were found dorsal to those of C2.

The most detailed analysis of dorsal root projections to the cuneate nucleus in the cat using the Nauta technic was made by Kuypers and Tuerk ('64). Dorsal rhizotomies included C2 and C3 (multiple section), C7 and C8 (multiple section) and T12. While the number of dorsal roots sectioned was small and no overall pattern of termination in the cuneate nucleus was established, certain data seem significant. These authors found degeneration from C2 and C3 dorsal roots in caudal parts of the nucleus cuneatus was dense ventrolaterally and diffuse in the base of the nucleus; in rostral parts of the nucleus degeneration was diffuse. Ascending degeneration from C7 and C8 dorsal roots was dense in dorsomedial parts of the nucleus and diffuse in basal regions caudally; rostrally degeneration from these roots seemed widespread. These authors indicated overlapping projections from these different dorsal roots in rostral parts of the cuneate nucleus. Ascending fibers from T12 dorsal root were distributed to cells in a transitional area between the nuclei gracilis and cuneatus; additional degeneration projected diffusely to basal parts of the cuneate nucleus rostrally. While limited, these observations suggested a dual pattern of fiber termination in the nucleus cuneatus not seen in our material.

On the basis of single dorsal rhizotomies and cordotomies at various levels Walker and Weaver ('42) reported a topical laminar pattern of fiber termination in the mediolateral and rostrocaudal dimensions of the cuneate nucleus of the monkey. Ascending fibers from T1 through T7 projected to medial and caudal regions of the cuneate nucleus; all cervical dorsal roots projected fibers to lateral and rostral parts of the nucleus. These authors noted that fibers passing to the cuneate nucleus seemed to be derived from basal parts of the fasciculus cuneatus while fibers entering the accessory cuneate nucleus were located more dorsally in the fasciculus.

Data concerning the termination of fibers from the upper three cervical dorsal roots in the cuneate nucleus heretofore have been fragmentary. Fibers from these cervical dorsal roots terminate in the most ventrolateral part of the nucleus at different rostrocaudal levels. Fibers of the C1 dorsal root are located most ventrolaterally in the rostral two-thirds of the nucleus. Fibers of the C2 dorsal root occupy the ventrolateral border of the caudal third of the nucleus, but at more rostral levels form a band medial to the C1 fibers. In the caudal pole of the cuneate nucleus fibers of the C3 dorsal root occupy the ventrolateral border of the nucleus, but at more rostral levels form a band medial to the fibers of C2. This pattern of somatotopic termination of upper cervical dorsal root fibers in the nucleus cuneatus appears more specific than in any previous study.

Ascending dorsal root fibers from C4 through C7 are distributed in crescent-shaped bands in progressively more dorsomedial parts of the cuneate nucleus; the hilus of these serial crescents is directed dorsomedially. Fibers of the C8 dorsal root terminate in a large circular or oval cell mass in the dorsal and central parts of the nucleus. Root fibers from T1 terminate in a large inverted "U" shaped area with the open part directed ventrolaterally. The T1 terminal zone caps the region receiving fibers from C8. Upper thoracic dorsal root fibers terminate in narrow, overlapping crescent-shaped bands oriented along the dorsomedial border of the cuneate nucleus. Fibers from all of these dorsal roots terminate throughout the length of the nucleus (fig. 5).

In the monkey the number of ascending dorsal root fibers projecting to the cuneate nucleus and the areas of their laminar

scribed for other root fibers. The smallest contribution of ascending fibers to the fasciculus cuneatus is from C1. These fibers form a narrow band in the lateral part of the fasciculus immediately dorsal to the nucleus caudalis of of the spinal trigeminal complex.

According to classical descriptions descending branches of cervical and thoracic dorsal roots form the small compact fasciculus interfascicularis, or comma tract of Schultze, situated in or near the posterior intermediate septum (Flechsig, 1876; Schultze, 1883). In the monkey descending fibers from cervical and upper thoracic dorsal roots form bands in the posterior funiculus, lateral and parallel to, but at varying distances from the posterior intermediate septum, rather than forming a compact bundle in the septum's middle portion. Descending fibers tend to mimic the position of the ascending fibers, except they do not migrate as far dorsomedially, and there is a greater concentration of fibers in ventromedial parts of the bundle close to the spinal gray. At successive caudal levels there is a gradual reduction in the number of fibers and those in ventromedial locations are the last to disappear. The number and length of the descending fibers in the posterior funiculus varies according to the dorsal root. Fibers from the upper three cervical roots descend from one to two segments, those from C4 through T1 dorsal roots descend from three to eight segments, and those from T2 through T7 dorsal roots extend caudally from two to six segments. Pass ('33) traced degeneration in the posterior funiculus from C6 and C7 dorsal roots caudally for about four segments; similar fibers could be followed caudally for five segments in the present study.

The present findings are in accord with those of Papez ('29), Ranson, Davenport and Doles ('32) and Corbin and Hinsey ('35) indicating that fibers from the upper three cervical dorsal roots do not descend more than two segments, but differ from those of van Gehuchten ('01), Corbin, Lhamon and Petit ('37), Yee and Corbin ('39) and Liu ('56) who reported that fibers of C1 and C2 dorsal roots extend caudally for three segments, and those of C3 for three or four segments. Observations demonstrating that fibers from C4 and (dorsal roots extend caudally for four or fi segments (Yee and Corbin, '39; Liu, '5 were confirmed in the monkey.

The amount, extent, and location of d scending fibers from a single dorsal r showed regional variations in the mo key's spinal gray. Earlier investigators d not indicate the extent of the distributi of these fibers in the spinal gray. Aft sectioning dorsal roots of the brachi plexus, Sterling and Kuypers ('67) report many degenerated fibers from a single do sal root distributed to laminae III-VI throughout the brachial cord. In the mo key, some fibers from C5 through T1 dors roots supplied certain of these lamina throughout the brachial enlargement, b not all of these roots supplied all of thes laminae throughout this region.

Medullary projections

Nucleus cuneatus. The nucleus cune atus exhibits regional differences in it cytoarchitecture, position and shape (Fer raro and Barrera, '35a; Meesen an Olszewski, '49; Olszewski and Baxter, '54 In Nissl and Golgi stained sections Kuy pers and Tuerk ('64) reported rostrocauda and dorsoventral variations in the cellula and dendritic architecture of this nucleu in the cat. Dorsal areas of the cuneat nucleus contained clusters of round cel with many bushy dendrites, while bas areas contained triangular, multipolar an fusiform cells with long, sparsely radiatin dendrites. These authors indicated that th mode of distribution of dorsal root fibers i the cuneate nucleus is related to certai features of the nuclear organizatio Round cell clusters with multiple bush dendrites, considered to receive afferen principally from distal parts of the extrem ties, were thought to be related to sma cutaneous receptive fields. Triangular an multipolar cells with sparsely radiatin dendrites, considered to receive afferen primarily from proximal parts of the ex tremities and trunk, were regarded as bein related to larger cutaneous fields.

All dorsal roots studied in this series (C through T7) projected fibers to specif parts of the cuneate nucleus (fig. 5). I addition, dorsal roots T8 through L1 (Ca penter, Stein and Shriver, '68), project

ıall number of fibers to this nucleus. ırsal root projections to the cuneate ıcleus have been reported from levels as ıv as T8 in the rhesus monkey (Walker ıd Weaver, '42) and from levels as caudal T12 (Kuypers and Tuerk, '64) and S1 land, '66) in the cat.

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Data concerning the termination of fibers from the upper three cervical dorsal roots in the cuneate nucleus heretofore have been fragmentary. Fibers from these cervical dorsal roots terminate in the most ventrolateral part of the nucleus at different rostrocaudal levels. Fibers of the C1 dorsal root are located most ventrolaterally in the rostral two-thirds of the nucleus. Fibers of the C2 dorsal root occupy the ventrolateral border of the caudal third of the nucleus, but at more rostral levels form a band medial to the C1 fibers. In the caudal pole of the cuneate nucleus fibers of the C3 dorsal root occupy the ventrolateral border of the nucleus, but at more rostral levels form a band medial to the fibers of C2. This pattern of somatotopic termination of upper cervical dorsal root fibers in the nucleus cuneatus appears more specific than in any previous study.

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In the monkey the number of ascending dorsal root fibers projecting to the cuneate nucleus and the areas of their laminar

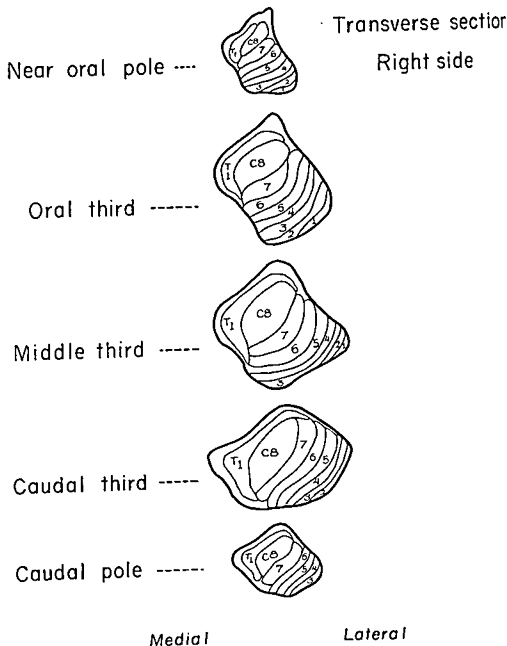


Fig. 5 Schematic diagram of the terminal zones in the cuneate nucleus receiving fibers from dorsal roots C1 through T1. The zone for each root shows the principal area of fiber termination but not the extent of overlap. Fibers from dorsal roots T2 through T7 terminate in overlapping fashion mainly along the dorsomedial border of the nucleus.

termination are roughly related to the size of the dorsal root. The largest areas of terminal degeneration in the cuneate nucleus are those related to dorsal roots C5 through T1. In absolute size the areas of termination of C7, C8 and T1 dorsal root fibers are the largest. While C1 through T1 dorsal root fibers projecting to the cuneate nucleus terminate in exclusive and overlapping zones, the degree of overlap of adjacent dorsal roots appears less than the accessory cuneate nucleus. Overlap dorsal root fibers in the cuneate nucleus primarily within the territory of the ne highest root. The present study does r provide detailed information concerni the manner in which fibers from through T7 dorsal roots terminate in t cuneate nucleus. Fibers from t thoracic dorsal roots are modest in

..d appear to have almost entirely overlap- ..ig terminations in dorsomedial parts of ..: nucleus.

..ccessory cuneate nucleus. The acces- ..y cuneate nucleus is considered to be ..: medullary equivalent of the dorsal ..cleus of Clarke (*Hollis, 1884; Blume- ..u,* 1891; Sherrington, 1893; Pass, '33; ..odal, '41). These nuclei share the follow- ..g anatomical and functional characteris- ..s: (1) cells are morphologically similar ..th eccentric nuclei, (2) afferent fibers ..e derived from dorsal roots, (3) both ..iclei give rise to uncrossed cerebellar ..ferent fibers, and (4) both nuclei relay ..ipulses from muscle spindles, type II ..uscle afferents, and cutaneous affer- ..its to the cerebellum (Oscarsson, '65). ..lthough impulses from Golgi tendon ..gans are relayed via the dorsal nucleus, ..iis has not been established for the ac- ..ssory cuneate nucleus.

Dorsal roots C1 through T7 project fibers .. the accessory cuneate nucleus (fig. 6). ..revious investigations in the rhesus mon- ..ey have reported ascending dorsal root ..rojections to the accessory cuneate ..ucleus from levels as caudal as T4 (Fer- ..rro and Barrera, '35b) and T8 (Walker and ..leaver, '42). In the cat (Liu, '56) T5 was ..ie most caudal dorsal root reported to ..ontribute fibers to this nucleus, but dorsal ..oots T6 through T9 were not sectioned in ..iis study. Hand ('66) observed some ..egeneration from lumbar and S1 dorsal ..oots projecting to the rostral part of the ..ccessory cuneate nucleus. Classically, the ..uneocerebellar tract is considered to be ..elated primarily to the upper extremity ..nd neck, while the posterior spinocerebel- ..ar tract is considered to be related ..rimarily to the lower extremity and trunk ..Oscarsson, '65). The present study indi- ..ates that C5 through T7 dorsal roots ..roject to both the accessory cuneate and ..orsal nuclei, while C1 through C4 dorsal ..oots supply only the accessory cuneate ..nucleus. Since dorsal roots conveying ..afferents from the upper extremity and ..upper trunk have central projections to ..both of these nuclei, dual pathways exist ..or the conduction of impulses to the cere- ..bellum.

Previous investigators have reported pro- ..jections to the accessory cuneate nucleus ..after section of selected cervical dorsal roots in the marmoset monkey (Dunn and Matzke, '67), cat (Ranson, Davenport and Doles, '32; Pass, '33; Escolar, '48), and rabbit (Yee and Corbin, '39). None of these studies indicated the pattern or extent of the dorsal root projection.

According to Torvik's ('56) studies in the rat fibers from the upper three cervical dorsal roots terminate in the most ventral and lateral parts of the accessory cuneate nucleus. In the only detailed study of dorsal root projections to the accessory cuneate nucleus in the cat (Liu, '56), dorsal root fibers from spinal segments C1 through T5 were found to terminate in oblique, overlapping spiral laminae. Fibers from the C7 dorsal root terminated in the central core of the nucleus about which oblique laminae from other roots were arranged. Dorsal root fibers from T5 through C8 were found dorsal to the central core with T5 fibers most rostral and lateral, and C8 fibers most caudal and medial. Spiral laminae ventral to the central core were formed by dorsal root fibers from the upper six cervical segments with fibers from C6 most rostral and medial and fibers from C1 most caudal and lateral. Our interpretation of these observations, presented in a schematic reconstruction, suggests that medial and lateral as indicated in Liu's ('56) figure 8 are reversed.

Observations concerning the laminar termination of ascending dorsal root fibers in the accessory cuneate nucleus in the monkey have been based upon Marchi stained material. Sherrington (1893) found that C2 dorsal root fibers were distributed primarily to dorsal and lateral regions of the accessory cuneate nucleus. The arrangement described by Ferraro and Barrera ('35b) indicated that fibers from high cervical dorsal roots appeared to end in ventral and lateral parts of the nucleus, while those from lower cervical and upper thoracic dorsal roots appeared to end in dorsomedial regions of the accessory cuneate nucleus. Walker and Weaver ('42) reported similar findings except that fibers from C1 through C8 appeared to end in lateral regions of the nucleus while those of T1 through T8 passed to more medial locations. The latter authors indicated that while all of these dorsal roots projected fibers to both the cuneate and accessory cuneate nuclei, fibers passing to the acces-

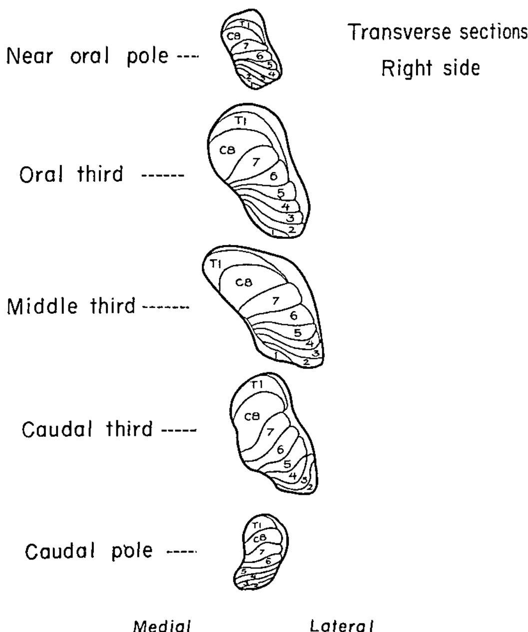


Fig. 6 Schematic diagram of the terminal zones in the accessory cuneate nucleus receiving fibers from dorsal roots C1 through T1. The zone for each root shows the principal area of fiber termination but not the extent of overlap. Fibers from dorsal roots T2 through T7 terminate in overlapping fashion mainly along the lateral border of the accessory cuneate nucleus.

sory cuneate nucleus were more numerous and more heavily myelinated. None of these authors specifically mentioned differences in the rostrocaudal distribution of various dorsal root fibers within the accessory cuneate nucleus.

In spite of specific differences, the laminar pattern of topical termination of dorsal root fibers in the accessory cuneate nucleus described by Liu ('56) appears to correspond more with the current observ tions than prior studies done in the mo key. While prior observations made in tl monkey are in general true, they lack si nificant details provided by silver stainir methods. In the monkey the central co region of the nucleus is the area of term nation of fibers from C5 through C8 dors roots. Other differences in the pattern laminar termination noted in the curre

dy concern the rostrocaudal disposition ibers from the upper three cervical dor- roots.

Although a C1 dorsal root is not present every monkey, when present, fibers of s root do not project to the caudal third the nucleus; fibers from this root are stributed only to ventromedial regions of e rostral two-thirds of the accessory ineate nucleus. Fibers from C2 and C3 orsal roots project to ventromedial gions of the nucleus throughout its ttent. Liu ('56) found fibers from C1 dor- l root localized to the caudal pole of the ccessory cuneate nucleus while fibers om C2 and C3 projected to ventrolateral egions of the caudal half of the nucleus ig. 6). Dorsal root projections from C8 ppear more extensive in the monkey than 1 the cat, and in the monkey they project aroughout dorsomedial regions of the ucleus. In the cat fibers of C8 were de- cribed as terminating only in dorsocaudal egions of the nucleus. Dorsal root fibers om T1 terminate dorsomedial to those of 8 and appear to cap the dorsomedial pole f the nucleus. The laminar terminations f dorsal root fibers T2 through T7 were not lotted in the present study, but it was vident that fibers from these roots rojected primarily to the lateral surface of he accessory cuneate nucleus.

Findings in the monkey related to scending dorsal root fibers terminating in he accessory cuneate nucleus (fig. 6) uggest that: (1) the number of fibers rojected to this nucleus is related to the ize of the dorsal root, (2) fibers from all ervical and the first thoracic dorsal roots erminate in an orderly sequence in both xclusive and overlapping zones, and (3) ibers from upper thoracic dorsal roots (i.e., 2 through T7) are less numerous, termi- iate in smaller regions of the nucleus and xhibit a higher degree of overlap. While lata are not precise with respect to over- apping terminations in this nucleus, fibers rom C1 through T1 dorsal roots partially overlap only the territory of the next high- est root. Comparisons of these results with hose of Liu ('56) indicate that the degree of overlapping projections in the monkey is much less than in the cat.

When the patterns of dorsal root termina- ions in the accessory cuneate and cuneate nuclei are compared, pertinent similarities and differences are evident. The patterns of termination of C1 through T1 dorsal root fibers in these nuclei are remarkably simi- lar. The C5 through C8 fibers terminated in the central core region about which fibers from the other dorsal roots are arranged in oblique laminae. The laminar pattern of termination of these dorsal roots is such that fibers from the highest root are ventro- lateral and those from the lowest root are dorsomedial. All of these dorsal roots, except C1 and C2, terminate throughout the rostrocaudal extent of both nuclei.

The patterns of termination of T2 through T7 dorsal root fibers in these nu- clei are different. Fibers from T2 through T7 dorsal roots terminate primarily in dorsomedial parts of the cuneate nucleus; in the accessory cuneate nucleus fibers from these roots terminate in lateral regions. Dorsal roots T5, T6 and T7 project fibers within both the medial border of the cuneate nucleus and the lateral border of the nucleus gracilis.

Central cervical and intermediate nuclei. Several authors have provided anatomical descriptions of the intermedi- ate nucleus in the upper cervical spinal cord and in the medulla of the cat (Cajal, '09; Papez, '29; Ranson, Davenport and Doles, '32; Rexed, '54) and the rat (Torvik, '56). These nuclei also are mentioned in studies of the cervical dorsal roots in the rabbit (Yee and Corbin, '39), cat (Kerr, '61) and monkey (Corbin, Lhamon and Petit, '37). The present study provides a descrip- tion of these nuclei in the rhesus monkey. According to Torvik ('56), the intermediate nucleus in the upper cervical spinal cord (Cajal, '09) probably is identical with the central cervical nucleus of Rexed ('54). Most authors (Papez, '29; Ranson, Daven- port and Doles, '32; Corbin, Lhamon and Petit, '37; Torvik, '56) indicate that the intermediate nucleus of the medulla is the rostral continuation of the intermediate, or central cervical, nucleus of the spinal cord. Rexed ('54), however, is firm in his belief that the central cervical nucleus is not directly continuous with any cell column in the medulla.

The present study indicates that the cen- tral cervical and intermediate nuclei are parts of the same cell column (figs. 1, 2),

within which cytological variations are seen. As Torvik ('56) indicated, cells at spinal levels and in caudal parts of the medulla are larger than those in more rostral regions. There has been little agreement concerning the rostral extent of this cell column. Corbin, Lhamon and Petit ('37) could not distinguish the intermediate nucleus at caudal levels of the hypoglossal nucleus in the monkey, and concluded that cells of the nucleus do not extend as far rostrally in the monkey as in the cat and rabbit. According to Torvik ('56) the rostral part of the intermediate nucleus lies slightly above the level of the caudal limit of the hypoglossal nucleus. Papez ('29) and Ranson, Davenport and Doles ('32) considered the oral part of the intermediate nucleus to lie along the lateral border of the hypoglossal nucleus and to virtually merge with the nucleus intercalatus. The present study indicates that the most rostral part of the intermediate nucleus in the monkey extends slightly beyond the caudal pole of the dorsal motor nucleus of the vagus. Although the rostral part of the nucleus is small, it is distinct and separate from the nucleus intercalatus. In normal material the intermediate nucleus does not stand out as an impressive cell column, but following section of the C2 dorsal root, extensive arborizations about cells of the nucleus outline the borders of the cell column in an impressive way (figs. 33, 34). The thesis that the central cervical nucleus and intermediate nucleus of the medulla constitute a single cell column is based upon: (1) anatomical evidence of a continuous aggregation of cells in corresponding locations in the upper cervical spinal cord and lower medulla, (2) the observation that cells in various parts of this column share at least one common source of afferent fibers, and (3) the finding that dorsal root fibers in all parts of the cell column terminate in similar profuse pericellular arborizations. The manner in which dorsal root fibers terminate upon cells in these nuclei (fig. 35) resembles that seem in the dorsal nucleus (Szentágothai and Albert, '55).

The present observations confirm that single or multiple section of the upper three cervical dorsal roots produces profuse degeneration in the central cervical nucleus (Ranson, Davenport and D[illegible] '32; Corbin and Hinsey, '35; Corb[illegible] Lhamon and Petit, '37; Yee and Corb[illegible] '39; Escolar, '48; Torvik, '56). Maxim[illegible] degeneration in this nucleus has be[illegible] found at the level of the sectioned dor[illegible] root. Current data also are consistent w[illegible] the observations of Corbin and Hinsey ('[illegible] and those of Yee and Corbin ('39) that [illegible] and C5 dorsal roots project fibers throug[illegible] out the rostrocaudal extent of the centr[illegible] cervical nucleus. All of the upper fi[illegible] cervical dorsal roots, except C1, proje[illegible] fibers to all parts of the central cer[illegible] nucleus and to portions of the intermedi[illegible] nucleus of the medulla. Dorsal root fibe[illegible] from C1 project to the central cervic[illegible] nucleus in the upper two cervical se[illegible] ments and contribute modestly to caud[illegible] parts of the intermediate nucleus of t[illegible] medulla. Profuse degeneration from t[illegible] C2 dorsal root projects to all parts of t[illegible] central cervical and intermediate nucl[illegible] Dorsal roots C3 through C5 contribu[illegible] fibers to the central cervical nucleus at [illegible] spinal levels and to all, except the mo[illegible] rostral, parts of the intermediate nucle[illegible] Ascending fibers from these dorsal ro[illegible] extend rostrally to levels slightly caudal [illegible] the hypoglossal nucleus. The upper fi[illegible] cervical dorsal roots all have extensi[illegible] overlapping projections to the cent[illegible] cervical and intermediate nuclei with t[illegible] C2 dorsal root having the most extensi[illegible] distribution. The overlapping nature [illegible] dorsal root projections to the central cer[illegible] cal nucleus has been demonstrated [illegible] Schimert ('39) and Kerr ('61).

Dorsal root projections to the interme[illegible] ate nucleus of the medulla have been [illegible] scribed only from the upper three cervi[illegible] roots (Ranson, Davenport and Doles, '[illegible] Corbin, Lhamon and Petit, '37; Yee [illegible] Corbin, '39). The present findings d[illegible] from those of Yee and Corbin ('39) w[illegible] found that the C1 dorsal root projects [illegible] greatest number of fibers to the interme[illegible] ate nucleus in the rabbit.

Hypotheses concerning the functio[illegible] significance of the intermediate nucle[illegible] seem to have been based mainly upon a[illegible] tomical considerations. Papez ('29) su[illegible] gested that the intermediate nucleus w[illegible] associated with the hypoglossal nucle[illegible] and received muscle afferents from t[illegible]

ngue via a reflexo-motor bundle from C1 rsal root (i.e., descendens hypoglossi). anson, Davenport and Doles ('32) considered that this nucleus might be involved in nic neck and righting reflexes, but aditted that the bulbar portion of the nueus could not be essential since the tonic eck reflex can still be obtained after tranction of the spinal cord above the level of e C1 dorsal root. Rexed ('54) suggested at the central cervical nucleus might be caudal continuation of the hypoglossal ucleus. He postulated that fibers from is nucleus might exit with the ventral oots of C1 through C3 (or C4), join the ypoglossal nerve to form the descending amus, and innervate muscles attached to e hyoid bone.

Evidence from the present study suggests *that the central cervical and intermediate* nuclei may constitute an important relay nucleus in an ascending pathway. Extensive cerebellar cortical ablaions in two monkeys have not provided lata indicating that fibers of this nucleus roject to the cerebellum, but these experinents cannot be regarded as conclusive.

Supraspinal nucleus. Throughout levels of the pyramidal decussation, scattered neurons of the ventromedial nuclear group of the anterior gray horn extend rostrally nto the medulla. These neurons, termed he supraspinal nucleus (Jacobsohn, '09; Olszewski and Baxter, '54) connect spinal and medullary (hypoglossal nucleus) porions of the general somatic efferent cell column. The supraspinal nucleus is considered to give rise to the C1 ventral root ibers.

In the monkey, the upper five cervical dorsal roots project fibers to the supraspinal nucleus. Within the nucleus most fibers terminate dorsomedially. The first two dorsal roots contribute the greatest number of fibers to the supraspinal nucleus; projections from the remaining cervical dorsal roots progressively decrease in a caudal sequence. Although numerous investigators have observed dorsal root projections to the ventromedial portion of the anterior horn at the C1 level, projections to its rostral continuation, the supraspinal nucleus, have not been reported.

Other medullary nuclei. Several authors have reported that ascending fibers from upper cervical dorsal roots project to: (1) nucleus and tractus solitarius (Ranson, Davenport and Doles, '32 Corbin and Hinsey, '35; Yee and Corbin, '39; Liu, '56; Torvik, '56), (2) the inferior vestibular nucleus (Corbin and Hinsey, '35; Yee and Corbin, '39; Escolar, '48; Dunn and Matzke, '67), (3) the commissural nucleus of the vagus (Schwartz, Roulhac, Lam and O'Leary, '51; Torvik, '56), and (4) the nucleus intercalatus and the nucleus of Roller (Torvik, '56).

In the monkey a few ascending fibers from the C2 dorsal root projected to parts of the nucleus solitarius in two animals (C-1006 and C-1024). In these animals small, but definite, projections from the fasciculus cuneatus entered the magnocellular part of the spinal trigeminal nucleus, the *nucleus of Roller, the nucleus* intercalatus and the parvocellular reticular formation. One other animal (C-912) had minimal degeneration in the nucleus intercalatus associated with a C3 dorsal rhizotomy. These observations with respect to the nucleus solitarius differ from those of Corbin, Lhamon and Petit ('37), Schwartz, Roulhac, Lam and O'Leary ('51) and Kerr ('61).

One point which might account for the above discrepancies relative to the inferior vestibular nucleus concerns cell group x (Brodal and Pompeiano, '57), a small aggregation of cells lateral to the caudal part of the inferior vestibular nucleus and rostral to the accessory cuneate nucleus. This cell group has been considered by some as part of the inferior vestibular nucleus, although it does not receive primary vestibular fibers (Walberg, Bowsher and Brodal, '58). Spinovestibular afferents to this nucleus course with fibers of the posterior spinocerebellar tract. No degenerated fibers projecting to this nuclear group were found following C6 through C8 dorsal rhizotomies or cord lesions restricted to the posterior columns at C2 and C4 (Pompeiano and Brodal, '57). Observations in this study indicate that inadvertent trauma to the posterior spinocerebellar tract invariably resulted in degeneration projecting to cell group x. The extent of the degeneration in this nucleus seemed proportional to that in the posterior spinocerebellar tract. However,

within which cytological variations are seen. As Torvik ('56) indicated, cells at spinal levels and in caudal parts of the medulla are larger than those in more rostral regions. There has been little agreement concerning the rostral extent of this cell column. Corbin, Lhamon and Petit ('37) could not distinguish the intermediate nucleus at caudal levels of the hypoglossal nucleus in the monkey, and concluded that cells of the nucleus do not extend as far rostrally in the monkey as in the cat and rabbit. According to Torvik ('56) the rostral part of the intermediate nucleus lies slightly above the level of the caudal limit of the hypoglossal nucleus. Papez ('29) and Ranson, Davenport and Doles ('32) considered the oral part of the intermediate nucleus to lie along the lateral border of the hypoglossal nucleus and to virtually merge with the nucleus intercalatus. The present study indicates that the most rostral part of the intermediate nucleus in the monkey extends slightly beyond the caudal pole of the dorsal motor nucleus of the vagus. Although the rostral part of the nucleus is small, it is distinct and separate from the nucleus intercalatus. In normal material the intermediate nucleus does not stand out as an impressive cell column, but following section of the C2 dorsal root, extensive arborizations about cells of the nucleus outline the borders of the cell column in an impressive way (figs. 33, 34). The thesis that the central cervical nucleus and intermediate nucleus of the medulla constitute a single cell column is based upon: (1) anatomical evidence of a continuous aggregation of cells in corresponding locations in the upper cervical spinal cord and lower medulla, (2) the observation that cells in various parts of this column share at least one common source of afferent fibers, and (3) the finding that dorsal root fibers in all parts of the cell column terminate in similar profuse pericellular arborizations. The manner in which dorsal root fibers terminate upon cells in these nuclei (fig. 35) resembles that seem in the dorsal nucleus (Szentágothai and Albert, '55).

The present observations confirm that single or multiple section of the upper three cervical dorsal roots produces profuse degeneration in the central cervical nucleus (Ranson, Davenport and Dol '32; Corbin and Hinsey, '35; Corb Lhamon and Petit, '37; Yee and Corb '39; Escolar, '48; Torvik, '56). Maxi degeneration in this nucleus has be found at the level of the sectioned dor root. Current data also are consistent wi the observations of Corbin and Hinsey ('3 and those of Yee and Corbin ('39) that C and C5 dorsal roots project fibers throug out the rostrocaudal extent of the centr cervical nucleus. All of the upper fi cervical dorsal roots, except C1, proje fibers to all parts of the central cervic nucleus and to portions of the intermedia nucleus of the medulla. Dorsal root fiber from C1 project to the central cervica nucleus in the upper two cervical seg ments and contribute modestly to caud parts of the intermediate nucleus of th medulla. Profuse degeneration from th C2 dorsal root projects to all parts of th central cervical and intermediate nucle Dorsal roots C3 through C5 contribute fibers to the central cervical nucleus at al spinal levels and to all, except the mos rostral, parts of the intermediate nucleus Ascending fibers from these dorsal roots extend rostrally to levels slightly caudal to the hypoglossal nucleus. The upper five cervical dorsal roots all have extensiv overlapping projections to the centra cervical and intermediate nuclei with th C2 dorsal root having the most extensiv distribution. The overlapping nature c dorsal root projections to the central cerv cal nucleus has been demonstrated b Schimert ('39) and Kerr ('61).

Dorsal root projections to the intermedi ate nucleus of the medulla have been de scribed only from the upper three cervica roots (Ranson, Davenport and Doles, '3 Corbin, Lhamon and Petit, '37; Yee an Corbin, '39). The present findings diffe from those of Yee and Corbin ('39) wh found that the C1 dorsal root projects th greatest number of fibers to the intermed ate nucleus in the rabbit.

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rminate primarily in *dorsomedial parts*
the cuneate nucleus and in lateral parts
the accessory cuneate nucleus.

15. Cytological and degeneration studies iggest that the central cervical nucleus nd the intermediate nucleus of the iedulla: (a) constitute parts of the same ell column, and (b) *receive fibers from the* pper five cervical dorsal roots which overap in large regions of the cell column and erminate in rich pericellular arborizaions. Afferent fibers to all parts of this cell olumn are derived from the C2 dorsal oot.

16. The upper five cervical dorsal roots project fibers to the supraspinal nucleus, primarily to the dorsomedial part. The greatest number of fibers terminating in his nucleus are derived from the C1 and C2 *dorsal roots.*

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in some animals small amounts of terminal degeneration were present in cell group x, even though none was detectable in the posterior spinocerebellar tract or in the inferior cerebellar peduncle.

CONCLUSIONS

The following conclusions were drawn from this study:

1. Degeneration in the spinal cord is most profuse at the level of dorsal rhizotomy and progressively diminishes in segments rostral and caudal to the sectioned root; progressive diminution of degeneration is greatest in the anterior horn.

2. At the level of dorsal rhizotomy, profuse terminal degeneration is present in laminae III and IV, central and medial portions of laminae V and VI, and among certain cell groups of lamina IX. Fewer fibers appear to terminate in laminae VII and VIII.

3. No dorsal root fibers from C1 project into the head of the posterior horn in the upper cervical region and fibers passing to the anterior horn are sparse.

4. Terminal fibers from dorsal roots C2 through C5 are localized to particular portions of laminae III and IV in a manner suggesting a somatotopic arrangement.

5. Fibers from dorsal roots of the brachial enlargement project to central and medial parts of lamina VI from four (C5) to eight (T1) segments rostral to their entry.

6. Cervical and upper thoracic dorsal roots project fibers to cell groups of lamina IX by distinctive pathways through either medial, or central, parts of laminae VII and VIII. At different levels the number of fibers following these pathways varies so that the greatest number of fibers from: (a) C5 through T1 dorsal roots course through central parts of lamina VII, (b) T2 through T7 dorsal roots project via medial parts of laminae VII and VIII, and (c) C2 through C4 dorsal roots project through both medial and central parts of laminae VII and VIII.

7. Cell groups of the spinal accessory nucleus in the upper cervical cord receive fibers from C2 through C5 dorsal roots with C3 contributing the greatest number of fibers.

8. Overlapping projections to the dors nucleus are derived from C5 through dorsal roots; single dorsal roots distribu fibers to this cell column in four to spinal segments, with the most profu degeneration: (a) derived from thorac dorsal roots, and (b) terminating in tions of the cell column nearest the sec tioned root.

9. A small, but distinct, bundle of from all cervical and all upper thoraci dorsal roots terminate in the intermedi medial nucleus and the area ventral to at the level of root entrance, and in adj cent spinal segments.

10. Ascending fibers in the fasciculu cuneatus in the monkey: (a) arise from the cervical and upper seven thoracic dorsal roots, and (b) exhibit only a general segregation because of extensive overlap. Dor sal roots of the brachial enlargemen contribute the largest number of fibers

11. Descending collaterals of cervical and upper thoracic dorsal roots form discrete bands in the posterior funiculus, lateral and parallel to, but at varying distances from the posterior intermediate septum. These bands resemble those of ascending fibers of the same root, except that they do not migrate as far dorsomedially, have a greater concentration of fibers in ventromedial locations, and are smaller

12. Ascending branches of fibers from all cervical and the upper seven thoracic dorsal roots project somatotopically to both the cuneate and accessory cuneate nuclei The pattern of termination of fibers from C1 through T1 dorsal roots in these nuclei is similar in that fibers from: (a) C5 through C8 terminate in a central core region about which other dorsal root fibers terminate in oblique serial laminae, (b) rostral roots terminate in ventrolateral regions, while those from caudal roots end in dorsomedial regions, and (c) all of these roots, except C1 and C2, terminate throughout the rostrocaudal extent of both nuclei.

13. Ascending fibers from C1 through T1 dorsal roots projecting to the cuneate and accessory cuneate nuclei terminate in both exclusive and overlapping zones; fiber from one dorsal root partially overlap th territory of the next highest root.

14. Fibers from T2 through T7 dorsa

nts projecting to the cuneate and accessory cuneate nuclei terminate in smaller regions of the nuclei, exhibit greater overlap and are distinctive in that these fibers terminate primarily in dorsomedial parts of the cuneate nucleus and in lateral parts of the accessory cuneate nucleus.

15. Cytological and degeneration studies suggest that the central cervical nucleus and the intermediate nucleus of the medulla: (a) constitute parts of the same cell column, and (b) receive fibers from the upper five cervical dorsal roots which overlap in large regions of the cell column and terminate in rich pericellular arborizations. Afferent fibers to all parts of this cell column are derived from the C2 dorsal root.

16. The upper five cervical dorsal roots project fibers to the supraspinal nucleus, primarily to the dorsomedial part. The greatest number of fibers terminating in this nucleus are derived from the C1 and C2 dorsal roots.

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in some animals small amounts of terminal degeneration were present in cell group x, even though none was detectable in the posterior spinocerebellar tract or in the inferior cerebellar peduncle.

CONCLUSIONS

The following conclusions were drawn from this study:

1. Degeneration in the spinal cord is most profuse at the level of dorsal rhizotomy and progressively diminishes in segments rostral and caudal to the sectioned root; progressive diminution of degeneration is greatest in the anterior horn.

2. At the level of dorsal rhizotomy, profuse terminal degeneration is present in laminae III and IV, central and medial portions of laminae V and VI, and among certain cell groups of lamina IX. Fewer fibers appear to terminate in laminae VII and VIII.

3. No dorsal root fibers from C1 project into the head of the posterior horn in the upper cervical region and fibers passing to the anterior horn are sparse.

4. Terminal fibers from dorsal roots C2 through C5 are localized to particular portions of laminae III and IV in a manner suggesting a somatotopic arrangement.

5. Fibers from dorsal roots of the brachial enlargement project to central and medial parts of lamina VI from four (C5) to eight (T1) segments rostral to their entry.

6. Cervical and upper thoracic dorsal roots project fibers to cell groups of lamina IX by distinctive pathways through either medial, or central, parts of laminae VII and VIII. At different levels the number of fibers following these pathways varies so that the greatest number of fibers from: (a) C5 through T1 dorsal roots course through central parts of lamina VII, (b) T2 through T7 dorsal roots project via medial parts of laminae VII and VIII, and (c) C2 through C4 dorsal roots project through both medial and central parts of laminae VII and VIII.

7. Cell groups of the spinal accessory nucleus in the upper cervical cord receive fibers from C2 through C5 dorsal roots with C3 contributing the greatest number of fibers.

8. Overlapping projections to the dors nucleus are derived from C5 through dorsal roots; single dorsal roots distrib fibers to this cell column in four to t spinal segments, with the most profu degeneration: (a) derived from dorsal roots, and (b) terminating in tions of the cell column nearest the se tioned root.

9. A small, but distinct, bundle of fibe from all cervical and all upper thorac dorsal roots terminate in the intermedi medial nucleus and the area ventral to at the level of root entrance, and in ad cent spinal segments.

10. Ascending fibers in the fascicul cuneatus in the monkey: (a) arise from t cervical and upper seven thoracic dor roots, and (b) exhibit only a general seg gation because of extensive overlap. D sal roots of the brachial enlargem contribute the largest number of fibers.

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12. Ascending branches of fibers all cervical and the upper seven thor dorsal roots project somatotopically to the cuneate and accessory cuneate nu The pattern of termination of fibers f C1 through T1 dorsal roots in these nu is similar in that fibers from: (a) through C8 terminate in a central region about which other dorsal root fi terminate in oblique serial laminae rostral roots terminate in ventrola regions, while those from caudal roots in dorsomedial regions, and (c) al these roots, except C1 and C2, termi throughout the rostrocaudal extent of nuclei.

13. Ascending fibers from C1 throug dorsal roots projecting to the cuneate accessory cuneate nuclei terminate in exclusive and overlapping zones; fi from one dorsal root partially overla territory of the next highest root.

14. Fibers from T2 through T7 d

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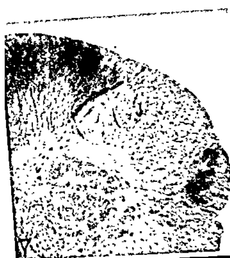
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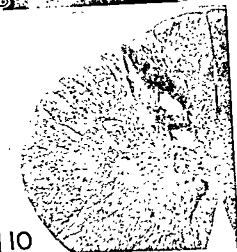
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PLATE 1

EXPLANATION OF FIGURES

7 Rhesus C-1008. Photomicrograph of degeneration following section of the C1 dorsal root on the right. Degeneration is present dorsal to the head of the posterior horn at C1. Nauta-Gygax. × 15.

8 Rhesus C-978. Photomicrograph of degeneration in root entry zone following section of the C3 dorsal root on the right. Note degeneration in the central part of laminae III and IV and fibers coursing ventrally through lamina V into the central cervical nucleus. Nauta-Gygax. × 16.

9 Rhesus C-1014. Photomicrograph of degeneration at C5 following section of the dorsal root at that level. Note fibers passing medially to the intermediomedial nucleus ventrolateral to the central canal. Nauta-Gygax. × 21.

10 Rhesus C-913. Photomicrograph of degeneration at T2 following section of the dorsal root at that level. Root fibers enter laminae III and IV mainly by coursing through medial parts of lamina II. Nauta-Gygax. × 16.

11 Rhesus C-1014. Photomicrograph of ascending degeneration in the fasciculi cuneatus resulting from section of dorsal roots C5 (left) and C8 (right). Less intense degeneration in the fasciculi gracilis resulted from section of dorsal roots L6 (left) and Co1 (right). Nauta-Gygax. × 13.

12 Rhesus C-1025. Photomicrograph of degeneration at T7 following section of the dorsal root at that level. Degeneration is present in the fasciculus cuneatus, laminae III and IV and in the dorsal nucleus. Nauta-Gygax. × 64.

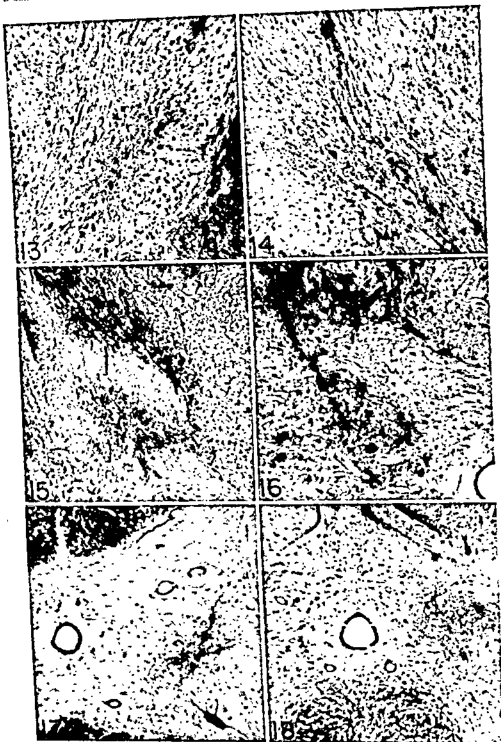

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PLATE 2

EXPLANATION OF FIGURES

13 Rhesus C-981. Photomicrograph of terminal degeneration localized to the lateral parts of laminae III and IV on the right side at C2 following section of the dorsal root at that level. Nauta-Gygax. ×95.

14 Rhesus C-981. Photomicrograph of terminal degeneration localized to the lateral parts of laminae III and IV on the left side at C5 following section of the C4 dorsal root. Nauta-Gygax. × 105.

15 Rhesus C-913. Photomicrograph of terminal degeneration in laminae III and IV on the left side at T2 following section of the dorsal root at that level. Nauta-Gygax. × 50.

16 Rhesus C-913. Photomicrograph of degenerated fibers from the T2 dorsal root projecting to the dorsal nucleus at T3 on the left. Nauta-Gygax. ×95.

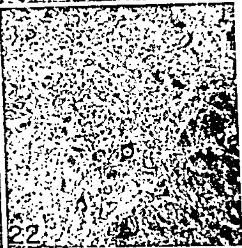
17 Rhesus C-1014. Photomicrograph of C8 dorsal root fibers projecting to the intermediomedial nucleus at C7 on the right side. Nauta-Gygax. ×95.

18 Rhesus C-1024. Photomicrograph of T5 dorsal root fibers projecting to the dorsal nucleus at the level of root section. Nauta-Gygax. ×95.


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PLATE 3

EXPLANATION OF FIGURES

19 Rhesus C-1006. Photomicrograph of fibers from the C2 dorsal root arborizing about cells of the central cervical nucleus at the C2 level. Nauta-Gygax. × 100.

20 Rhesus C-913. Photomicrograph of fibers from the C2 dorsal root arborizing about cells of the central cervical nucleus at the level of root section. Nauta-Gygax. × 240.

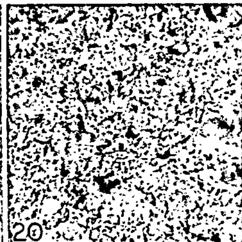
21 Rhesus C-1014. Photomicrograph of C8 dorsal root fibers passing from the fasciculus cuneatus into the medial part of lamina VI at C5. Nauta-Gygax. × 50.

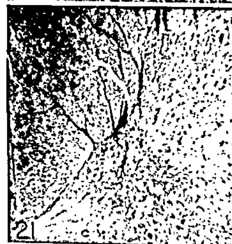
22 Rhesus C-978. Photomicrograph of C3 dorsal root fibers passing to the spinal accessory cell group of lamina IX at the C3 level. Nauta-Gygax. × 125.

23 Rhesus C-1014. Photomicrograph of C8 dorsal root fibers on the right projecting into motor cell groups of lamina IX at the C7 level. Nauta-Gygax. × 240.

24 Rhesus C-970. Photomicrograph of T4 dorsal root fibers on the left projecting ventrally through the medial parts of laminae VII and VIII to cell groups of lamina IX at T4. Nauta-Gygax. × 100


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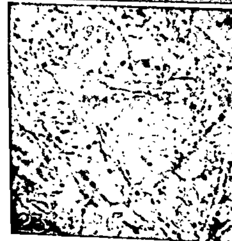




PLATE 4

EXPLANATION OF FIGURES

25 Rhesus C-1008. Photomicrograph of degenerated fibers from the C1 dorsal root terminating in the most ventral part of the accessory cuneate nucleus. Nauta-Gygax. × 40.

26 Rhesus C-1006. Photomicrograph of degenerated fibers from the C2 dorsal root terminating in ventral portions of both the cuneate and accessory cuneate nuclei. Nauta-Gygax. × 16.

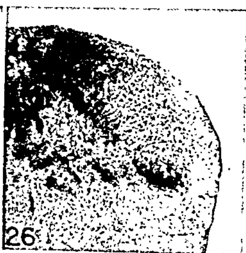
27 Rhesus C-974. Photomicrograph of degenerated C5 dorsal root fibers terminating in localized zones of the cuneate and accessory cuneate nuclei. Nauta-Gygax. × 16.

28 Rhesus C-967. Photomicrograph of degenerated C6 dorsal root fibers terminating in localized zones of the cuneate and accessory cuneate nuclei. Nauta-Gygax. × 13.

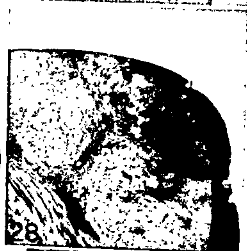
29 Rhesus C-1014. Photomicrograph of C8 dorsal root fibers terminating in localized zones of the cuneate and accessory cuneate nuclei. Nauta-Gygax. × 16.

30 Rhesus C-913. Photomicrograph of a horizontal brain stem section showing terminal degeneration from the T2 dorsal root in the lateral part of the left accessory cuneate nucleus. Nauta-Gygax. × 50.


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PLATE 4

EXPLANATION OF FIGURES

25 Rhesus C-1008. Photomicrograph of degenerated fibers from the C1 dorsal root terminating in the most ventral part of the accessory cuneate nucleus. Nauta-Gygax. ×40.

26 Rhesus C-1006. Photomicrograph of degenerated fibers from the C2 dorsal root terminating in ventral portions of both the cuneate and accessory cuneate nuclei. Nauta-Gygax. × 16.

27 Rhesus C-974. Photomicrograph of degenerated C5 dorsal root fibers terminating in localized zones of the cuneate and accessory cuneate nuclei. Nauta-Gygax. × 16.

28 Rhesus C-967. Photomicrograph of degenerated C6 dorsal root fibers terminating in localized zones of the cuneate and accessory cuneate nuclei. Nauta-Gygax. × 13.

29 Rhesus C-1014. Photomicrograph of C8 dorsal root fibers terminating in localized zones of the cuneate and accessory cuneate nuclei. Nauta-Gygax. × 16.

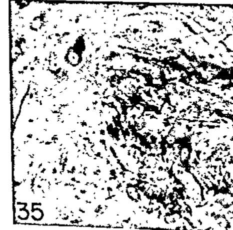
30 Rhesus C-913. Photomicrograph of a horizontal brain stem section showing terminal degeneration from the T2 dorsal root in the lateral part of the left accessory cuneate nucleus. Nauta-Gygax. × 50.


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PLATE 5

EXPLANATION OF FIGURES

31 Rhesus C-913. Photomicrograph of horizontal brain stem section showing ascending fibers from the C2 dorsal root projecting profusely to the accessory cuneate nucleus and giving off small fascicles to the cuneate nucleus. Nauta-Gygax. × 15.

32 Rhesus C-915. Photomicrograph of a horizontal brain stem section showing terminal degeneration from the C7 dorsal root in the central part of the cuneate nucleus. Nauta-Gygax. × 14.

33-34 Rhesus C-970. Photomicrographs of C2 dorsal root fibers arborizing about clusters of cells in the intermediate nucleus of the medulla. Nauta-Gygax. × 40, × 100

35 Rhesus C-970. Photomicrograph of terminal degeneration from the C2 *dorsal root in the rostral part of the intermediate nucleus. A few cells of* the dorsal motor nucleus of the vagus nerve can be seen in the upper left Nauta-Gygax. × 260.

36 Rhesus C-1008. Photomicrograph of degenerated C1 dorsal root fibers projecting into the region of the supraspinal nucleus. Nauta-Gygax. × 240.

Central Projections of Spinal Dorsal Roots in the Monkey[1]

II. LOWER THORACIC, LUMBOSACRAL AND COCCYGEAL DORSAL ROOTS

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ABSTRACT A study of the central distribution of lower thoracic, lumbosacral and coccygeal dorsal roots (DR) was made in 11 monkeys. Degeneration resulting from 17 individual dorsal rhizotomies was studied in multiple spinal and medullary sections stained by the Nauta technic. At the rhizotomy level all DR project fibers to: (1) portions of laminae III and IV, (2) the intermediomedial nucleus, and (3) portions of laminae VII, VIII and IX. Lower thoracic, lumbar and the first sacral (S1) DR project in overlapping fashion to particular portions of Clarke's column, ascending and descending fibers terminate in a spatially organized manner. Lower lumbar DR project to medial lamina VI throughout the lumbar enlargement. DR fibers reach cell groups of lamina IX via two trajectories, one of which is dominant in the lumbar enlargement. Ascending DR fibers in the fasciculus gracilis. (1) arise caudal to T7, (2) exhibit only general segregation due to overlap, and (3) project in an overlapping somatotopic fashion throughout the rostrocaudal extent of the nucleus gracilis. Zones of the nucleus gracilis receiving fibers are organized, so that (1) DR of the lumbar enlargement project to a central core region, (2) lower thoracic and upper lumbar DR project to narrow oblique laminae lateral to the core, and (3) S1 and Co1 DR project to crescent-shaped dorsomedial regions. Less autonomous terminal representation of DR fibers was found in the nucleus gracilis than in other posterior column nuclei.

As emphasized in Part I of this study (Shriver, Stein and Carpenter, '68), most previous anatomical investigations of the central projections of dorsal root fibers have been based upon the Marchi method and section of a limited number of dorsal roots in restricted regions of the spinal cord. Studies of the projections of certain lower thoracic, lumbosacral and coccygeal dorsal roots, done in the cat and dog (Szentágothai and Albert, '55; Liu, '56; Grant and Rexed, '58; Szentágothai, '64; Sprague and Ha, '64; Kuypers and Tuerk, '64; Hand, '66), have utilized silver staining methods. Investigations in the monkey (Ferraro and Barrera, '34, '35; Walker and Weaver, '42; Chang and Ruch, '47), based upon partial cordotomies and selective dorsal root sections, have used the Marchi method. Although Marchi studies have contributed little information concerning projections of dorsal root fibers within the spinal gray, they have provided information concerning the somatotopic organization of ascending fibers in the posterior columns and in the nucleus gracilis. Studies using silver staining methods have provided important details concerning the distribution of dorsal root fibers in the spinal gray (Szentágothai and Albert, '55; Liu, '56; Grant and Rexed, '58; Szentágothai, '64; Sprague and Ha, '64) and information regarding termination of fibers in the nucleus gracilis (Kuypers and Tuerk, '64; Hand, '66). Recent anatomical (Kuypers and Tuerk, '64; Hand, '66) and physiological (Kuhn, '49; Gordon and Paine, '60; Perl, Whitlock and Gentry, '62; McComas, '63; Winter, '65) studies suggest that dorsal root fibers may terminate in a dual fashion in different parts of the nucleus gracilis, which has been interpreted to suggest rostrocaudal differences in the size of the receptor fields and/or differences in modal-

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the intermediolateral cell group. Fibers passed from lamina VII ventrally into medial parts of lamina VIII and followed the border of the spinal gray into lamina IX. Within lamina IX degeneration was maximal about cells of the ventromedial group. Only a few fibers traversing central parts of laminae VII and VIII reached cells of the lateromedial cell group of lamina IX.

At the T9 spinal segment a marked reduction in the quantity of degeneration was observed. In rhesus C-970 degeneration in laminae III and IV was seen in lateral regions, while degeneration was present in medial parts of these laminae in rhesus C-1017. Slight differences were noted in the location of degeneration in the dorsal nucleus. In rhesus C-970 it was present in lateral portions, but in C-1017 it occupied dorsal regions of the nucleus. Only relatively sparse degeneration was seen in laminae VIII and IX.

At the T7 spinal segment reduced degeneration similar to that present at the T9 level was observed. This degeneration was principally in the central and medial portions of laminae III and IV, dorsolateral portions of the dorsal nucleus, and adjacent portions of lamina VII. Degeneration was sparse in the medial part of lamina VIII and in lamina IX.

Descending degeneration from the T8 dorsal root in both animals formed a narrow band in the lateral third of the posterior column separated from the posterior horn by a zone of normal fibers. This degeneration was traced as far caudally as T12 (C-970) and L2 (C-1017). Fibers given off in lower thoracic segments in rhesus C-970 were confined principally to parts of lamina VII and the dorsal nucleus; degeneration was maximal in lateral portions of the dorsal nucleus caudal to T10 in both animals.

Similar ascending degeneration in both animals occupied progressively more medial parts of the posterior column. At the T4 level the ventromedial portion of the band of ascending degeneration was present in the lateral part of the fasciculus gracilis, while the dorsolateral portion was located in medial parts of the fasciculus cuneatus (fig. 13). A gradual shift of the degeneration occurred rostrally so that at C8 only a tiny amount of the degeneration was present in the dorsomedial part of the fasciculus cuneatus and the majority of the degeneration occupied the lateral border of the fasciculus gracilis. Thus, ascending degeneration was present in different locations on both sides of the posterior intermediate septum in upper thoracic segments. Ascending degeneration within the spinal gray rostral to T7 was confined to the dorsal nucleus and the adjacent portions of lamina VII. Degeneration within the dorsal nucleus, primarily the dorsolateral part, was seen as far rostrally as T5 (C-970) and T2 (C-1017).

Sections of the medulla were cut horizontally (C-970) and transversely (C-999 and C-1017). Posterior column degeneration in the three animals was contained in lateral portions of the fasciculus gracilis at caudal medullary levels. Relatively few fibers appeared to terminate in ventral parts of the nucleus gracilis at this level. In sections through mid- and rostral parts of the nucleus gracilis moderate amounts of terminal degeneration were seen within the lateral border of the nucleus (fig. 31). Initially these degenerated fibers occupied nearly the entire lateral border of the nucleus, but further rostrally they were confined to a smaller, more dorsal region (fig. 5). Fiber terminations were greatest in the rostral third of the nucleus gracilis. A small, but distinct, number of fibers entered the most medial part of the nucleus cuneatus in those regions where the nuclei gracilis and cuneatus have a common border. Rostrally degenerated fibers tended to invade more dorsal portions of the medial border of the cuneate nucleus.

In rhesus C-999 moderate degeneration was seen in cell group x and in portions of the inferior vestibular nucleus (Brodal and Pompeiano, '57). Degeneration in these nuclei appeared to be related to ascending degeneration of spinovestibular fibers (Pompeiano and Brodal, '57) which course in association with fibers of the posterior spinocerebellar tract.

Ninth thoracic root. In rhesus C-999 the ninth thoracic dorsal root was sectioned, but this operation, like that on the opposite side at T8, was complicated by unintentional injury to the lateral and anterior funiculi of the spinal cord.

ity segregation. In view of these data which are difficult to reconcile with classical concepts, further information concerning the termination of dorsal root fibers in the nucleus gracilis seems needed.

Material presented in this report is a continuation of Part I of a study of the central projections of dorsal root fibers in the monkey. In Part II anatomical data concerning lower thoracic, lumbosacral and coccygeal dorsal root projections are presented. Attempts have been made to compare observations in the two parts of this study, and to correlate certain anatomical and physiological data.

MATERIAL AND METHODS

In 11 rhesus monkeys single dorsal rhizotomies were done at different levels and on different sides. As reported in Part I, three or four different dorsal roots at selected levels were sometimes sectioned in individual animals to conserve time, effort and material. Table 1 supplements similar data contained in Part I of this study. Some of the animals were used in both parts of this study. Dorsal roots sectioned included four lower thoracic (i.e., T8, T9, T10 and T12), all seven lumbar, and the first sacral and coccygeal. A total of 17 dorsal rhizotomies were done at the levels mentioned.

The operative procedures, histological technics and methods of study used here were the same as those described in Part I of this study.

OBSERVATIONS

Lower thoracic dorsal roots

Four lower thoracic dorsal roots (i.e., T8 T9, T10 and T12) were sectioned individually in five monkeys.

Eighth thoracic dorsal root. The eighth thoracic dorsal root was sectioned in three animals (C-970, C-999 and C-1017). In one animal (C-999) dorsal rhizotomy was complicated by bilateral injury to the anterior and lateral funiculi which precluded evaluation of the degenerat within the spinal gray. Spinal and medullary degeneration in the other two animals was similar but not identical.

Degenerated fibers from the root en zone projected into the most lateral portio of the posterior column and into the zone of Lissauer. Fibers in ventromedial parts of the fasciculus gracilis approached the posterior median septum. Degenerated fibers coursed obliquely through medial portions of lamina II and arborize profusely about cells in medial and centr parts of laminae III and IV (fig. 7). Relatively sparse degeneration was seen in lamina V. In rhesus C-970 diffuse degeneration was present in all parts of the dors nucleus, but in rhesus C-1017 degenera tion in the dorsal nucleus was foun mainly in ventrolateral regions. A small amount of degeneration was seen in po tions of lamina VII lateral to the dors nucleus, and an island of degener was noted within the intermediomedi nucleus. No degeneration was present

TABLE 1

Dorsal roots sectioned in eleven monkeys

Animal Number	Spinal roots																								
	C1	C2	C3	C5	C6	C7	C8	T1	T2	T4	T6	T7	T8	T9	T10	T12	L1	L2	L3	L4	L5	L6	L7	S1	Col
C-912			R					L									R							L	
C-913*		R							L									R							
C-914				R			L												R				L		
C-915*						R						L								R					
C-916	R														L							R			
C-970*		R								L			R												
C-999													R	L											
C-1011																				R					
C-1012					L											L					R				
C-1014				L			R															L			R
C-1017											L		R												

*In 11 animals 32 dorsal roots were sectioned individually on different sides and at different levels. Some of the observatio on these animals were reported in Part I of this study. Letters R and L refer to right and left sides and indicate roots section Asterisks indicate that sections of the medulla were cut in horizontal planes.

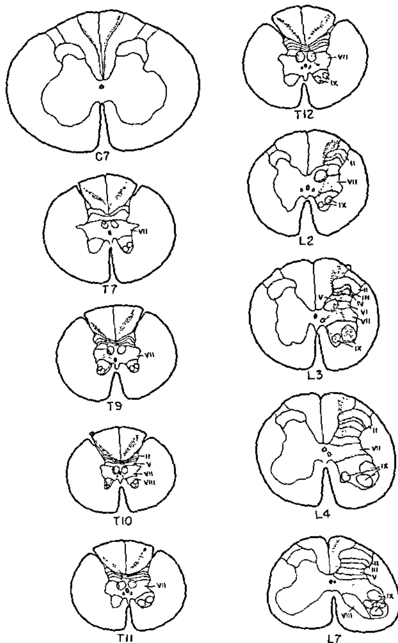


FIG 1 Drawings of the central distribution of degenerated fibers at spinal levels following section of T10 (left) and L3 (right) dorsal roots. Projected spinal cord sections of rhesus C-916 (T10) and C-914 (L3) were used in preparing these drawings.

Ascending degeneration, resulting from trauma, was present in both posterior spinocerebellar tracts. At the level of rhizotomy a moderate number of degenerated fibers from the root entry zone occupied a band along the lateral aspect of the posterior column. From this region degenerated fibers coursed obliquely through and around the medial portion of lamina II. Extensive degeneration was seen throughout the spinal gray except in lamina II. Because of these complicating features no description of degeneration within the spinal gray will be given.

In the posterior column, where no trauma was evident, descending degeneration from the T9 dorsal rhizotomy formed a narrow band in the lateral third of the posterior column, separated from the posterior horn by a narrow band of normal fibers. This degeneration was traced caudally as far as the L1 level. Ascending degeneration, localized to a diagonal band in the lateral portion of the fasciculus gracilis, shifted progressively medial at rostral spinal levels. In upper thoracic and cervical segments the most peripheral portion of this degenerated band occupied the dorsomedial border of the fasciculus cuneatus. As this band of degeneration extended ventromedially it crossed the posterior intermediate septum and lay along the lateral border of the fasciculus gracilis; the most ventral fibers were adjacent to the posterior median septum. A smaller proportion of the ascending degeneration was within the fasciculus cuneatus than following T8 dorsal rhizotomy (C-970, C-999 and C-1017).

Sections of the medulla were cut transversely. Degeneration from the T9 dorsal rhizotomy was located primarily within the gracile nucleus, but a few fibers projected to the medial parts of the cuneate nucleus. Extensive degeneration which appeared to originate from the posterior spinocerebellar tract was observed to project into cell group x (Brodal and Pompeiano, '57; Pompeiano and Brodal, '57).

Degeneration within the fasciculus gracilis was located laterally in sections through caudal portions of the nucleus gracilis. This portion of the nucleus remained relatively free of preterminal degeneration except for a small area ventrolaterally. In mid-portions of the nu-
cleus degeneration was located in a ban
along the dorsolateral border of the nu-
cleus. In ventral portions of the nucle
this band lay medial to, and somewha
removed from, the lateral border of th
nucleus. In the rostral portions of th
nucleus gracilis degeneration diminishe
in amount, and although located predom
nantly in the dorsolateral portions of th
nucleus, was not restricted to this are
Small amounts of degeneration were sca
tered throughout the central portion of th
nucleus.

The pattern of degeneration seen in th
nucleus gracilis was similar to that see
after T8 dorsal rhizotomy except that
was slightly more profuse and extende
into somewhat more ventromedial portio
of the nucleus.

Tenth thoracic root. In rhesus C-9
section of the tenth thoracic dorsal ro
produced a relatively narrow band of d
generated fibers along the entire later
border of the fasciculus gracilis adjace
to the posterior horn (fig. 1). Part of the
fibers projected to medial parts of lamin
III and IV by coursing medially throu
parts of lamina II. Other fibers from th
fasciculus gracilis passed directly
ventrolateral parts of the dorsal nucle
Relatively few degenerated fibers we
seen in lamina V or in portions of lami
VII removed from the dorsal nucleus.
few fibers were given off to the intermed
medial nucleus, but the major portion
the degeneration from lamina VII p
jected ventrally into medial parts of la
ina VIII. Fibers coursing through t
medial part of lamina VIII followed t
curve of the anterior horn and terminat
primarily in the ventromedial cell group
lamina IX. Relatively sparse degenerati
in central parts of laminae VII and V
projected towards the lateromedial c
group of lamina IX.

In adjacent thoracic segments, T11 a
T9, oblique narrow bands of degenerati
in the fasciculus gracilis were remov
slightly from the medial surface of t
posterior horn. The only degeneration
the spinal gray at T11 was in the later
part of the dorsal nucleus. In the T9 spin
segment a moderate reduction in t
amount of degeneration was observed

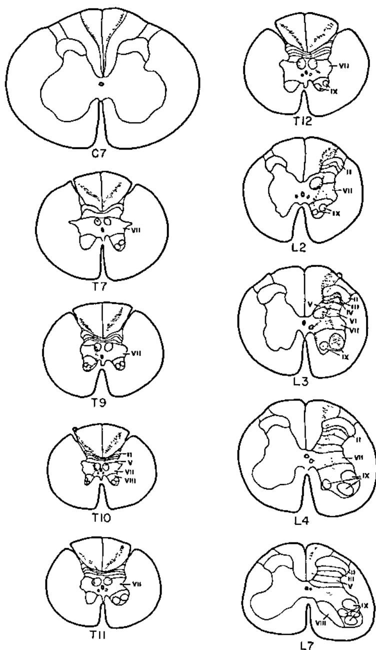


Fig. 1 Drawings of the central distribution of degenerated fibers at spinal levels following section of T10 (left) and L3 (right) dorsal roots. Projected spinal cord sections of rhesus C-916 (T10) and C-914 (L3) were used in preparing these drawings.

the spinal gray. This degeneration was found throughout laminae III and IV, in the ventral part of the dorsal nucleus and in adjacent central parts of lamina VII. Only sparse degeneration was present in medial parts of lamina VIII and virtually no degeneration was seen in lamina IX. (fig. 1).

Descending root fibers of T10 passed caudally in lateral parts of the fasciculus gracilis as far as the L1 spinal segment. At T12 and L1 spinal segments fibers entering the spinal gray projected exclusively into lateral parts of the dorsal nucleus.

Ascending T10 root fibers shifted to more medial portions of the fasciculus gracilis at higher levels, and where a well defined fasciculus cuneatus was present, they formed a narrow band medial, parallel and close to the posterior intermediate septum (fig. 14). These fibers ascended in this position to medullary levels. Collateral fibers were given off exclusively to dorsomedial parts of the dorsal nucleus in T8 and T7 spinal segments.

In transverse sections of the medulla ascending fibers from T10 were oriented along the lateral border of the nucleus gracilis with fewer fibers in the most dorsal and the most ventral regions (fig. 2). Very few of these fibers appeared to terminate about cells in ventrocaudal parts of the nucleus. In mid- and rostral portions of the nucleus most of the degenerated fibers terminated about small clusters of cells along the ventrolateral border of the nucleus gracilis near its common border with the cuneate nucleus. A few fibers seemed to extend into the dorsal apex of the nucleus cuneatus, but these were less numerous than those seen with rhizotomies at T8 or T9. In rostral parts of the nucleus gracilis, near the area postrema, degeneration was limited to cells in the dorsolateral part of the nucleus. No degeneration was seen in other medullary relay nuclei.

Twelfth thoracic root. The twelfth thoracic dorsal root was sectioned in rhesus C-1012. At the level of rhizotomy a narrow band of degenerated fibers from the root entry zone entered the lateral portions of the fasciculus gracilis. From this region fibers passed both through and around the medial portion of lamina II to enter medial portions of laminae III and I (fig. 8). Modest terminal degeneration ar fibers of passage were located in the ce tral portion of lamina V. From this regi degenerated fibers projected to the centr portion of lamina VII where they we especially prominent adjacent to the dors nucleus. From this site fibers passed ir the dorsal nucleus, medial portions lamina VIII, and to cell groups in lami IX.

Degeneration within the dorsal nucle was located principally in the ventrolate parts of the nucleus (fig. 8). Small fascic of degeneration projected to the inter diomedial nucleus located ventromedia the dorsal nucleus. Fibers travers lamina VII projected ventrally into lam VIII which at the T12 level extends a broad band across the anterior horn. most concentrated bundle of fibers pas into the medial part of lamina VIII coursed along the medial and ventral r gin of the anterior horn distributing fi to both ventromedial and lateromedial groups in lamina IX. Smaller, somev diffuse, fascicles of fibers coursed ve laterally in central parts of lamina VI reach the lateromedial cell group in lar IX.

At the L1 level degeneration in posterior column consisted of a na band slightly dorsomedial to the post horn. Virtually no degeneration was ent in laminae III and IV, but mc degeneration was seen in ventrola parts of the dorsal nucleus, surroun portions of lamina VII and the dorsom part of lamina VIII. No descending de eration was present in the post column at L2, but a few fibers were within lateral parts of the dorsal nuc No degeneration was detectable in caudal spinal segments.

At the T11 level, ascending degener in the posterior column had shifted r ally and was separated from the post horn by a band of normal fibers. De erated fibers could be seen enterinp spinal gray through medial portior laminae III and IV; these fibers were tributed to medial parts of these lam except for a lateral island of degenera The amount of degeneration was co erably less than at T12. Fibers proje

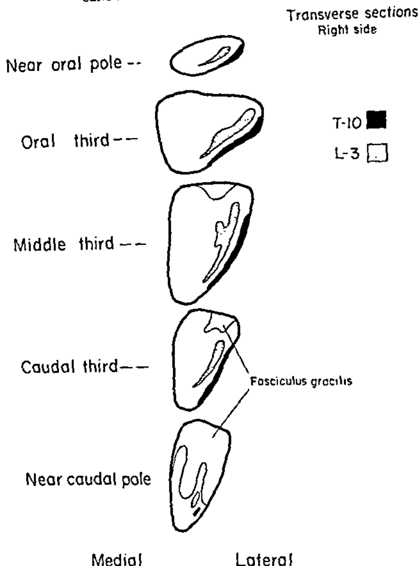


Fig. 2 Diagram of the principal zones of termination in the nucleus gracilis of ascending dorsal root fibers from T10 and L3. Because of the extensive overlap of fibers from certain dorsal roots, four separate drawings (also figs. 4, 5, 6) have been made of the terminal zones in the nucleus gracilis.

'entrally passed mainly to ventrolateral arts of the dorsal nucleus. Rostral to this evel the principal degeneration in the spinal gray was in the dorsal nucleus, but minimal, scattered degeneration was evident in laminae III and IV, as high as T10. Degenerated T12 dorsal root fibers projected to parts of the dorsal nucleus as far rostrally as T4. In spinal segments L2 to T10 degeneration in the dorsal nucleus was greatest in ventrolateral regions; from T9 to T4 it was greatest in dorsal parts of the nucleus.

Ascending degeneration in the posterior column shifted medially at higher levels. In upper thoracic spinal segments where the fasciculi cuneatus and gracilis were clearly separated, fibers from the T12 dorsal root formed a narrow band along the lateral border of the fasciculus gracilis;

the spinal gray. This degeneration was found throughout laminae III and IV, in the ventral part of the dorsal nucleus and in adjacent central parts of lamina VII. Only sparse degeneration was present in medial parts of lamina VIII and virtually no degeneration was seen in lamina IX. (fig. 1).

Descending root fibers of T10 passed caudally in lateral parts of the fasciculus gracilis as far as the L1 spinal segment. At T12 and L1 spinal segments fibers entering the spinal gray projected exclusively into lateral parts of the dorsal nucleus.

Ascending T10 root fibers shifted to more medial portions of the fasciculus gracilis at higher levels, and where a well defined fasciculus cuneatus was present, they formed a narrow band medial, parallel and close to the posterior intermediate septum (fig. 14). These fibers ascended in this position to medullary levels. Collateral fibers were given off exclusively to dorsomedial parts of the dorsal nucleus in T8 and T7 spinal segments.

In transverse sections of the medulla ascending fibers from T10 were oriented along the lateral border of the nucleus gracilis with fewer fibers in the most dorsal and the most ventral regions (fig. 2). Very few of these fibers appeared to terminate about cells in ventrocaudal parts of the nucleus. In mid- and rostral portions of the nucleus most of the degenerated fibers terminated about small clusters of cells along the ventrolateral border of the nucleus gracilis near its common border with the cuneate nucleus. A few fibers seemed to extend into the dorsal apex of the nucleus cuneatus, but these were less numerous than those seen with rhizotomies at T8 or T9. In rostral parts of the nucleus gracilis, near the area postrema, degeneration was limited to cells in the dorsolateral part of the nucleus. No degeneration was seen in other medullary relay nuclei.

Twelfth thoracic root. The twelfth thoracic dorsal root was sectioned in rhesus C-1012. At the level of rhizotomy a narrow band of degenerated fibers from the root entry zone entered the lateral portions of the fasciculus gracilis. From this region fibers passed both through and around the medial portion of lamina II to enter medial portions of laminae III and IV (fig. 8). Modest terminal degeneration and fibers of passage were located in the central portion of lamina V. From this region degenerated fibers projected to the central portion of lamina VII where they were especially prominent adjacent to the dorsal nucleus. From this site fibers passed into the dorsal nucleus, medial portions of lamina VIII, and to cell groups in lamina IX.

Degeneration within the dorsal nucleus was located principally in the ventrolateral parts of the nucleus (fig. 8). Small fascicles of degeneration projected to the intermediomedial nucleus located ventromedial to the dorsal nucleus. Fibers traversing lamina VII projected ventrally into lamina VIII which at the T12 level extends as a broad band across the anterior horn. The most concentrated bundle of fibers passed into the medial part of lamina VIII and coursed along the medial and ventral margin of the anterior horn distributing fibers to both ventromedial and lateromedial cell groups in lamina IX. Smaller, somewhat diffuse, fascicles of fibers coursed ventrolaterally in central parts of lamina VIII to reach the lateromedial cell group in lamina IX.

At the L1 level degeneration in the posterior column consisted of a narrow band slightly dorsomedial to the posterior horn. Virtually no degeneration was present in laminae III and IV, but modest degeneration was seen in ventrolateral parts of the dorsal nucleus, surrounding portions of lamina VII and the dorsomedial part of lamina VIII. No descending degeneration was present in the posterior column at L2, but a few fibers were seen within lateral parts of the dorsal nucleus. No degeneration was detectable in more caudal spinal segments.

At the T11 level, ascending degeneration in the posterior column had shifted medially and was separated from the posterior horn by a band of normal fibers. Degenerated fibers could be seen entering the spinal gray through medial portions of laminae III and IV; these fibers were distributed to medial parts of these laminae except for a lateral island of degeneration. The amount of degeneration was considerably less than at T12. Fibers projecting

Second lumbar root. The second lum-r dorsal root was sectioned in rhesus C-3. At the level of rhizotomy degeneration ɔm the root entry zone passed medially to the lateral part of the fasciculus graci-; dorsomedial to the posterior horn. From ıis region degenerated fibers coursed ırough and around the medial portion of ımina II and terminated profusely about ɜlls in the medial portions of laminae III nd IV. A modest number of fibers of pas-ıge was observed in the central portion of ımina V. These fibers projected into the entrolateral portion of the dorsal nucleus nd adjacent portions of lamina VII. De-enerated fibers reached the anterior horn y two routes. The major degeneration oursed medially through lamina VII, giv-ng off a few fascicles to the intermedio-nedial nucleus and curving ventrally hrough the medial portion of lamina VIII o terminate sparsely about both ventro-medial and centromedial cell groups of ıamina IX. A smaller amount of degenera-tion reached lamina IX through the central portions of laminae VII and VIII.

At the L3 spinal segment markedly reduced degeneration was confined to the medial portions of laminae III and IV. No dorsal nucleus was present in our sections of this level. No descending degeneration was observed in the fasciculus gracilis at L3 or in the spinal gray caudal to L3, presumably because of oversuppression of differential staining. Descending degeneration in the spinal gray at the L4 spinal level was meager and predominantly confined to parts of laminae III, IV and VII; very sparse degeneration occupied lamina IX.

At the L1 spinal segment modestly reduced degeneration was located predominantly in the medial portions of laminae III and IV and in ventrolateral portions of the dorsal nucleus. Only sparse, scattered degeneration was present in laminae VII and VIII.

Degeneration ascending in the fasciculus gracilis shifted progressively medial throughout lower thoracic segments. At levels with a well defined fasciculus cuneatus, degeneration occupied a spindle-shaped area in the lateral third of the fasciculus gracilis slightly medial to the region occupied by degeneration from the L1 dorsal root. Degeneration in the spinal gray rostral to the L1 segment extended to T7 and was confined to the dorsal nucleus. At the T12 and T11 segments degeneration in the dorsal nucleus was ventrolateral, but it shifted to lateral regions of the nucleus at T10. At the T7 level only scattered sparse degeneration was present in the nucleus.

In horizontal sections of the medulla degeneration was confined to the nucleus gracilis. Fibers from the fasciculus gracilis formed a small band of degeneration in the caudocentral portion of the nucleus gracilis. This band of degenerated fibers extended rostrally in dorsolateral parts of the nucleus for most of its length. A secondary smaller group of fibers, branching off from the main band, passed more medially in the middle third of the nucleus. Degenerated fibers of passage and occasional terminal fibers occupied the central portions of the nucleus gracilis between these lateral and medial bands. Only scant degeneration was scattered in the most ventral portions of the nucleus (fig. 6). No degeneration was seen in other medullary relay nuclei.

Third lumbar root. In rhesus C-914 the third lumbar dorsal root was sectioned. At the level of the rhizotomy degenerated fibers entered the lateral part of the fasciculus gracilis and were concentrated in an area dorsal to the posterior horn (figs. 1, 16). Root fibers traversing and coursing around medial parts of lamina II arborized about cells in all parts of laminae III and IV. Relatively sparse degeneration was seen in laminae V and VI centrally. Degeneration was scattered throughout lamina VII but was maximal in the central and medial regions adjacent to the dorsal nucleus. Within the dorsal nucleus degeneration was located principally in lateral portions. A small fascicle of fibers from lamina VII ended in the intermediomedial nucleus, while a concentrated bundle of fibers curved ventrally through the medial portion of lamina VIII to terminate in the ventromedial cell group of lamina IX. More diffuse groups of degenerated fibers traversed central portions of laminae VII and VIII to terminate predominantly about the centromedial cell group of lamina IX.

At the L4 spinal segment a marked

dorsolaterally a few of these fibers seemed to be present in the medial part of the fasciculus cuneatus. At cervical levels ascending degeneration was confined to the lateral margin of the fasciculus gracilis.

In transverse medullary sections modest amounts of degeneration formed a narrow band paralleling the lateral border of the nucleus gracilis. In caudal portions of the nucleus gracilis the major degeneration lay within the fasciculus gracilis; only a few terminal fibers were seen about the ventrolateral cells of the nucleus. The most profuse terminal degeneration was observed in mid-portions of the nucleus gracilis, where fibers formed an oblique band just inside the lateral border of the nucleus (fig. 6). At more rostral levels through the nucleus gracilis, terminal degeneration diminished gradually especially in dorsolateral and ventromedial regions, although the basic pattern of the degeneration and its orientation persisted into the most oral parts of the nucleus. No degeneration was observed in other medullary relay nuclei.

Lumbar dorsal roots

All seven lumbar dorsal roots were sectioned individually in seven animals. The L4 dorsal root was sectioned in two animals.

First lumbar root. The first lumbar dorsal root was sectioned in rhesus C-912. At the level of root section, degeneration from the root entry zone was distributed along nearly the entire lateral part of the fasciculus gracilis. Fascicles of degenerated fibers traversed medial portions of lamina II and terminated in medial and central parts of laminae III and IV. Other fascicles of degenerated fibers coursed through medial parts of lamina V to enter medial lamina VII (fig. 25). Part of these fibers passed into ventrolateral portions of the dorsal nucleus where they arborized about individual cells; a smaller group of fibers, projecting ventrally, terminated in the intermediomedial nucleus. The principal ventrally projecting fibers entered medial parts of lamina VIII and terminated about ventromedial cells of lamina IX. A few of these fibers followed the peripheral border of the anterior horn and entered the lateromedial cell group of lamina IX. A small, more loosely organized, group of fibers passed ventrolaterally through central parts of laminae VII and VIII towards the lateromedial cell group of IX.

Relatively modest descending degeneration from the L1 dorsal root occupied lateral portions of the fasciculus gracilis medial to normal fibers from the root [illegible] zone. The distribution of [illegible] degeneration in the L2 spinal gray was similar to that described at L1, except the number of fibers was considerably reduced and degeneration was scattered unevenly throughout the dorsal nucleus. Evaluation of descending L1 degeneration at more caudal spinal levels was complicated by extraneous degeneration in the L3 dorsal root zone.

Ascending L1 dorsal root degeneration occupied progressively more medial parts of the fasciculus gracilis in thoracic spinal segments. At upper thoracic and all cervical spinal levels these fibers formed a narrow band along the lateral border of the fasciculus gracilis. Degeneration in the spinal gray at T12 was similar to that at the rhizotomy level except for more modest degeneration in the anterior horn. In T10, T9 and T8 spinal segments degeneration in the spinal gray was confined to the dorsal nucleus; these fibers were distributed diffusely throughout the nucleus.

In transverse sections of the medulla degenerated dorsal root fibers formed a narrow band in the lateral portion of the nucleus gracilis throughout its rostrocaudal extent. Caudally, degeneration was maximal dorsally in the lateral part of the nucleus, excluding only the most lateral border. In middle portions of the nucleus degeneration was most abundant in the dorsal two-thirds of this lateral portion, again excluding the lateral border. However, some fibers extended ventrally along the lateral nuclear border. Rostrally terminal degeneration occupied the entire dorsoventral extent of the lateral portion of the nucleus. In sections near the oral pole of the nucleus only modest degeneration was seen near the lateral border (fig. 4). A few fibers from the main band of degeneration appeared to pass laterally and to reach the most medial border of the cuneate nucleus.

ıe posterior median septum close to the ›sterior gray commissure. Degeneration ı the spinal gray was greatly reduced at ıe L5 level; a few fibers arched ventrally › enter medial parts of the posterior horn ut little degeneration was seen in laminae II and IV. These fibers, entering through nedial parts of lamina V, mainly termi-.ated about clusters of cells in central amina VI. Fibers traversing the above aminae projected ventrally in central ›arts of lamina VII; only a few fibers eached cell groups of lamina IX. At L6, L7 .nd S1 levels the principal degeneration vas present in central and medial parts of aminae V and VI; fibers projected to these ocations directly from the posterior col-ımn. Scattered degeneration was seen in .amina IV at L6, and in dorsal parts of ıamina VII, at L6, L7 and S1; no degen-erated fibers were seen in lamina IX in these spinal segments. More caudal spinal segments were not available in rhesus C-1011.

Immediately rostral to the rhizotomy (i.e., L3) degenerated fibers were seen throughout lamina I and profusely in all parts of laminae III and IV. Similar profuse degeneration passed through lamina V into central parts of lamina VI. From this locus in lamina VI fibers pursued two courses: (1) ventromedially, through medial parts of laminae VII and VIII to lamina IX, and (2) ventrolaterally, through central parts of laminae VII and VIII to terminate about centromedial cell groups of lamina IX. A few fibers terminated about large cells of the dorsal nucleus in medial lamina VII. At the L2 level, moderate degeneration was present in laminae III and IV, but the principal degeneration was distributed to all parts of the dorsal nucleus (figs. 19, 20). In successively more rostral spinal segments, the degeneration in the spinal gray was seen only in the ipsilateral dorsal nucleus. Degeneration in the dorsal nucleus extended as high as T8. At L1 and T12 degeneration in this nucleus was mainly in lateral regions, but at higher levels it shifted to dorsal portions of the nucleus.

Ascending fibers in the fasciculus gracilis formed a relatively broad band in upper lumbar spinal segments which was displaced progressively medially by entering normal fibers. In lower thoracic segments the band appeared narrower in the lateral half of the fasciculus but did not extent ventromedially to the spinal gray. In upper thoracic and cervical spinal segments ascending degeneration in the lateral part of the fasciculus gracilis was less concentrated and did not form as discrete a band as at lower levels.

In transverse medullary sections caudal cell groups of the nucleus gracilis were interlaced with degenerated fibers concentrated in the lateral third of the fasciculus gracilis, but removed from the lateral border. Only sparse degeneration was seen about central cell groups at this level. In middle portions of the nucleus degeneration, prominent in central and lateral parts of the nucleus, extended dorsally (fig. 4). Less degeneration was evident in ventromedial regions. Rostrally the principal terminal degeneration was seen in an irregularly shaped area in dorsolateral portions of the nucleus gracilis, but scattered islands of degeneration were present in central regions (fig. 32). Degeneration, seen in the oral pole of the nucleus, was scattered in central and lateral regions.

Fifth lumbar root. The fifth lumbar dorsal root was sectioned in rhesus C-1012. At the level of dorsal rhizotomy degenerated fibers occupied the root entry zone and lateral portions of the fasciculus gracilis dorsal to the posterior horn. Degenerated fibers coursed through and around the medial portion of lamina II to terminate in the central and medial parts of laminae III and IV. Ventrally coursing fascicles of fibers traversed central and medial parts of lamina V. Part of the fibers coursing from this lamina arborized about cells in the medial part of lamina VI, while others entered lamina VII. The majority of fibers from this area coursed ventrally through central portions of lamina VII to terminate in the ventral and ventrolateral cell groups of lamina IX. A small bundle of fibers coursing through medial parts of lamina VII projected to the intermediomedial nucleus and into medial parts of lamina VIII. A few of these fibers entered medial cell groups of lamina IX.

At the L6 level the band of degeneration in the fasciculus gracilis had moved ventromedially to occupy the central por-

reduction in the amount of degeneration occurred, so that only sparse and scattered degeneration was seen in parts of laminae III, IV, V, VI and VII. More degeneration was seen in medial parts of laminae III and IV at L2 and fibers passing through lamina V projected to ventrolateral parts of the dorsal nucleus and adjacent areas of lamina VII. Only a few scattered fibers were seen in laminae VIII and IX (fig. 1).

Descending branches of the L3 dorsal root shifted to more medial and dorsal parts of the fasciculus gracilis caudal to the rhizotomy. These fibers were traced as far as L7 in the fasciculus gracilis. A modest number of fibers were given off in lower lumbar segments primarily to medial parts of laminae VI and VII (figs. 24, 26).

Ascending degeneration from the L3 dorsal rhizotomy occupied an oblique band in the lateral half of the fasciculus gracilis that migrated medially at successively rostral levels. Fibers were given off to the dorsal nucleus and adjacent areas of lamina VII from L2 to T10 spinal segments. The location of degeneration within the dorsal nucleus changed as follows: (1) at L2 and L1 it was ventrolateral, (2) at T12 it was dorsolateral, and (3) at T10 it was dorsomedial. In upper thoracic and cervical spinal segments ascending degeneration formed a narrow band just medial to the posterior intermediate septum. More degenerated fibers were seen dorsally than ventrally, but none of the degenerated fibers reached the dorsal margin of the spinal cord.

In transverse sections of the lower medulla L3 root fibers continued to occupy a small dorsolateral part in the fasciculus gracilis; few fibers terminated in caudal parts of the nucleus gracilis. In the middle third of the nucleus degenerated fibers were most concentrated laterally near fibers of the fasciculus gracilis but scattered degenerated fibers began to appear in more medial parts of the nucleus. In the rostral third of the nucleus gracilis terminal degeneration was seen dorsally just inside the lateral border. Although the area containing terminal degeneration varied slightly from section to section, maximal degeneration persisted to the oral pole in a slightly curved band medial to, and separated from, the lateral border of the nucleus (fig. 2). At these levels all other parts of the nucleus were free of degeneration.

Fourth lumbar root. The fourth lumbar dorsal root was sectioned in two animals (C-915 and C1011) but spinal trauma in rhesus C-915 caudal to the rhizotomy resulted in extraneous spinal degeneration. Only the reliable observations in rhesus C-1011 will be reported.

At the level of rhizotomy degenerated fibers entered the dorsolateral part of the fasciculus gracilis dorsal to the posterior horn. From this region a large number of root fibers passed through medial portions of laminae I and II to enter central and medial parts of laminae III and IV. Modest terminal degeneration was seen about cells in central and medial parts of lamina I, but no similar degeneration was seen in underlying parts of lamina II. Although terminal degeneration was seen in central and medial parts of laminae III and IV, the most medial portions of these laminae were free of degenerated fibers (fig. 9). A small number of fibers entered ventrolateral parts of lamina II from the zone of Lissauer. These fibers terminated in lateral parts of laminae II through IV. A relatively clear zone was present in the lateral half of laminae III and IV which did not receive fibers from fascicles entering either medially or laterally. Ventrally projecting fibers passed through central parts of laminae V and VI into dorsal parts of lamina VII. A number of these fibers terminated in central parts of lamina VI. From the large collection of degenerated fibers in lamina VII, a small, distinct bundle projected ventromedially to the intermediomedial nucleus, and continued ventrally into the dorsomedial part of lamina VIII (fig. 23). None of these fibers could be traced into cell groups of lamina IX. Fibers projecting to cell groups of lamina IX coursed through central parts of lamina VII in numerous small fascicles.

Descending branches of dorsal root fibers shifted to central regions of the fasciculus gracilis caudal to the rhizotomy; these fibers appeared diffuse and scattered at L5, but at the L6 level formed a relatively thin band in the medial part of the fasciculus. At the S1 level a small number of descending fibers were localized near

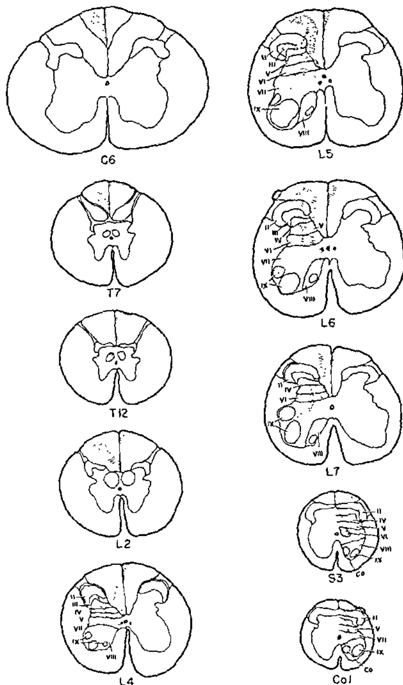


Fig. 3 Drawings of the central distribution of degenerated fibers at spinal levels following section of L6 (left) and Co1 (right) dorsal roots in rhesus C-1014.

tion of the fasciculus gracilis, leaving a band of normal fibers adjacent to the posterior horn. Root fibers entered the spinal gray at L6 chiefly through the medial portion of lamina V and were distributed primarily to medial parts of lamina VI. Smaller amounts of degeneration were scattered in central regions of lamina VII. No degeneration was observed in the intermediomedial nucleus at this level. Progressively diminishing amounts of descending degeneration were present in similar locations in the fasciculus gracilis and spinal gray as far caudally as S1.

At the L4 spinal segment the amount of degeneration in the spinal gray nearly equalled that observed at L5. This degeneration was distributed profusely throughout laminae III and IV, about the central cell clusters of lamina VI, and in central portions of lamina VII. Although a significant number of degenerated fibers traversed central portions of laminae VII and VIII, the number of degenerated fibers terminating in cell groups of lamina IX was less than at L5. A small amount of degeneration entered the intermediomedial nucleus while only sparse degeneration was present in the medial portion of lamina VIII. At the L3 spinal segment the greatest reduction in the amount of degeneration occurred within laminae III, IV and IX. Maximum degeneration persisted in the medial portion of lamina VI and adjacent portion of lamina VII. At the L2 spinal level and above, degeneration in the spinal gray was virtually limited to the dorsal nucleus; degeneration in this nucleus was seen rostrally as far as T7. At L2, L1 and T12 spinal segments profuse degeneration occupied virtually the entire cross-sectional area of the dorsal nucleus, but in more rostral segments degeneration was localized to dorsomedial portions of the nucleus.

Ascending L5 dorsal root fibers in the fasciculus gracilis occupied a broad band which progressively shifted medially. At the L1 level this band of degeneration occupied nearly the medial *half of the fasciculus gracilis* except for a dorsomedial wedge of normal fibers. At levels where a well defined fasciculus cuneatus was present, only the above-mentioned dorsomedial wedge and the lateral border of the fasciculus gracilis were devoid of ascen*ing degeneration.*

In transverse sections through the ca dal pole of the nucleus gracilis a few arb rizations were noted about cells in th ventromedial part of the nucleus. In th caudal third of the nucleus gracilis degen eration occupied an irregular central are which extended into ventromedial regions in middle portions of the nucleus degenera tion was most profuse centrally and ven trally (figs. 5, 33). Only extreme dorso medial and lateral portions of the nucleu were free of degeneration. At rostral level the major area of degeneration shifte dorsomedially. In the oral pole of th nucleus degeneration was dorsomedial. The impressive feature of terminal degen eration from the L5 dorsal rhizotomy within the nucleus gracilis was its diffuse distribution within the central core of the nucleus.

Sixth lumbar root. The sixth lumbar dorsal root was sectioned in two animals (C-916 and C-1014). Observations in these animals were remarkably similar. At the level of the dorsal rhizotomy degenerated root fibers were most numerous in the lateral part of the fasciculus gracilis immediately dorsal to the posterior horn (fig. 3) Fibers from this area coursed through medial parts of laminae I and II, as well as around the medial border of these laminae. to enter laminae III and IV. A moderate number of fibers terminated in medial parts of lamina I, while no fibers appeared to end in lamina II. In laminae III and IV terminal degeneration was maximal in medial and central regions. From lamina IV fibers projected ventrally through central parts of laminae V and VI to medial parts of lamina VII. Terminal arborizations were seen about clusters of cells in medial parts of lamina VI. A small, distinct bundle of fibers projected medially to the intermediomedial nucleus. Fibers bypassing this nucleus projected ventrally *in the* most medial part of lamina VIII; a few of these fibers ended upon cells of the centromedial cell group *of lamina IX*. A larger number of fibers radiated through central parts of lamina VII to enter large ventral and lateral cell groups of lamina IX.

At the L7 level descending degenerated fibers in the fasciculus gracilis had moved

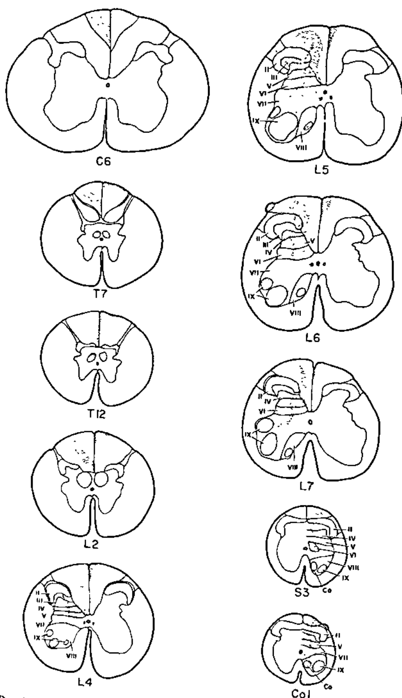


Fig 3 Drawings of the central distribution of degenerated fibers at spinal levels following section of L6 (left) and Co1 (right) dorsal roots in rhesus C-1014.

nearer the midline where they formed a narrow band extending from the dorsal border of the cord to the posterior gray commissure. Relatively sparse degeneration was present in medial parts of laminae III and IV. The most impressive degeneration was in medial and central parts of lamina VI; fibers projected to this location directly from the fasciculus gracilis by traversing medial parts of lamina V (fig. 27). A modest number of fibers, bypassing cells of lamina VI, radiated through central parts of lamina VII; part of these fibers terminated in the dendritic fields of cells in ventral and lateral regions of lamina IX (fig. 3).

At the S1 level descending degenerated fibers from the fasciculus gracilis swept ventrally through extreme medial parts of the posterior horn into medial parts of laminae V, VI and VII. In rhesus C-1014 a small number of these fibers entered lateral cell groups of lamina IX. In rhesus C-916 observations were similar, except that a distinct bundle of fibers, traversing medial parts of lamina VIII, projected to the oral pole of the commissural nucleus within this lamina. More caudal spinal sections were not available in this animal, but it seemed likely that descending degeneration extended further caudally.

At the L5 level ascending fibers from the L6 dorsal root shifted to central parts of the fasciculus gracilis where they formed a slightly curved broad band. Fewer fibers traversed laminae I and II and terminal degeneration in laminae III and IV was less than at L6. In rhesus C-916 degeneration was seen in lateral parts of laminae III and IV, while in rhesus C-1014 a larger number of fibers occupied only medial portions of these laminae. In both animals the most profuse degeneration was seen in medial parts of laminae V and VI; fibers entered these laminae directly from the fasciculus gracilis. Relatively modest degeneration passed through central parts of lamina VII to reach internal portions of lateral cell groups of lamina IX. At successively rostral spinal levels, progressively diminished degeneration within the spinal gray was seen in all locations except for medial portions of laminae V and VI which continued to receive direct fibers from the fasciculus gracilis as far rostrally as L3.

Persistent degeneration in medial parts of lamina VI was characterized by arborizations about clusters of large cells (fig. [illegible]). At the L3 level medial parts of lamina VI appeared to merge with the dorsal nucleus. Rostral to L3 degeneration within the spinal gray was confined to portions of the dorsal nucleus. Degeneration within caudal parts of the dorsal nucleus (L3-L2) was distributed throughout its entire cross-sectional area (fig. 22). Ascending fibers in the fasciculus gracilis projected to portions of the dorsal nucleus as far rostrally as T9 (C-916) and T7 (C-1014). In these lower thoracic segments degeneration within the dorsal nucleus was localized to small dorsomedial regions (fig. 21).

Ascending dorsal root fibers formed a relatively broad, obliquely oriented bundle in central parts of the fasciculus gracilis throughout the lumbar segments. In lower thoracic spinal segments ascending fibers from the L6 dorsal root shifted to more medial and dorsal parts of the fasciculus gracilis, but still left a small wedge of normal fibers in the most dorsomedial part of the fasciculus. This pattern persisted with minor variations throughout upper thoracic and cervical segments, but at these levels degenerated fibers seemed to occupy a smaller area (figs. 3, 18).

In caudal transverse medullary sections L6 dorsal root fibers were concentrated in medial parts of the fasciculus gracilis. When cell groups of the nucleus gracilis appeared, degenerated fibers shifted ventromedially and some fibers terminated about ventromedial cell clusters. As the nucleus gracilis enlarged in its middle third degenerated terminal fibers were distributed over a large area comprising nearly the medial two-thirds of the nucleus (figs. 4, 34, 36). Terminal degeneration was maximal in central and ventral regions in this part of the nucleus. In the rostral third of the nucleus profuse degeneration still occupied ventromedial parts, but it did not extend to either the medial or ventral borders of the nucleus. Scattered terminal degeneration was seen in central and ventral parts of the nucleus near its oral pole. Terminal degeneration in the nucleus gracilis was maximal in ventromedial regions of the middle and rostral parts of the nucleus. No L6 dorsal root fibers

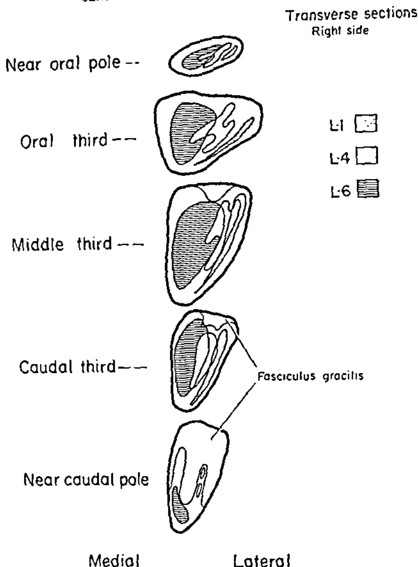


Fig 4 Diagram of the principal zones of termination in the nucleus gracilis of ascending dorsal root fibers from L1, L4 and L6.

passed to other medullary relay nuclei.

Seventh lumbar root. Section of the seventh lumbar dorsal root (C-914) produced degeneration, at the level of section, in the medial root entry zone and in the lateral two-thirds of the fasciculus gracilis dorsal and medial to the posterior horn (fig. 10). Fascicles of degeneration traversed and coursed around medial parts of lamina II in their projection to predominantly lateral cell groups of laminae III and IV. Other degenerated fascicles coursing around laminae II, III and IV entered medial parts of lamina V directly. Part of these fibers terminated in medial parts of laminae V and VI, while others, in passage, entered dorsocentral regions of lamina VII. The majority of degenerated fibers in this location coursed ventrolaterally through central parts of lamina VII and entered ventral and ventrolateral cell groups in lamina IX. A smaller bundle of fibers

passed ventromedially to the intermediomedial nucleus and medial portions of lamina VIII. A few of these fibers reached centromedial cell groups of lamina IX.

A marked reduction in the amount of degeneration in the spinal gray was evident in the L6 segment. The most impressive degeneration at this level was seen in medial parts of laminae V and VI; fibers in this region appeared to enter the gray directly from the posterior columns. Degeneration in laminae III and IV was modest in amount and scattered, except for a discrete island of terminal fibers in the lateral part of lamina III. Relatively scant degeneration was seen in laminae VII and IX. Similarly distributed degeneration was seen in the spinal gray at L5; the most profuse degeneration continued to be present in medial parts of laminae V and VI. At L4 degeneration within the spinal gray was seen only in medial portions of lamina VI. No sections of the spinal cord caudal to the L7 segment were available for study.

In their ascending course dorsal root fibers of L7 shifted rapidly to medial portions of the fasciculus gracilis, where they formed a relatively narrow band extending from the periphery of the cord to the posterior gray commissure. This band of fibers was widest dorsally and tapered ventrally where it was adjacent to the posterior median septum. None of these ascending fibers extended into the most dorsomedial part of the fasciculus gracilis; a clear triangular area in this location was devoid of degenerated fibers (figs. 15, 16). This pattern within the fasciculus gracilis persisted with slight variation until the L1 segment. At this level the degenerated fibers shifted dorsally, forming a dorsomedial diagonal band adjacent and lateral to a wedge-shaped area of normal fibers in the most dorsomedial part of the fasciculus gracilis. At T12 and higher levels of the spinal cord, these fibers formed a "boomerang-shaped" configuration with one arm parallel to the posterior median septum and the other arm following the curvature of the spinal cord. These degenerated fibers were separated from the posterior median septum and the dorsal margin of the cord by a narrow zone of normal fibers. This basic "boomerang" configuration of ascending degeneration in the fasciculus gracilis persisted with minor variation throughout thoracic and cervical spinal segments.

As previously described, ascending fibers from the fasciculus gracilis entered lamina V and projected to cells in medial parts of laminae V and VI throughout the lumbar enlargement. At the L3 level and above, similar fibers projecting to the dorsal nucleus constituted the only degenerated fibers from the L7 root distributed to the spinal gray. Root fibers passing to the dorsal nucleus terminated in different locations at various levels. At L3 and L1 relatively profuse degeneration was seen in lateral and ventrolateral portions of the nucleus while at T12 and T10 degenerated fibers occupied only dorsal portions of the nucleus.

In transverse medullary sections, L7 dorsal root fibers ascending in the fasciculus gracilis retained their basic "boomerang" configuration, although this pattern was not so sharp due to scattering of fibers. At caudal medullary levels degenerated fibers in the fasciculus gracilis were dorsal and medial to the principal cell groups of the nucleus gracilis. A number of terminal fibers were seen in medial parts of the nucleus caudally. At levels where most of the fibers of the fasciculus gracilis had disappeared, terminal degeneration was scattered throughout the medial half of the nucleus; more concentrated degeneration was seen in dorsomedial areas removed from the margins of the nucleus (fig. 5). Through rostral parts of the nucleus gracilis terminal degeneration was profuse and localized to dorsocentral areas. Surrounding regions of the nucleus were free of degeneration except for scattered terminations ventromedially. Near the oral pole of the nucleus gracilis the area containing terminal degeneration extended obliquely from the dorsocentral region to the most ventromedial area. At this level the lateral third of the nucleus and the extreme dorsomedial area were entirely devoid of degeneration.

First sacral root. In rhesus C-912 the first sacral dorsal root was sectioned. At the rhizotomy level degeneration occupied the lateral third of the fasciculus gracilis dorsal to the posterior horn. From the latter

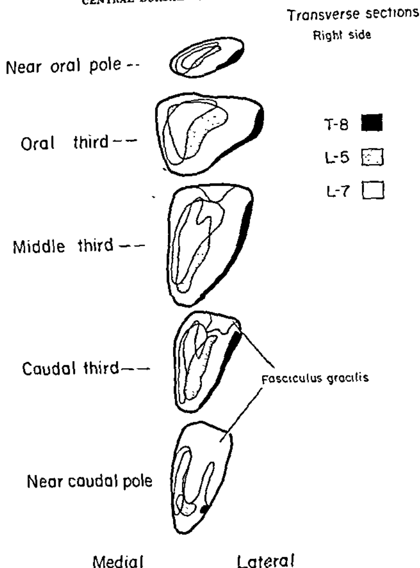


Fig 5 Diagram of the principal zones of termination in the nucleus gracilis of ascending dorsal root fibers from T8, L5 and L7. Fibers from dorsal roots of the lumbar enlargement (fig. 4) project to irregular shaped areas in the central core of the nucleus

region fascicles of degeneration traversed medial portions of lamina II to terminate primarily in medial and central portions of laminae III and IV (fig. 11). A smaller number of fibers projected into the medial and central portions of laminae V and VI, where some of these fibers appeared to terminate. A modest number of degenerated fibers coursed ventrally through medial portions of lamina VII, giving off fibers to the intermediomedial nucleus, and continuing ventrally in medial parts of lamina VIII. Fibers with this course entered mainly the ventromedial cell group of lamina IX. Smaller fascicles of degenerated fibers traversed central parts of lamina VII in passage toward ventral and lateral cell groups of lamina IX, but few of these fibers reached these cell groups.

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In their ascending course dorsal root fibers of L7 shifted rapidly to medial portions of the fasciculus gracilis, where they formed a relatively narrow band extending from the periphery of the cord to the posterior gray commissure. This band of fibers was widest dorsally and tapered ventrally where it was adjacent to the posterior median septum. None of these ascending fibers extended into the most dorsomedial part of the fasciculus gracilis; a clear triangular area in this location was devoid of degenerated fibers (figs. 15, 16). This pattern within the fasciculus gracilis persisted with slight variation until the L1 segment. At this level the degenerated fibers shifted dorsally, forming a dorsomedial diagonal band adjacent and lateral to a wedge-shaped area of normal fibers in the most dorsomedial part of the fasciculus gracilis. At T12 and higher levels of the spinal cord, these fibers formed a "boomerang-shaped" configuration with one arm parallel to the posterior median septum and the other arm following the curvature of the spinal cord. These degenerated fibers were separated from the posterior median septum and the dorsal margin of the cord by a narrow zone of normal fibers. This basic "boomerang" configuration of ascending degeneration in the fasciculus gracilis persisted with minor variations throughout thoracic and cervical spinal segments.

As previously described, ascending fibers from the fasciculus gracilis entered lamina V and projected to cells in medial parts of laminae V and VI throughout the lumbar enlargement. At the L3 level and above, similar fibers projecting to the dorsal nucleus constituted the only degenerated fibers from the L7 root distributed to the spinal gray. Root fibers passing to the dorsal nucleus terminated in different locations at various levels. At L3 and L1 relatively profuse degeneration was seen in lateral and ventrolateral portions of the nucleus while at T12 and T10 degenerated fibers occupied only dorsal portions of the nucleus.

In transverse medullary sections, L7 dorsal root fibers ascending in the fasciculus gracilis retained their basic "boomerang" configuration, although this pattern was not so sharp due to scattering of fibers. At caudal medullary levels degenerated fibers in the fasciculus gracilis were dorsal and medial to the principal cell groups of the nucleus gracilis. A number of terminal fibers were seen in medial parts of the nucleus caudally. At levels where most of the fibers of the fasciculus gracilis had disappeared, terminal degeneration was scattered throughout the medial half of the nucleus; more concentrated degeneration was seen in dorsomedial areas removed from the margins of the nucleus (fig. 5). Through rostral parts of the nucleus gracilis terminal degeneration was profuse and localized to dorsocentral areas. Surrounding regions of the nucleus were free of degeneration except for scattered terminations ventromedially. Near the oral pole of the nucleus gracilis the area containing terminal degeneration extended obliquely from the dorsocentral region to the most ventromedial area. At this level the lateral third of the nucleus and the extreme dorsomedial area were entirely devoid of degeneration.

First sacral root. In rhesus C-912 the first sacral dorsal root was sectioned. At the rhizotomy level degeneration occupied the lateral third of the fasciculus gracilis dorsal to the posterior horn. From the latter

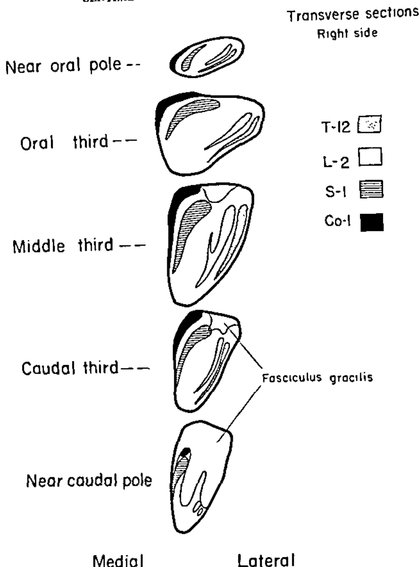


Fig. 6 *Diagram of the principal zones of termination in the nucleus gracilis of ascending dorsal* root fibers from T12, L2, S1 and Co1. Lower thoracic and upper lumbar roots project fibers to narrow oblique laminae lateral to the core regions (figs. 2, 4), while sacral and coccygeal dorsal roots project fibers to crescent-shaped laminae in dorsomedial parts of the nucleus.

ventrally in medial lamina V to be distributed in a triangular area in central lamina VII (fig. 12). The apex of this triangular area was directed dorsally. Cells in this part of lamina VII were large and intimately surrounded by degenerated fibers. Fibers *projecting ventrally from* lamina VII traversed medial parts of lamina VIII and arborized about cells of the commissural nucleus within this lamina (figs. 29, 30). Very few dorsal root fibers entered ventromedial cell groups of lamina IX, although small bundles of *fibers passed ventrolaterally through cen-*

At the S2 level descending degeneration still occupied the lateral third of the fasciculus gracilis, and degenerated fibers entered the posterior horn in the same manner as described at S1. Reduced amounts of degeneration in the spinal gray were distributed to medial and lateral parts of laminae III and IV, central portions of laminae V and VI, and dorsal parts of lamina VII. A small number of fibers from lamina VII reached cell groups in lamina IX. In the S3 spinal segment traces of descending degeneration were seen in the lateral part of the fasciculus gracilis but little degeneration was seen in the spinal gray.

Rostral to the dorsal rhizotomy (i.e., L7) reduced amounts of degeneration were seen in medial parts of laminae III and IV. Fibers entering medial parts of lamina V directly from the posterior column projected into medial lamina VI where some of these fibers terminated. Fibers entering dorsal parts of lamina VII projected to the intermediomedial nucleus and into medial parts of lamina VIII. Part of these fibers entered ventromedial cell groups of lamina IX.

No sections of spinal segments L6 through L4 were available for study. At the L3 level sparse degeneration was seen in medial parts of laminae III, IV and VII. In more rostral spinal segments degeneration in the spinal gray was confined to the dorsal nucleus. At L2 degeneration was localized to the dorsomedial part of the dorsal nucleus, while at L1 and T12 only sparse scattered degeneration was seen within this nucleus.

Ascending degeneration in the fasciculus gracilis at the L7 level formed a narrow, oblique band, contiguous ventrally with the posterior median septum, but separated from it dorsally. The largest amount of degeneration, located dorsally, extended laterally for a short distance along the dorsal border of the spinal cord. A small dorsomedial wedge of normal fibers separated degenerated fibers from the dorsal part of the posterior median septum. At upper lumbar levels the degenerated fibers shifted dorsally and laterally, forming a narrow, oblique band adjacent to a small dorsomedial wedge of normal fibers (fig. 7). As this band of fibers ascended maximal degeneration appeared to be dorsolateral. From the T9 level rostrally ascending degeneration formed a "boomerang-shaped" configuration, similar to that described for L7, except that the arm parallel to the posterior median septum was shorter and the lateral arm extended along nearly the entire dorsal border of the fasciculus gracilis. A small wedge of normal fibers always remained in the dorsomedial part of the fasciculus. In cervical regions a modest number of degenerated fibers appeared within the concavity of the "boomerang," giving a somewhat diffuse pattern to the ascending degeneration.

Ascending degeneration from the S1 dorsal root was studied in transverse sections of the medulla. Degenerated fibers occupied dorsocentral parts of the fasciculus gracilis in the lower medulla, where they were arranged in a less pronounced "boomerang" configuration. Near the middle third of the nucleus fibers passed ventromedially to terminate about cells in a "comma-shaped" area in the medial part of the nucleus gracilis. The most ventral projections at this level surrounded cells along the medial border of the nucleus. In more rostral parts of the nucleus several distinct and separate groups of fibers passed ventromedially in small bands that coursed between and around certain cell groups. Terminal degeneration was seen about cells in the medial two-thirds of the nucleus dorsally, but at most levels several groups of cells were completely free of degeneration (figs. 6, 35). A large oblique band on the lateral border of the nucleus was devoid of degeneration. Near the oral pole of the nucleus the amount of degeneration diminished, but that present persisted in a ventromedial location; at these levels the degeneration did not extend to either the medial or ventral borders of the nucleus.

First coccygeal root. The first coccygeal dorsal root was sectioned in rhesus C-1014. At the level of the rhizotomy dorsal root fibers were scattered throughout the fasciculus gracilis, but with the greatest number of fibers in lateral regions (fig. 31). These fibers entered the spinal gray medially at a considerable distance from the apex of the posterior horn, and terminated about cells in medial portions of laminae III and IV. Degenerated fibers projected

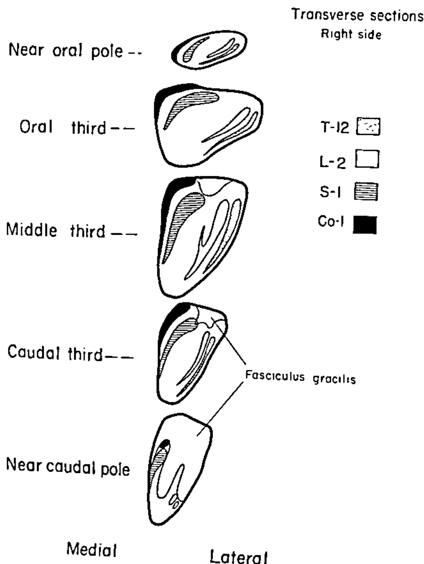


Fig. 6 Diagram of the principal zones of termination in the nucleus gracilis of ascending dorsal root fibers from T12, L2, S1 and Co1. Lower thoracic and upper lumbar roots project fibers to narrow oblique laminae lateral to the core regions (figs. 2, 4), while sacral and coccygeal dorsal roots project fibers to crescent-shaped laminae in dorsomedial parts of the nucleus

ventrally in medial lamina V to be distributed in a triangular area in central lamina VII (fig. 12). The apex of this triangular area was directed dorsally. Cells in this part of lamina VII were large and intimately surrounded by degenerated fibers. Fibers projecting ventrally from lamina VII traversed medial parts of lamina VIII and arborized about cells of the commissural nucleus within this lamina (figs. 29, 30). Very few dorsal root fibers entered ventromedial cell groups of lamina IX, although small bundles of fibers passed ventrolaterally through cen-

tral parts of lamina VIII in the direction of this cell group. No spinal sections caudal to Co1 were available for study.

At the S3 level diffuse degeneration in the fasciculus gracilis was maximal in medial and dorsal regions. A small number of fibers from the fasciculus gracilis entered medial parts of the posterior horn to terminate in medial portions of laminae III and IV, as well as dorsomedial parts of lamina VII. Little degeneration was present in lamina VIII and none was seen in lamina IX. At the S2 level degeneration was similar in distribution but reduced in amount. No degeneration was seen in the spinal gray at S1 or in more rostral spinal sections.

Ascending degeneration in the fascicu-

confined to a small dorsomedial region, at and above the S1 level. These ascending fibers formed a narrow triangular area, with the long side against the posterior median septum, throughout most of the lumbar segments. At the second lumbar segment the bundle of degenerated fibers in the fasciculus gracilis shifted dorsally to occupy a less perfectly shaped triangle in which the long side conformed to the dorsal margin of the spinal cord. At midthoracic levels ascending fibers from Co1 formed a narrow band extending over nearly the entire dorsal surface of the fasciculus gracilis (figs. 3, 18); this pattern of fiber disposition persisted into lower cervical segments. In cervical segments C6 through C3, degenerated Co1 dorsal root fibers formed an inverted "L"-shaped pattern in the dorsomedial part of the fasciculus gracilis with the base of the "L" elongated. In the highest cervical segments these ascending fibers formed a thin band along the dorsal margin of the fasciculus gracilis.

Medullary degeneration resulting from section of the Co1 dorsal root was confined to dorsomedial parts of the nucleus gracilis. As ascending fibers entered the nucleus gracilis they were distributed to medial regions and along the dorsal border of the nucleus, in a "boomerang" configuration (fig. 36). Degeneration in this location was seen throughout the nucleus gracilis but was greatest in its middle third (fig. 6).

DISCUSSION

Part II of this study provides data for rhesus monkey concerning the ... distribution and termination of lo... thoracic, lumbosacral and coccygeal do... root fibers. Observations reported here c... cern the distribution of dorsal root fibe... the spinal gray, ascending and descend... dorsal root fibers in the posterior funic... and the pattern of termination of ... ing fibers in the nucleus gracilis. T... data together with those from Part I of ... study permit certain comparisons rega... ing ascending fibers passing to medull... relay nuclei.

Spinal projections

Fiber projections to the spinal gr... The spinal distribution of lower thorac... lumbosacral and coccygeal dorsal ro... in the monkey are similar to those ... the cervical and upper thoracic dor... roots, except for certain local difference... In lower thoracic spinal segments a small... number of dorsal root fibers mainly ent... more medial regions of the posterior hor... This appears to be due to the fact th... laminae II through V extend across ... midline and fuse with corresponding lam... nae of the opposite side. The number ... degenerated fibers in the zone of Lissau... appears relatively greater and more even... distributed in lower thoracic segmen... probably because this zone is so narrow. ... these spinal segments dorsal root fibers ... very nearly confined to their segment... entry except for fibers passing to the dor... nucleus. This may be due to the long... rostrocaudal extent of thoracic spinal seg... ments (Thomas and Combs, '65).

Observations concerning the upper thre... lumbar dorsal roots are similar to those ... the lower thoracic segments except tha... the number of root fibers is greater an... moderate numbers of fibers are distribute... to portions of the spinal gray in adjacen... spinal segments in locations other than th... dorsal nucleus.

Lower lumbar spinal segments (i.e., L... to L7) are characterized by larger dors... roots, more extensive spinal gray, enlarge... and broader laminae in the posterior hor... that are not fused across the midline, an... well developed cell groups in lamina I... especially in ventral and lateral region...

eneration in the zone of Lissauer is less :entrated than in lower thoracic and er lumbar segments and some fibers n this zone enter *lateral parts of the* :erior horn. Dorsal root fibers in the ıal gray are most abundant in the seg- ıt of entry, but are more widely distrib- d in the gray of adjacent spinal seg- nts than other lumbar, or thoracic, dor- roots. The distribution of dorsal root ːrs in the two segments rostral to a zotomy is greater than in adjacent cau- segments. In adjacent segments root ːrs project most abundantly to cell ıups of the posterior horn.

The configuration and laminar organiza- n of the spinal gray at S1 in the monkey fers from that in the cat (Rexed, '52, ı). The S1 spinal segment in the monkey ɜembles S3 in the cat in that laminae III, and V of the posterior horn extend in ntinuity across the midline as in most of e thoracic segments. However, laminae ', VII, VIII and IX at S1 are similar in the t and monkey. At this level where the mbosacral enlargement is disappearing, ɔers of the fasciculus gracilis form a flat- ned, horizontal wedge overlying the ɔrsomedial part of the posterior horn. Fi- ːrs entering the gray traverse medial parts laminae I, II and III and terminate ırgely in medial parts of laminae III and /. The distribution of S1 dorsal root fibers ı certain respects resembles that of ıoracic dorsal roots.

Coccygeal spinal segments in the mon- ey, though smaller, resemble sacral seg- ıents in laminar organization, except that ıere is no lamina VI, cell groups of lamina X are smaller, and the commissural ucleus develops in the medial part of ımina VIII. Dorsal root fibers at this level nter the gray dorsally near the midline at ı considerable distance from the apex of he posterior horn. Dorsal root fibers pass- ng into the spinal gray of the two adjacent ostral segments are distributed princi- ɔally to laminae of the posterior horn. A systematic discussion of the distribution of lower thoracic, lumbosacral and coccygeal dorsal root fibers in the spinal gray must be made with respect to the laminae of Rexed.

Laminae I and II. Observations con- cerning laminae I and II in the lower thoracic, lumbosacral and coccygeal spinal segments are essentially the same as in cervical and upper thoracic segments of the spinal cord. A small number of dorsal root fibers end in medial parts of lamina I. These fibers are inconspicuous in lower thoracic and upper lumbar segments where this lamina is small.

As discussed in Part I of this study, most of the dorsal root fibers seen in lamina II are in passage to deeper laminae of the posterior horn. While the largest number of the dorsal root fibers pass through medial parts of lamina II, there are some which curve around its medial border. Fibers entering in thoracic (fig. 8), upper lumbar, and sacrococcygeal (figs. 11, 12) segments where laminae II through V are fused across the midline seem to traverse more medial parts of lamina II than do fibers in the lumbar enlargement (fig. 10). A modest number of fibers course from the zone of Lissauer through, and to, small lateral parts of lamina II, but this is not constant. These observations are the same as those of Anderson ('60), Ralston ('65), Stein and Carpenter ('65) and Sterling and Kuypers ('67) which indicate that few dorsal root fibers terminate in lamina II.

Using more sophisticated technics Ral- ston ('65) has demonstrated that the sub- stantia gelatinosa corresponds only to lamina II and that the afferent inputs to laminae II and III differ significantly. From these studies it was concluded that lamina III receives a major projection of dorsal root collaterals, while the substantia gelatinosa receives very few. According to Szentágothai ('64), the substantia gelati- nosa is essentially a closed system in which cells are interconnected with each other either by axons running within the substantia gelatinosa or passing via the zone of Lissauer to re-enter the substantia gelatinosa in neighboring segments. This paucity of terminal degeneration in lamina II following dorsal rhizotomy has sug- gested that the cells of the substantia gelatinosa may be excited polysynaptically via interneurons in deeper portions of the posterior horn. There appears to be little evidence that cells of lamina II are acti- vated monosynaptically. In addition to the fibers projected to the spinal gray, fine

fibers from the root entry zone project into the zone of Lissauer. These small fascicles, located mainly in medial portions of the zone of Lissauer, are distributed to a number of segments both caudal and rostral to the root entry zone (Ranson, '13, '14; Earle, '52).

Laminae III and IV. While it is widely acknowledged that dorsal root fibers at all levels project profusely to laminae III and IV (i.e., the nucleus proprius), there are differences of opinion concerning localization of dorsal root terminals in these laminae. As discussed in Part I of this study, Szentágothai and Kiss ('49) consider medial parts of laminae II and III to be related to ventral cutaneous regions, and lateral parts of these laminae to be associated with dorsal cutaneous regions. Studies of dorsal root fibers entering the brachial enlargement of the cord in the cat (Sterling and Kuypers, '67) suggest a differential distribution of fibers in laminae III and IV in which: (1) more rostral roots supplying proximal parts of a limb, or preaxial regions, project preferentially to lateral parts of these laminae, and (2) caudal roots supplying distal parts of a limb, or postaxial regions, project to medial parts of these laminae. Physiological data (Wall, '60, '67) indicate that this type of localization is present also in the lumbar enlargement of the cat. In the most extensive anatomical study (Sprague and Ha, '64) of lumbar dorsal roots in the cat, these authors noted variations in the terminal fields of lumbosacral dorsal roots in laminae III and IV at levels above and below dorsal rhizotomies. This finding and those of Szentágothai and Kiss ('49) and Szentágothai ('64) have supported the thesis of radially oriented "synaptic compartments" or "lobuli" within these laminae. The variations in patterns of localization within these laminae have made it difficult to postulate a general scheme. Data in the monkey are similar to those reported for the cat.

In the monkey lower thoracic, upper lumbar and sacrococcygeal dorsal root fibers enter the posterior horn more medially than do the dorsal root fibers of the lumbar enlargement. Fibers from these roots radiate into medial and central parts of laminae III and IV (figs. 7, 8, 11). degeneration is seen in lateral portions these laminae, it usually is modest amount. In segments above and below t dorsal rhizotomy, fibers enter even m medial regions of the posterior horn a are distributed in essentially the same p tern as at the rhizotomy level, but reduced amounts.

Observations concerning fibers of dors roots of the lumbar enlargement are sim lar in that the greatest concentration terminal fibers at the level of root section seen in medial and central parts of lamin III and IV (fig. 9). However, degeneration these laminae rostral and caudal to th rhizotomies sometimes revealed a diffe ent pattern of localization. In only one ani mal (C-914) with an L7 dorsal rhizotom degeneration in laminae III and IV wa seen predominantly in lateral regions a levels of the root section. Adjacent rostral spinal segments in this animal revealed less degeneration in these laminae distrib uted in "synaptic compartments" as de scribed by Sprague and Ha ('64). Data available from this study suggest that the pattern of distribution of dorsal root fiber in laminae III and IV of the lumbar en largement in the monkey probably i similar to that described by Sterling and Kuypers ('67) for the brachial enlargemen of the cat. Precise confirmation of thes data would seem to require staining o virtually serial sections through severa segments of the spinal cord, a procedur not done in the current study. Further some of the variations seen in our materia undoubtedly are the result of comparing patterns of degeneration at different loca tions within various spinal segments. The establishment of precise patterns of local ization of terminal degeneration in these laminae would seem to require a detailed study of the fibers of each dorsal root in several animals. Even when this is done (Sprague and Ha, '64), it is difficult to generalize broadly on the patterns of local izations in laminae III and IV. It is apparent from the present investigation that more detailed studies of the distribution of dorsal root fibers in these laminae are necessary.

Laminae V and VI. While lamina V is present at all spinal levels, lamina VI is

ound only in the brachial and lumbar nlargements and in upper cervical segnents. Dorsal root fibers enter central and nedial parts of *lamina* V almost excluively from lamina IV, except in the spinal nlargements. In lower *thoracic and upper* umbar segments, dorsal root fibers raversing central and medial regions of amina V appear to be mainly fibers of assage which project ventrally into laminae VII, VIII *and* IX. Virtually no degenerated fibers are present in lateral parts of amina V.

In the cord enlargements a considerable number of dorsal root fibers appear to erminate in medial parts of laminae V and VI. According to Sterling and Kuypers '67), fibers from a single dorsal root of the brachial enlargement of the cat are distributed *to parts of these laminae throughout* the brachial cord. However, there is no evident topical distribution of fibers within laminae V and VI. A slightly different arrangement is present in the brachial enlargement in the monkey (Shriver, Stein and Carpenter, '68) in that brachial dorsal roots *project fibers to medial and central* parts of lamina VI which is maximal in segments rostral to their entry, and present in lesser amounts for variable distances in more caudal segments.

In the lumbar enlargement root fibers enter laminae V and VI from lamina IV, while other fibers enter lamina V directly from the posterior funiculus. Fibers entering lamina V directly are *seen at the level* of the rhizotomy but frequently these are *more impressive* in adjacent spinal segments. These fibers, like those at the level of the root section, appear mainly to arborize about individual cells, but some fibers are in passage to more ventral laminae. Fibers enter laminae V and VI at all levels above the cut root until the L3 level is reached and the dorsal nucleus is evident. At the L3 level and above most of the entering fibers *seem to merely traverse* medial parts of lamina V *en route* to cells of the dorsal nucleus. Degenerated fibers and cells in the medial part of laminae VI seem to merge imperceptibly into the caudal pole of Clarke's column at L3. The relationship between cells of the medial part of lamina VI at this level to the dorsal nucleus resembles a similar relationship of medial lamina VI *in the upper cervical* cord to the caudal cuneate nucleus (Shriver, Stein and Carpenter, '68).

Dorsal roots of the lumbar enlargement also contribute fibers to medial parts of laminae V and VI at levels below the rhizotomy. Fibers descending in the fasciculus *gracilis project to these* laminae as far caudally as S1. Medial parts of laminae V and VI appear to be the principal regions of termination of fibers descending in the fasciculus gracilis from dorsal roots of the lumbar enlargement (figs. 26, 27, 28).

According to Sprague and Ha's ('64) analysis and *correlation of anatomical and* physiological findings (Eccles, Eccles and Lundberg, '60), dorsal root fibers terminating in laminae V and VI are group Ia and Ib muscle afferents, group III muscle afferents, and some cutaneous fibers. Recent physiological studies (Wall, '67) of the laminar *organization of the posterior horn* in the cat indicate that cells in laminae IV, V and VI all respond to cutaneous stimulation but only cells of lamina VI respond to joint movement. Cells of lamina IV differ from those of lamina V in that receptive cutaneous fields are smaller. Cells of *lamina V are described as responding as if* impulses from lamina IV converge upon them. These observations suggest that dorsal root afferents projecting to laminae V and VI may be concerned with the integration of impulses from more than one type of peripheral receptor.

Laminae VII and VIII. Although laminae VII and VIII are recognized at all spinal levels, the configurations of these laminae show variations at different levels. Lamina VII has a much greater extent in the cord enlargements, where its lateral part extends into the anterior horn. Conversely, *lamina* VIII *is reduced greatly in* the cord enlargements where it occupies only the medial part of the anterior horn.

The distribution of dorsal root fibers in laminae VII and VIII appears different in the lower thoracic and upper lumbar regions than in the lumbar enlargement. While the distribution of dorsal root fibers in sacral and coccygeal segments is similar to that of the *thoracic cord, there are* certain distinctive features.

In the lower thoracic and upper lumbar spinal segments (i.e., T8 to L2), dorsal root

fibers from the root entry zone project into the zone of Lissauer. These small fascicles, located mainly in medial portions of the zone of Lissauer, are distributed to a number of segments both caudal and rostral to the root entry zone (Ranson, '13, '14; Earle, '52).

Laminae III and IV. While it is widely acknowledged that dorsal root fibers at all levels project profusely to laminae III and IV (i.e., the nucleus proprius), there are differences of opinion concerning localization of dorsal root terminals in these laminae. As discussed in Part I of this study, Szentágothai and Kiss ('49) consider medial parts of laminae II and III to be related to ventral cutaneous regions, and lateral parts of these laminae to be associated with dorsal cutaneous regions. Studies of dorsal root fibers entering the brachial enlargement of the cord in the cat (Sterling and Kuypers, '67) suggest a differential distribution of fibers in laminae III and IV in which: (1) more rostral roots supplying proximal parts of a limb, or preaxial regions, project preferentially to lateral parts of these laminae, and (2) caudal roots supplying distal parts of a limb, or postaxial regions, project to medial parts of these laminae. Physiological data (Wall, '60, '67) indicate that this type of localization is present also in the lumbar enlargement of the cat. In the most extensive anatomical study (Sprague and Ha, '64) of lumbar dorsal roots in the cat, these authors noted variations in the terminal fields of lumbosacral dorsal roots in laminae III and IV at levels above and below dorsal rhizotomies. This finding and those of Szentágothai and Kiss ('49) and Szentágothai ('64) have supported the thesis of radially oriented "synaptic compartments" or "lobuli" within these laminae. The variations in patterns of localization within these laminae have made it difficult to postulate a general scheme. Data in the monkey are similar to those reported for the cat.

In the monkey lower thoracic, upper lumbar and sacrococcygeal dorsal root fibers enter the posterior horn more medially than do the dorsal root fibers of the lumbar enlargement. Fibers from these roots radiate into medial and central parts of laminae III and IV (figs. 7, 8, 11). If degeneration is seen in lateral portions of these laminae, it usually is modest in amount. In segments above and below the dorsal rhizotomy, fibers enter even more medial regions of the posterior horn and are distributed in essentially the same pattern as at the rhizotomy level, but in reduced amounts.

Observations concerning fibers of dorsal roots of the lumbar enlargement are similar in that the greatest concentration of terminal fibers at the level of root section is seen in medial and central parts of laminae III and IV (fig. 9). However, degeneration in these laminae rostral and caudal to the rhizotomies sometimes revealed a different pattern of localization. In only one animal (C-914) with an L7 dorsal rhizotomy degeneration in laminae III and IV was seen predominantly in lateral regions at levels of the root section. Adjacent rostral spinal segments in this animal revealed less degeneration in these laminae distributed in "synaptic compartments" as described by Sprague and Ha ('64). Data available from this study suggest that the pattern of distribution of dorsal root fibers in laminae III and IV of the lumbar enlargement in the monkey probably is similar to that described by Sterling and Kuypers ('67) for the brachial enlargement of the cat. Precise confirmation of these data would seem to require staining of virtually serial sections through several segments of the spinal cord, a procedure not done in the current study. Further some of the variations seen in our material undoubtedly are the result of comparing patterns of degeneration at different locations within various spinal segments. The establishment of precise patterns of localization of terminal degeneration in these laminae would seem to require a detailed study of the fibers of each dorsal root in several animals. Even when this is done (Sprague and Ha, '64), it is difficult to generalize broadly on the patterns of localizations in laminae III and IV. It is apparent from the present investigation that more detailed studies of the distribution of dorsal root fibers in these laminae are necessary.

Laminae V and VI. While lamina V is present at all spinal levels, lamina VI is

:is nucleus varies. Most authors indicate :at following section of a single dorsal 'ot, degeneration projecting to the dorsal ucleus extends in a span of six or seven gments. Liu ('56) reported the most tensive study of dorsal root projections to e dorsal nucleus. According to this uthor, the T10 dorsal root projected fibers the dorsal nucleus as far rostral as T3 nd as far caudal as L1. The distribution of orsal root fibers from different segments ndicated that those concerned with the unk were found in all regions of the ucleus, while those roots conveying afferents from the tail, hindlimb and forelimb ere confined to specific regions.

The distribution of fibers from a single orsal root in the monkey to cells of the orsal nucleus at various levels seems ven greater than in the cat. Lower horacic and upper lumbar (i.e., T8 through 2) dorsal roots contribute fibers to cells of he dorsal nucleus three to eight segments bove the level of the rhizotomy and one to our segments below. Dorsal roots of the umbar enlargement (i.e., L3 to L7) project nly ascending fibers to the dorsal nucleus. Although these fibers ascend in the osterior columns for variable distances audal to the column of Clarke, they all ontribute fibers to approximately the same portions of this cell column, namely, hat part of the cell column between L3 and T7 segments. While dorsal root fibers from the lumbar enlargement overlap those from upper lumbar and lower thoracic segments, their terminations are restricted to definite portions of Clarke's column. The first sacral dorsal root projects fibers to more restricted caudal parts of Clarke's column. Coccygeal roots do not appear to project fibers to the dorsal nucleus in the monkey, although they have been described in the cat (Liu, '56) as projecting fibers to the dorsal nucleus at the L4 level. Data in the monkey support the observations of Liu ('56) that dorsal root fibers from different parts of the body project to the cells of the dorsal nucleus at particular spinal levels.

There is evidence that dorsal root fibers from various spinal levels project to different parts of the dorsal nucleus. Szentágothai and Albert ('55) noted that in the dog sacral dorsal rhizotomies produced degeneration in the medial part of Clarke's column, while degeneration resulting from lumbar rhizotomies occupied lateral parts of Clarke's column. In the cat Grant and Rexed ('58) found that fibers from sacral dorsal roots were distributed throughout the cross-sectional area of the dorsal nucleus in the four caudal segments containing this nucleus, while more rostral fibers terminated in medial or dorsomedial parts of the column. Lumbar dorsal roots projected fibers to lateral parts of the cell column caudally and to dorsomedial regions of the cell column rostrally. In intermediate regions of the cell column fibers seemed distributed throughout the cross-sectional area of the nucleus.

Dorsal root fibers in lower thoracic and upper lumbar segments of the monkey enter the dorsal nucleus by coursing along its lateral and ventral borders. While at some levels dorsal root fibers are distributed throughout the dorsal nucleus at the level of rhizotomy (e.g., T8, L2 and L3), most of these fibers project to ventrolateral parts of the nucleus (figs. 8, 25). Degenerated fibers from these dorsal roots terminating in the dorsal nucleus at more rostral levels also are localized to parts of the cell group. In the two or three adjacent rostral spinal segments terminal degeneration usually is localized to ventrolateral parts of the nucleus, but further rostrally terminal degeneration occupies small dorsal parts of the nucleus. This change in the intranuclear distribution of fibers appeared related to the fact that at more rostral levels fibers enter the dorsal nucleus directly from the posterior column. Branches of dorsal roots projecting fibers to cells of the dorsal nucleus caudal to the level of the rhizotomy consistently terminate in lateral parts of the nucleus.

Dorsal root fibers of lumbar segments caudal to L4 ascend in the posterior column and enter the column of Clarke by passing ventrally through medial parts of lamina V. It is also possible that some dorsal root fibers entering the medial part of lamina VI throughout the lumbar enlargement may reach the dorsal nucleus by passing rostrally through this lamina. Fibers from all dorsal roots of the lumbar enlargement project profusely throughout the cross-sectional area of the dorsal nucleus

fibers traverse medial parts of laminae III, IV and V and enter portions of lamina VII lateral to the dorsal nucleus. Part of these fibers pass directly to cells of the dorsal nucleus, while others project to the intermediomedial nucleus. Fibers passing through, and adjacent to, the intermediomedial nucleus enter the medial part of lamina VIII, course ventrally along the medial border of the anterior horn, and follow the curve of the spinal gray into ventral and lateral parts of the anterior horn. Fascicles of fibers following this path project to ventromedial and lateromedial cell groups of lamina IX. A smaller and variable number of fibers pass ventrolaterally through central parts of laminae VII and VIII to reach the lateromedial cell group of lamina IX.

Beginning at the L3 level and extending through L7, quantitative changes are seen in the distribution of fibers in laminae VII and VIII. From the region of concentrated fibers in dorsal and central parts of lamina VII, one small bundle of fibers projects ventromedially toward the intermediomedial nucleus (figs. 23, 24). Some fibers with this course terminate in the latter nucleus while others bypass it and enter the dorsomedial part of lamina VIII. The above fibers follow the pattern of distribution previously described for lower thoracic and upper lumbar segments. The major difference in the lumbar enlargement is that fewer fibers projecting in this manner reach cell groups of lamina IX; fibers pursuing this course project mainly to ventromedial and centromedial cell groups. The major fiber projection from dorsal and central parts of lamina VII in the lumbar enlargement consists of numerous small radiating fascicles which stream ventrolaterally through central regions of lamina VII and enter large cell groups of lamina IX located ventrally and laterally (fig. 23). In spinal segments caudal to the rhizotomies greatly reduced amounts of degeneration are seen passing ventrally in lamina VII and only a few of these reach cell groups in lamina IX. Rostral to the rhizotomies more degenerated fibers projecting through central parts of lamina VII reach cell groups of lamina IX.

While data regarding sacral and coccygeal roots in this study are limited, available evidence indicates that the majority [of] these dorsal root fibers course throu[gh] medial parts of laminae VII and V[III]. Sacral dorsal root fibers reach [...] medial cell groups of lamina IX while co[c]cygeal roots project to cells of the [com]missural nucleus (figs. 29, 30). A [sm]all number of diffuse and scattered fibers pa[ss] ventrally through central parts of lamin[ae] VII and VIII at S1 and Co1, but few of the[se] fibers reach cell groups of lamina IX.

Thus fibers entering lamina VII a[nd] projecting to cell groups of lamina I[X] appear to follow two pathways: (1) o[ne] ventromedially through the medi[al] [part of] lamina VIII, and (2) one ventr[olaterally] through central parts of lamina VII. Dor[sal] root collaterals follow both of these pa[th]ways at all spinal levels, but the vent[ro]medial projection is the major one [in] thoracic and upper lumbar segmen[ts], while the projection through central pa[rts] of lamina VII is predominant in the spi[nal] cord enlargements. These findings are [in] agreement with those of Sprague and [Ha] ('64), Sterling and Kuypers ('67), and [our] own observations concerning rostral p[or]tions of the spinal cord (Shriver, Stein a[nd] Carpenter, '68).

At levels studied here, three distinct cell groups can be identified within lami[na] VII: (1) the dorsal nucleus, (2) the inter[me]diomedial nucleus, and (3) the interme[dio]lateral nucleus. The intermediolate[ral] nucleus in thoracolumbar and sacral se[g]ments does not receive dorsal root affe[r]ents. The intermediomedial nucleus [re]ceives a constant small projection at th[e] level of root entrance in all spinal [seg]ments (Schimert, '39; Sprague and Ha[...] Sterling and Kuypers, '67; Shriver, Stei[n] and Carpenter, '68). While the significanc[e] of dorsal root projections to the intermedi[o]medial nucleus is not apparent, the co[n]stancy of this termination at all spinal le[v]els suggests that its cells are interneuro[ns] of importance.

Data concerning the extensive distrib[u]tion of fibers from a single dorsal root to th[e] dorsal nucleus of Clarke have bee[n] reported by Pass ('33), Szentágothai a[nd] Albert ('55), Liu ('56) and Grant and Rexe[d] ('58) for the cat and dog. The distanc[e] above and below the level of the rhizotom[y] at which degeneration has been found [...]

…), there is no clearcut lateral boundary …this fasciculus in the *mid-thoracic* …ion. Thus in the strict sense the fascicu… …s gracilis should consist only of posterior …umn fibers terminating in the nucleus …acilis. Our data indicate that a few fibers …m all lower thoracic and the L1 dorsal …t project to the cuneate nucleus, …hough the bulk of these fibers termi…tes in the nucleus gracilis. Ascending …rsal root fibers from lower thoracic and …per lumbar dorsal roots form oblique …rrow bands in the fasciculus gracilis …th fibers from thoracic segments most …teral (figs. 1, 13, 14). Central lumbar dor…l roots (L4, L5 and L6) contribute larger …mbers of fibers to the fasciculus gracilis …hich occupy larger areas (figs. 3, 18). …hile all ascending dorsal root fibers over…p those from adjacent segments, the …erlap of central lumbar fibers appears …eatest. Ascending fibers from the most …udal dorsal roots (i.e., L7, S1 and Co1) …itially occupy more medial regions of the …sciculus gracilis (figs. 15, 16, 17), but in …pper thoracic segments, these fibers …read out over the dorsal surface of the …sciculus (figs. 3, 18). At cervical levels …bers from these caudal roots assume a …urved configuration in which fibers are …resent medially near the raphe and …xtend dorsally along the surface of the …sciculus. The manner in which fibers …om caudal roots are organized in the …hesus monkey is remarkably similar to …at described in the spider monkey (Chang …nd Ruch, '47).

The number of fibers contributed to the …sciculus gracilis and the area which they …ccupy appears related to the size of the …orsal root. The physiological implication …f these findings is that the lower lumbar …oots (L4, L5, L6 and L7) probably convey …ensation from the most important cutane…us areas. This view appears consistent …vith the conclusions of Mott and Sherring…on (1895).

Dichotomizing descending branches of …orsal root fibers in the posterior columns …ave been described as relatively small …ompact bundles. Fibers from thoracic and …umbar dorsal roots are said to form an …val bundle adjacent to the posterior median septum (Flechsig, 1876), while similar fibers from sacral roots descend in an extreme dorsomedial part of the fasciculus gracilis (Gombault and Philippe, 1894). The extent of these descending fibers varies with different spinal roots. In the cat Liu ('56) found descending fibers, irregularly distributed in the posterior funiculus, extending four to six segments caudal to the level of entrance. In one human case (Collier and Buzzard, '03) fibers from the L3 dorsal root descended eight segments in the posterior columns.

Observations concerning descending branches of dorsal root fibers in the monkey indicate that these fibers are arranged in oblique bands within various parts of the fasciculus gracilis and do not form small compact bundles. The organization of these narrow bands resembles that of ascending fibers in the posterior columns, although descending bands undergo a relatively rapid attenuation and dissolution with increasing distance from the sectioned dorsal root, as fibers are given off to the spinal gray. Fibers from lower thoracic and upper lumbar roots occupy the lateral third of the fasciculus gracilis while fibers from lower lumbar roots are in more medial parts of the fasciculus gracilis. In most instances these fibers projected caudally two to four spinal segments. Descending fibers from lower thoracic and the upper three lumbar dorsal roots project to lateral parts of the dorsal nucleus and adjacent portions of lamina VII. More caudal lumbar dorsal roots project fibers into the medial part of lamina VI and adjacent parts of lamina VII.

Medullary projections

Dorsal root projections to the nucleus gracilis. According to Ferraro and Barrera ('35) and Walker and Weaver ('42), ascending dorsal root fibers projecting to the nucleus gracilis are somatotopically organized. Fibers from sacral dorsal roots were found to terminate in caudal and medial parts of the nucleus gracilis, while lumbar dorsal root fibers projected to slightly more rostral and lateral parts of the nucleus. Lower thoracic roots were described as terminating in the most rostral and lateral parts of the nucleus gracilis. The limited number of dorsal rhizotomies and limitations of the …

at L3 and L2 (figs. 19, 20, 22); at L1 and in lower thoracic segments terminal degeneration within the dorsal nucleus is consistently and discretely localized to dorsomedial regions (fig. 21). These observations are similar to those of Grant and Rexed ('58) and support the thesis that dorsal root afferents to the dorsal nucleus terminate in an overlapping but spatially organized topical manner. There is abundant physiological evidence that group Ia and Ib afferents from stretch receptors terminate upon cells of this nucleus which in turn conveys impulses to the cerebellum via the posterior spinocerebellar tract (Lloyd and McIntyre, '50; Oscarsson, '65).

Lamina IX. Collaterals of lower thoracic, lumbosacral and coccygeal dorsal roots have a more restricted segmental distribution in cell groups of lamina IX, than similar collaterals in laminae of the posterior horn or in lamina VII. In lower thoracic and upper lumbar segments collaterals of dorsal root fibers reaching lamina IX are virtually restricted to the segment of entry. In the lumbar enlargement the number of dorsal root collaterals reaching cell groups of lamina IX is greater and these fibers are distributed to this lamina not only at the level of root entry but in the two adjacent spinal segments above and below. Fibers passing to adjacent spinal segments are modest in number, but definite. The two pathways by which dorsal root fibers reach cell groups of lamina IX have been discussed in relationship to laminae VII and VIII. These observations for the lumbar enlargement are similar to those of Spargue and Ha ('64), but these authors made no comparisons between thoracic and lumbar levels. Sacral and coccygeal dorsal root fibers conform to the pattern described for the lower thoracic dorsal roots in that collaterals pass mainly to medial cell groups of lamina IX via the medial part of lamina VIII.

According to Sprague ('58), lumbar dorsal root collaterals in the cat establish both axosomatic and axodendritic contacts with motor neurons in their segment of entry, but in adjacent segments connections are primarily axodendritic. In the present study most dorsal root collaterals appeared to terminate in the dendritic fields of motor neurons, an observation which might be due only to longer postoperative survival of our animals. In this connection it is of interest that Golgi studies of motor and interneurons of the anterior horn (Sprague and Ha, '64) indicate that dendrites of motor neurons extend beyond the limits of Rexed's lamina IX into adjacent portions of laminae VII and VIII.

Projections in the posterior funiculus. Classic descriptions of ascending fibers in the posterior columns in the rhesus monkey have been based upon the study of Marchi degeneration following selective dorsal root section and various types of cordotomies (Ferraro and Barrera, '35; Walker and Weaver, '42). The most detailed study (Walker and Weaver, '42) included data from eight dorsal rhizotomies and 16 cordotomies; only six dorsal roots were sectioned caudal to T8. All authors have reported an oblique laminar arrangement of ascending fibers in the fasciculus gracilis in which lower thoracic fibers were most lateral and sacral fibers most dorsomedial. Composite diagrams illustrating the somatotopic arrangement of ascending fibers in the posterior columns show quantitative differences in the number of ascending fibers and in the areas they occupy for different dorsal rhizotomies. These authors emphasized that ascending fibers from adjacent dorsal roots exhibit extensive overlap in the posterior columns. Intermingling of ascending dorsal root fibers was especially pronounced for lumbar and sacral root fibers in cervical spinal segments. Observations concerning the organization of the posterior columns in the spider monkey (Chang and Ruch, '47) based upon complete and partial transections of the spinal cord, indicated that fibers ascending from coccygeal regions initially occupy dorsomedial parts of the fasciculus gracilis but at thoracic levels form a band or cap along the dorsal margin of the fasciculus gracilis.

The current study based upon more extensive data in the monkey confirms prior investigations and reconciles certain minor differences. Dorsal roots from the five lowest thoracic segments (T8 through T12) and all more caudal spinal segments project fibers rostrally in the fasciculus gracilis. As noted by Walker and Weaver

t" region in that they receive the princi- projections from the cerebral cortex.

The concept that neurons in various trocaudal parts of the nucleus gracilis hibit differences with respect to: (1) size their peripheral field, and (2) the sensory dality they relay to higher levels of the uraxis, is difficult to reconcile with the servations of Mountcastle and Powell 9) and Poggio and Mountcastle ('60) ncerning tactile sense and kinesthesis. ese authors conclude that: "the ventro- sal complex of the thalamus is the of thalamic transfer of the medial mniscus component of the somatic affer- t system, a component defined as com- ising those fibers whose cells of origin lie ithin the dorsal column nuclei." Each euron of the ventrobasal complex: (1) is lated to restricted, specific and unchang- receptive fields on the contralateral de of the body, and (2) can be activated by ither superficial mechanical stimulation f the skin or mechanical alteration of deep tructures (especially joint rotation), but nly by one of these. These neurons, egarded as modality specific, are thought o contribute almost exclusively to percep- ion of tactile sense and kinesthesis. These indings strongly suggest that the somato- opic organization for specific modalities, onveyed to the thalamus via the posterior olumns and the medial lemniscus, must e maintained at the only known synaptic junction of this system, the nuclei gracilis and cuneatus. Mountcastle and Powell ('59) clearly state: "At each level elements devoted to this submodality (i.e., kinesthe- sis) are intermingled with those of others (i.e., tactile sense) in a single and mutual somatotopic pattern."

The physiological studies of Kruger, Siminoff and Witkovsky ('61), based upon single neuron analysis of dorsal column nuclei in the cat, provide no clear sugges- tion of a rostro-caudal differentiation either in terms of somatotopy or modality segre- gation. It was further clear from this study that the entire body was represented at all rostrocaudal levels explored in detail. The evidence provided by these authors appears to substantiate the findings of Mountcastle and his colleagues and data available from the present investigation.

Observations in the current study concerning the somatotopic termination of ascending dorsal root fibers in the nucleus gracilis appear to support the studies of Mountcastle and his colleagues. The somatotopic termination of dorsal root fibers in the nucleus gracilis, while very irregular for certain dorsal roots, seems to extend throughout the nucleus. It is characterized by relatively regular zones of termination for dorsal root fibers related to the lower thoracic, upper lumbar and sacral segments, relatively irregular and larger zones of termination for fibers from the lumbar enlargement, and a high degree of overlapping terminations, especially for the lower lumbar roots (figs. 2, 4, 5, 6). The area of the terminal zones for fibers from different dorsal roots varies. Terminal zones for the lower thoracic and upper lumbar dorsal root fibers, arranged parallel to the lateral border of the nucleus gracilis, are relatively small (fig. 31). The terminal zone for root fibers of Co1, located dorsomedial throughout most of the nucleus, also is relatively small (fig. 36). The lower lumbar dorsal roots terminate in overlapping, irregularly shaped zones in the central region of the nucleus (figs. 4, 5, 34). The S1 dorsal root terminal zone is fairly large, located dorsomedially, and overlaps portions of the central part of the nucleus (fig. 35). The largest terminal zones in the nucleus gracilis are those related to the lower four lumbar dorsal roots.

Attempts to determine the somatotopic termination of dorsal root fibers in the nucleus gracilis are much more difficult than similar studies for the nucleus cuneatus, because: (1) the nucleus gracilis has a greater rostrocaudal extent, (2) the fibers of the fasciculus gracilis are intermingled with irregularly shaped cell groups for long distances, and (3) there is greater difficulty in establishing whether degeneration is terminal or in passage. While there probably are some areas within the nucleus gracilis that receive exclusively the ascending fibers from particular dorsal roots, such exclusive areas are much smaller than those in the nucleus cuneatus.

Comparison of dorsal root projections to the medullary relay nuclei. A comparison of dorsal root projections to different medullary relay nuclei in the monkey

precluded a more detailed analysis of terminations in this relay nucleus.

Recent studies (Hand, '66) of lumbosacral dorsal root terminations in the nucleus gracilis of the cat employing silver staining methods have demonstrated a different organization. This author relates his observations to cytologically distinct subdivisions of the nucleus gracilis. Three cytologically distinct regions of the nucleus gracilis have been recognized (Cajal, '09; Taber, '61; Kuypers and Tuerk, '64): (1) a reticular region rostral to the obex characterized by a loose organization of cells, (2) a "cell nest" region caudal to the obex characterized by cell clusters, and (3) a caudal region of the nucleus characterized by fewer cells occurring singly or in small clusters. Although a definite somatotopic lamination was found in the nucleus gracilis, generally similar to that reported by Ferraro and Barrera ('35) and Walker and Weaver ('42), this lamination was found chiefly in the "cell nest" region and was not demonstrable in the reticular region where terminal degeneration was diffuse with impressive intersegmental overlap. In the "cell nest" region, dorsal root fibers related to more distal parts of the body terminated more dorsally than those related to proximal parts of the body. Of special interest was the observation that the density and extent of degeneration in the nucleus gracilis did not appear to be related to the size of the peripheral dermatome, but to its proximal or distal position on the body surface. It has been suggested that this finding may be related to the greater density of cutaneous receptors in more distal dermatomes.

Physiological studies in the cat suggest that neurons in the nucleus gracilis exhibit rostrocaudal differences with respect to: (1) the size of peripheral receptive fields which supply afferent input (Gordon and Paine, '60; McComas, '63), and (2) segregation of sensory modality (Kuhn, '49; Perl, Whitlock and Gentry, '62; Gordon and Jukes, '64; Winter, '65). Rostral portions of the nucleus (i.e., reticular region) are said to be related to deep pressure and joint movement, while the "cell nest" and caudal regions of the nucleus are related to hair and skin receptors.

Anatomical studies (Kuypers and Tuerk, '64) of the nuclei cuneatus and gracilis in the cat provide information concerning dendritic architecture, dorsal root terminations, and cells contributing fibers to the medial lemniscus. Cell clusters in the nucleus gracilis are said to consist of large round cells arranged in a circular fashion which possess densely packed dendritic trees composed of many bushy dendrites. Cells in rostral and caudal parts of the nucleus gracilis appear to be triangular, smaller, and possess long radiating dendrites. Dorsal root fibers from L6, L7 and S1 distributed fibers to both rostral and caudal parts of the nucleus gracilis. Caudally fibers were distributed to cell clusters in medial parts of the nucleus. Rostrally fibers were distributed more diffusely. These authors postulated that regions of the nucleus gracilis having cells with small densely packed dendritic trees probably maintain small receptive fields, while those regions of the nucleus characterized by cells with radiating dendrites may maintain larger receptive fields. Their anatomical findings were considered to correlate closely with the physiological observations of Gordon and Paine ('60). These data have been interpreted (Kuypers and Tuerk, '64) as suggesting that the cell cluster regions of the nucleus gracilis possess a greater spatial resolving power than regions containing smaller cells with long radiating dendrites. Cell clusters in the so-called "cell nest" region were considered to be related to small receptive fields that convey impulses largely from hair receptors (Perl, Whitlock and Gentry, '62). Cells in the rostral part of the nucleus gracilis with radiating dendrites were considered to respond to proprioceptive stimuli (Kuhn, '49).

Following section of the medial lemniscus in the upper midbrain of kittens Kuypers and Tuerk ('64) reported chronic retrograde cell changes mainly in the cell clusters of the "cell nest" region of the nucleus gracilis. While these data were not interpreted to mean that cells in the rostral part of the nucleus gracilis do not contribute fibers to the medial lemniscus, they supported the thesis that rostrally located cells at least react differently to axonal interruption. These rostral cells of the nucleus gracilis also differ from those of the "cell

izotomy and progressively diminishes at creasing distances from the sectioned ot; progressive diminution of degeneration is greatest in the anterior horn and in segments caudal to the dorsal izotomy.

2. Lower thoracic and upper lumbar dorsal roots distribute fibers within the spinal gray, which are virtually limited to one segment above and below root entry, except for fibers projecting to the dorsal nucleus.

3. *Dorsal roots of the lumbar enlargement* distribute fibers within certain parts of the spinal gray at the level of rhizotomy and in the two adjacent spinal segments above and below, except for fibers projecting to medial parts of lamina VI and the dorsal nucleus. The first coccygeal dorsal root distributes fibers in the spinal gray in the segment of entry and in the two adjacent rostral spinal segments.

4. Lower thoracic, lumbar and sacrococcygeal dorsal roots project fibers to medial and central parts of laminae III and IV at the level of the rhizotomy; lower thoracic, upper lumbar and sacrococcygeal dorsal root fibers terminate in more medial regions of these laminae than do fibers of dorsal roots in the lumbar enlargement.

5. Dorsal root fibers of the lumbar enlargement project fibers to cells in the medial part of lamina VI throughout all levels of the lumbar enlargement.

6. Dorsal root fibers from lower thoracic, lumbar and sacral segments project to the dorsal nucleus; fibers from lower thoracic and upper lumbar dorsal roots reach portions of the dorsal nucleus three to eight segments above the rhizotomy level and one to four segments below it. Dorsal root fibers from the lumbar enlargement project to portions of Clarke's column caudal to T7 with the most profuse terminations at the L2 and L3 levels.

7. Dorsal root fibers projecting to portions of the dorsal nucleus above the level of rhizotomy change their dispostition at various levels, but ultimately become localized to dorsomedial parts of the nucleus; descending fibers from these roots terminate in lateral parts of the dorsal nucleus at nearly all levels.

8. Fibers from lower thoracic, lumbar, sacral and coccygeal dorsal roots project to the intermediomedial nucleus, usually maximally, at the segment of entry.

9. Fibers from lower thoracic, lumbar, and sacral dorsal roots project to cell groups of lamina IX by two pathways: (a) via medial parts of lamina VIII, and (b) via central parts of lamina VII. Lower thoracic, upper lumbar and sacral dorsal root fibers reach cell groups of lamina IX primarily via medial parts of lamina VIII, while dorsal root fibers of the lumbar enlargement mainly traverse central parts of lamina VII.

10. Ascending fibers in the fasciculus gracilis arise from the dorsal roots of T8 and all more caudal spinal segments; fibers issuing from dorsal roots of the lumbar enlargement contribute the largest number of fibers.

11. Ascending dorsal root fibers in the fasciculus gracilis exhibit only a general segregation because of extensive overlap; fibers from rostral spinal segments are lateral and those from caudal segments are medial.

12. Descending collaterals of dorsal root fibers, arranged in oblique bands in parts of the fasciculus gracilis comparable to that of ascending fibers of the same dorsal root, project caudally for two to four spinal segments, and enter: (a) lateral portions of the dorsal nucleus in lower thoracic or upper lumbar segments, or (b) medial parts of lamina VI throughout the lumbar enlargement.

13. Ascending collaterals of lower thoracic, lumbar, sacral and coccygeal dorsal roots project in a somatotopic and overlapping fashion throughout the rostrocaudal extent of the nucleus gracilis. Zones of the nucleus gracilis receiving terminal dorsal root fibers are organized, so that: (a) roots of the lumbar enlargement project to irregular shaped areas in the central core region of the nucleus, (b) lower thoracic and upper lumbar roots project in serial fashion to narrow oblique laminae lateral to the central core region, and (c) sacral and coccygeal dorsal roots project in serial fashion to crescent-shaped laminae in dorsomedial parts of the nucleus.

14. The areas of the terminal projection zones of dorsal root fibers in the nucleus gracilis: (a) are related to the size of the dorsal root and the number of ascending

reveals both similar and unique features. Fibers from all cervical and the upper seven thoracic dorsal roots project in a somatotopic fashion upon portions of the cuneate and accessory cuneate nuclei. Fibers from lower thoracic, lumbar, sacral and coccygeal dorsal roots project somatotopically upon portions of the nucleus gracilis. In addition a few fibers from the lower thoracic and the first lumbar dorsal roots appear to project within the medial border of the nucleus cuneatus. No fibers from dorsal roots caudal to T7 project fibers to the accessory cuneate nucleus.

In Part I (Shriver, Stein and Carpenter, '68) of this study comparisons were made of dorsal root projections to the cuneate and accessory cuneate nuclei. Briefly, this comparison indicated that the patterns of terminations of C1 to T1 dorsal root fibers in these nuclei was remarkably similar, in that: (1) fibers from C5 to C8 terminated in a central core region, while fibers from other dorsal roots were arranged in oblique laminae about this core, and (2) fibers from the most rostral root (i.e., C1) terminated ventrolaterally, while those from T1 ended dorsomedially. Ascending dorsal root fibers projecting to these nuclei terminated in relatively discrete, but overlapping areas; fibers from one dorsal root mainly overlapped part of the territory of the next highest root.

Ascending fibers from the lower four lumbar dorsal roots terminate in a central core region comparable to that described for the cuneate and accessory cuneate nuclei. The zones of termination of these fibers in the nucleus gracilis are characterized by their large size, irregular configurations and extensive overlap. Fibers projecting to the central core region in the cuneate nucleus terminate in relatively regular wedge-shaped areas that have a consistent patterned overlap. In both the nuclei gracilis and cuneatus the area of the terminal zones bears a definite relationship to the size of the dorsal root and the number of ascending fibers in the posterior columns.

Dorsal root fibers terminating in laminae surrounding the central core region in the nucleus gracilis form narrower, more regularly shaped, primarily vertically-oriented laminar zones which overlap each other in a lateral to medial sequence. Fibers f[rom] dorsal roots rostral to the lumbar [enlarge]ment terminate lateral to the central [core] region, while fibers from roots [caudal to] the enlargement terminate dorsome[dial] to the core region. In the cuneate [and] accessory cuneate nuclei the laminar [ar]rangement of the terminal zones [of] all cervical and the first thoracic dor[sal] roots is more nearly horizontal. Fi[bers of] the first thoracic dorsal root termi[nate] most dorsomedially, while those from [C1] are most ventrolateral. This difference [in] orientation of terminal laminar zones s[een] in these medullary relay nuclei may be d[ue] to the lateral shift and external rotation [of] the alar plate of the myelencepha[lon] which occurs with the development of t[he] fourth ventricle. Thoracic dorsal roots ca[u]dal to T1 form the narrowest termi[nal] laminae in all three of the medullary re[lay] nuclei. Upper thoracic dorsal root fib[ers] terminate in lateral parts of the access[ory] cuneate nucleus, and in dorsomedial pa[rts] of the cuneate nucleus. Lower thora[cic] dorsal root fibers terminate in lateral p[or]tions of the nucleus gracilis.

Comparisons of the laminar termin[a]tions in the medullary relay nuclei indica[te] that fibers of nearly all dorsal roots studi[ed] project throughout the rostrocaudal exte[nt] of their respective medullary relay nucl[ei]. The degree of overlapping fiber termina[tions] in the central core region is muc[h] greater in the nucleus gracilis than in t[he] cuneate or accessory cuneate nuclei. Th[is] suggests that in the cuneate and acc[essory] cuneate nuclei there is a higher degree [of] autonomous terminal representation [of] individual dorsal root fibers than in t[he] gracile nucleus. Further this pattern [of] autonomous and overlapping termina[l] zones, clearly demonstrated in the cuneat[e] and accessory cuneate nuclei, applies to a[ll] cervical and the first thoracic dorsal root[s]. Fibers from all more caudal dorsal roots, including those from the lumbar enlarge[-]ment, appear to have predominantly over[-]lapping terminal projections in their respective medullary relay nuclei.

CONCLUSIONS

The following conclusions were drawn from this study:

1. In the monkey spinal degeneration is most profuse at the level of the dorsal

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fibers in the fasciculus gracilis, and (b) are arranged in an overlapping lateral (rostral dorsal root) to medial (caudal dorsal root) sequence.

15. The degree of overlapping terminations of dorsal root fibers in the nucleus gracilis is more extensive and irregular than in the cuneate or accessory cuneate nuclei, and suggests that there is less autonomous terminal representation of individual dorsal root fibers in the nucleus gracilis.

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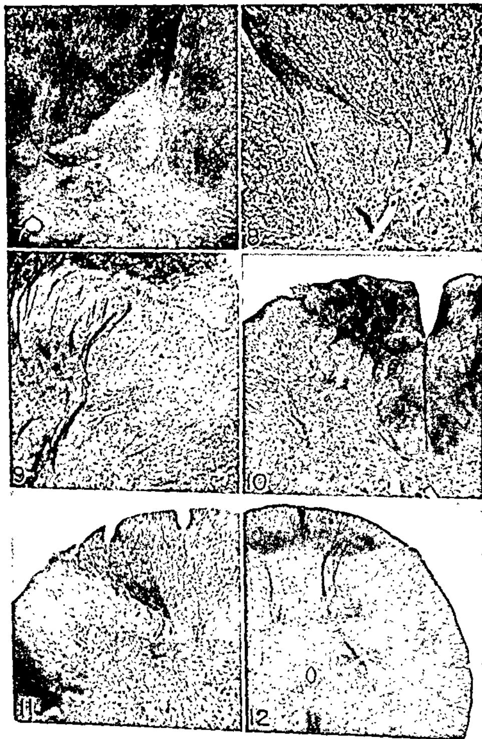

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PLATE 1

EXPLANATION OF FIGURES

7 Rhesus C-1017. Photomicrograph of degeneration at level of root entry following section of the right T8 dorsal root. Note degeneration in medial and central parts of laminae III and IV and in the dorsal nucleus. Nauta-Gygax. ×65.

8 Rhesus C-1012. Photomicrograph of degeneration at T12 following section of the left dorsal root at that level. Note fascicles of degeneration coursing through, and curving around, the medial portion of lamina II to enter medial parts of laminae III and IV and the ventrolateral portion of the dorsal nucleus. Nauta-Gygax. ×65.

9 Rhesus C-1011. Photomicrograph of degeneration at L4 following right dorsal rhizotomy at that level. Degeneration is present in medial and central parts of laminae III and IV and lateral parts of laminae II through IV. Note the relatively clear zones in the most medial portions, and the lateral half, of laminae III and IV. Nauta-Gygax. ×65.

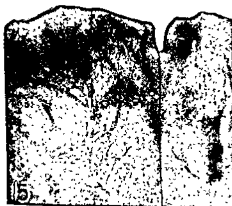
10 Rhesus C-914. Photomicrograph of degeneration at L7 following section of the left dorsal root at that level. Note degeneration in the lateral two-thirds of the fasciculus gracilis dorsal and medial to the posterior horn. Nauta-Gygax. ×37.

11 Rhesus C-912. Photomicrograph of degeneration at the level of root entry following left S1 dorsal rhizotomy Root fibers enter medial and central portions of laminae III and IV mainly by coursing through medial parts of lamina II. Nauta-Gygax. ×42.

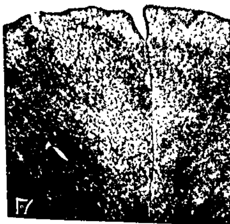
12 Rhesus C-1014. Photomicrograph of degeneration at Co1 following section of the right dorsal root at that level. Note the medial position of fascicles of fibers entering the spinal gray and the large triangular area of degeneration in central lamina VII. Nauta-Gygax. ×33.


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PLATE 2

EXPLANATION OF FIGURES

13 Rhesus C-1017. Photomicrograph of degeneration at the T2 level following section of the right T8 dorsal root. The ventromedial portion of the band of ascending degeneration is present in the lateral part of the fasciculus gracilis, while the dorsolateral portion is located in medial parts of the fasciculus cuneatus. Nauta-Gygax. × 65.

14 Rhesus C-916. Photomicrograph of degeneration at the T2 level following section of the left T10 dorsal root. These ascending fibers form a narrow band in the fasciculus gracilis medial, parallel and close to the posterior intermediate septum. Nauta-Gygax. × 65.

15-16 Rhesus C-914. Photomicrographs of ascending degeneration in medial portions of the fasciculus gracilis at the L5 (fig. 15) and the L3 (fig. 16) levels following left L7 dorsal rhizotomy. In figure 16 degenerated dorsal root fibers can be seen entering the dorsolateral part of the fasciculus gracilis following right L3 dorsal rhizotomy. Nauta-Gygax. × 37, × 33.

17 Rhesus C-912. Photomicrograph of degeneration in medial parts of the fasciculus gracilis at the T12 level after section of the left S1 dorsal root. Note that this crescent-shaped band of ascending fibers is contiguous with the posterior median septum ventrally, but separated from it dorsally by a small medial wedge of normal fibers. Nauta-Gygax. × 68.

18 Rhesus C-1014. Photomicrograph of ascending degeneration in the fasciculus gracilis at the T3 level resulting from section of dorsal roots L6 (left) and Co1 (right). The large band of degeneration in medial parts of the fasciculus cuneatus resulted from section of the right C8 dorsal root. Drawings in figure 3 show both ascending and descending degeneration in the posterior columns resulting from sections of these roots. Nauta-Gygax. × 40.

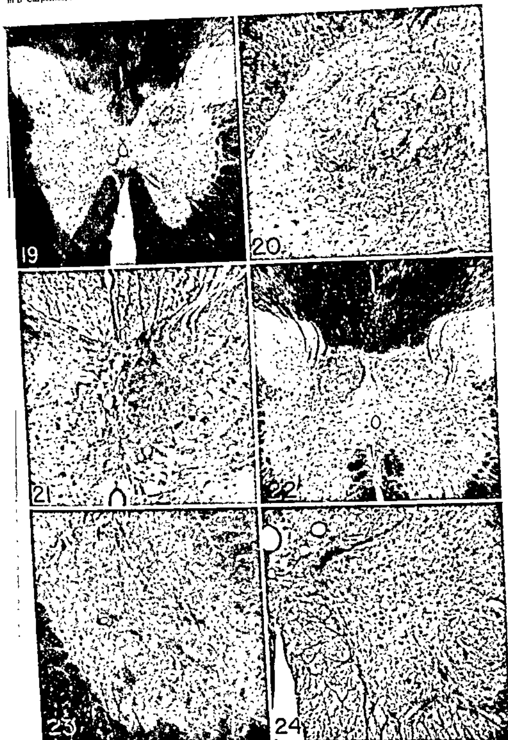

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PLATE 3

EXPLANATION OF FIGURES

19-20 Rhesus C-1011. Photomicrographs of terminal degeneration in the dorsal nucleus at the L2 level following section of the right L4 dorsal root. Note that the degeneration is distributed widely to virtually all parts of the dorsal nucleus. Nauta-Gygax. × 30, × 100.

21 Rhesus C-916. Photomicrograph of right degenerated L6 dorsal root fibers terminating within a small dorsomedial region of the dorsal nucleus at the L1 level. Nauta-Gygax. × 95.

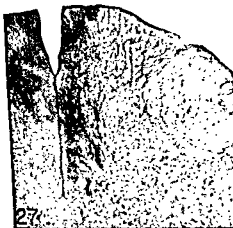
22 Rhesus C-1014. Photomicrograph of degenerated fibers from the left L6 dorsal root terminating throughout the entire cross-sectional area of the dorsal nucleus at the L2 level. Nauta-Gygax. × 45.

23 Rhesus C-1011. Photomicrograph of L4 dorsal root fibers on the right projecting ventromedially to the intermediomedial nucleus and the dorsomedial part of lamina VIII and ventrally through central parts of lamina VII to cell groups of lamina IX at L4. Nauta-Gygax. × 50.

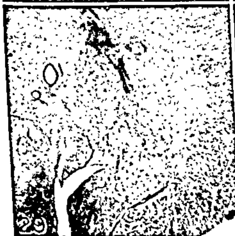
24 Rhesus C-914. Photomicrograph of L3 dorsal root degeneration on the right projecting to the intermediomedial nucleus and into the most medial part of lamina VIII at L5. Nauta-Gygax. × 35.


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PLATE 4

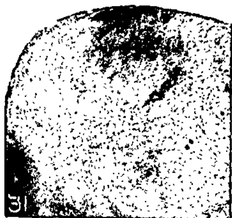
EXPLANATION OF FIGURES

25 Rhesus C-912. Photomicrograph of right L1 dorsal root fibers projecting into laminae III and IV, and directly and profusely to cells of the dorsal nucleus at the level of root section. Nauta-Gygax. × 30.

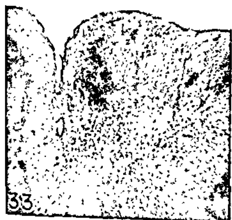
26 Rhesus C-914. Photomicrograph of descending L3 dorsal root fibers projecting to medial parts of lamina VI at L5 on the right side. Nauta-Gygax. × 165.

27-28 Rhesus C-916. Photomicrographs of right L6 dorsal root fibers projecting from the fasciculus gracilis into medial and central parts of lamina VI at L7 (fig. 27) and L3 (fig. 28) levels. Nauta-Gygax. × 40, × 100.

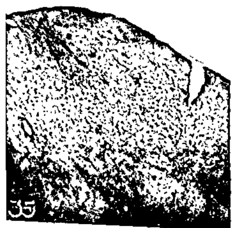
29-30 Rhesus C-1014. Photomicrographs of right Co1 dorsal root fibers passing ventrally from lamina VII through medial parts of lamina VIII to the commissural nucleus within this lamina. In figure 30 these fibers are seen closely surrounding cells of this nucleus. Nauta-Gygax. × 68, × 165.


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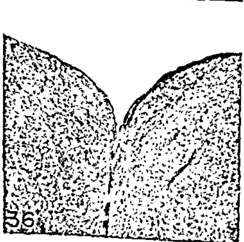

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PLATE 5

EXPLANATION OF FIGURES

31 Rhesus C-1017. Photomicrograph of degenerated fibers from the right T8 dorsal root terminating in the lateral border of the nucleus gracilis. Nauta-Gygax. × 30.

32 Rhesus C-1011. Photomicrograph of right degenerated L4 dorsal root fibers terminating rather diffusely in dorsolateral portions of the nucleus gracilis. Nauta-Gygax. × 35.

33 Rhesus C-1012. Photomicrograph of right degenerated L5 dorsal root fibers terminating ventrally and centrally in middle portions of the nucleus gracilis. Nauta-Gygax. × 30.

34 Rhesus C-1014. Photomicrograph of left L6 dorsal root fibers terminating in irregular shaped central portions of the nucleus gracilis. (See fig. 36). Nauta-Gygax. × 30.

35 Rhesus C-912. Photomicrograph of left S1 dorsal root fibers terminating in a crescent-shaped area in the dorsomedial portion of the nucleus gracilis. Nauta-Gygax. × 70.

36 Rhesus C-1014. Photomicrograph of L6 (left) and Co1 (right) dorsal root fibers terminating in portions of the nuclei gracilis. Nauta-Gygax. × 30.

ɔcutaneous Implantation 'genous Rat Molars[1]

JAMES T. IRVING AND JAMES A. BOND[2]
Forsyth Dental Center, Boston, Massachusetts

TRACT Maxillary molars were extracted, and decalcified or devitalized prior to utaneous implantation in the same animals. All molar implants attracted large numbers of ocytes during the first week of implantation. After two weeks small basophilic mononuclear . appeared which penetrated the dentinal tubules of decalcified molars and were associated ı considerable resorption of dentin and predentin. Giant cells were not usually observed near ılcified molars, but large numbers of them were attracted to FDNB treated molars. Giant ; appeared in moderate numbers around devitalized molars and were associated with ırption of intact root surfaces after 56 days. The significance of these results is discussed.

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; and Migliore ('65) conducted a sing implants of chemically treated .treated autogenous collagen and o determine what influence the al and physical nature of the t has on the cellular response. Giant 'ere not observed around collagen ts which were not chemically , and several different kinds of :al treatment had no effect on the r response. On the other hand, a giant cell response was observed collagen was treated prior to im- tion with 1-fluoro-2,4-dinitrobenzene 3). The *untreated bone* implants d a mile giant cell response which ıhanced in bone treated with FDNB. These findings showed that there is a high degree of specificity of the chemical changes induced in both isolated collagen and bone, which govern the appearance of giant cells.

Since teeth, under most circumstances, do not readily resorb, it appeared of interest to study the tissue reaction to implanted molars and to relate the findings to resorption of bone implants. The present study was designed to utilize the subcutaneous environment (Irving and Handelman, '63) for the implantation of autogenous rat molars.

METHODS AND MATERIALS

Holtzman strain 23 day old male albino rats (65–70 gm) were used in the study. Thirty to sixty animals were used in each experiment, with 4–6 animals for each implantation period.

Surgical procedures. Maxillary first molars were extracted under ether anesthesia, placed in test tubes, and treated as follows.

Preparation of molars for implantation

Group I. Devitalized molar implants. Extracted molars were scraped free of food debris and adherent alveolar bone, then placed in an aqueous antibiotic solu-

[1] This work was supported by U.S. Public Health grant DE-1592, National Institute of Dental Research.
[2] A Postdoctoral Research Fellow in Orthodontics supported by the National Institute of Dental Research while this work was in progress.

:he Subcutaneous Implantation ·f Autogenous Rat Molars [1]

JAMES T. IRVING AND JAMES A. BOND[2]
Forsyth Dental Center, Boston, Massachusetts

ABSTRACT Maxillary molars were extracted, and decalcified or devitalized prior to subcutaneous implantation in the same animals. All molar implants attracted large numbers of leukocytes during the first week of implantation. After two weeks small basophilic mononuclear cells appeared which penetrated the dentinal tubules of decalcified molars and were associated with considerable resorption of dentin and predentin. Giant cells were not usually observed near decalcified molars, but large numbers of them were attracted to FDNB treated molars. Giant cells appeared in moderate numbers around devitalized molars and were associated with resorption of intact root surfaces after 56 days. The significance of these results is discussed.

A method for studying bone resorption ınder relatively standardized environmenal conditions was described by Irving and ·Iandelman ('63). They implanted, sub:utaneously, pieces of autogenous rat scap.ılae and observed the cellular response. Large multinucleated giant cells rapidly aggregated around the bone implants and were associated with extensive resorption. The response was the same for bone implants which were either decalcified, or devitalized by repeated freezing and thawing. On the other hand, rachitic osteoid did not attract giant cells until it has become calcified after implantation. The giant cells were similar, in most respects, to osteoclasts, and Irving and Handelman ('63) concluded that the subcutaneous implantation of autogenous bone was a valid model for the study of osteoclasts.

Irving and Migliore ('65) conducted a study using implants of chemically treated and untreated autogenous collagen and bone to determine what influence the chemical and physical nature of the implant has on the cellular response. Giant cells were not observed around collagen implants which were not chemically treated, and several different kinds of chemical treatment had no effect on the cellular response. On the other hand, a strong giant cell response was observed when collagen was treated prior to implantation with 1-fluoro-2,4-dinitrobenzene (FDNB). The untreated bone implants showed a mile giant cell response which was enhanced in bone treated with FDNB.

These findings showed that there is a high degree of specificity of the chemical changes induced in both isolated collagen and bone, which govern the appearance of giant cells.

Since teeth, under most circumstances, do not readily resorb, it appeared of interest to study the tissue reaction to implanted molars and to relate the findings to resorption of bone implants. The present study was designed to utilize the subcutaneous environment (Irving and Handelman, '63) for the implantation of autogenous rat molars.

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TABLE 1

Number of animals	Treatment prior to implantation	Implantation periods
	Group 1 Devitalized molor implants	
56	Freeze-thaw	2, 7, 14, 21, 28, 56, 84, 112, 140 days
	Group 2 Decalcified molar implants	
63	Decalcification	Hourly intervals from 1-24 hours and 1, 2, 3, 4, 5, 6, 7, 14, 28, 56 days
8	Decalcification and FDNB treatment	14 days
24	Decalcification	4, 14, 35, 56 days followed by acid phosphatase determination.

RESULTS

ɩ.one of the implantation sites became ːptic, and most of the molars were ːcovered in highly vascular connective ssue capsules.

Reference molars. Reference sections f molars which were either devitalized or ecalcified, but not implanted, were judged o be nonvital by the torn and disrupted ıppearance of all pulpal cells when ·ompared with molars whose cells were ixed prior to decalcification.

Implanted molars. Histological examiıation of devitalized (Group I) and decalciied (Group II) molar implants revealed a narked difference in the tissue response.

Group I. The initial tissue response to he devitalized molar implants was the [illegible] of large numbers of inflammatory cells, primarily neutrophils and eosinophils, which surrounded each tooth, ınd invaded the pulp chambers through he root canals. On rare occasions, leukocytes penetrated and dentinal tubules as far as the predentin layer (fig. 1).

After 14 days the leukocytes decreased in numbers and were replaced with small, basophilic mononuclear cells, which also invaded the capsules and pulp cavities. They were also occasionally seen within tubules of the predentin layer. Unlike the leukocytes, the basophilic mononuclear cells were associated with mild resorption of predentin and dentin as evidenced by the scalloping of these usually smooth surfaces (fig. 2).

In most cases a decrease in cell numbers in the coronal pulp cavity was observed after 28 days, while multinucleated giant cells began to appear in significant numbers around fragmented or damaged roots. Little, if any, cellular activity was evident near intact roots or crown dentin surfaces at this stage. Deposition of an osteoid type of substance occurred in the pulp horns of some of the implants (fig. 3).

After 56 days moderate resorption of intact root surfaces by giant cells was evident (fig. 4).

Group II. The primary response to decalcified molar implants during the first week was very similar to that of the devitalized molar implants, but more invasive. After three hours of implantation, leukocytes surrounded the molars and invaded the pulp cavities by way of the root canals. Six hours later, these cells were observed penetrating deeply into the dentinal tubules. Penetration of the tubules was not restricted to the predentin layer (fig. 5).

After 14 days, the leukocytes decreased in numbers and were replaced with large numbers of small basophilic mononuclear cells which also invaded the pulp cavities. These cells aggregated on pulpal and external dentin surfaces (former dentoenamel junction) and penetrated deeply into the dentinal tubules (fig. 6). After penetration, the nuclei of the mononuclear cells became indistinguishable from the cytoplasm and the elongated cells

tion, and frozen. The antibiotic solution consisted of 100 μgm/ml streptomycin, 100 units/ml penicillin, and 100 μgm/ml mycostatin in 500 ml sterile distilled water (Goldhaber, '58). The antibiotic solution was used to eliminate bacterial contamination carried on the teeth from the oral cavity. Molars were devitalized by repeated slow freezing and thawing, which consisted of one hour at 1°C, four hours at −4°C, four hours at −60°C (dry ice-acetone bath) followed by a one hour thaw period at room temperature. This procedure was repeated four times. Destruction of vital tissue structures resulted from the formation of large ice crystals within the cells, and from the subsequent thaw. Following devitalization, teeth were implanted directly from the antibiotic solution.

Group II. Decalcified molar implants. After extraction, molars were placed in 5% EDTA solution pH 7.0 for decalcification. After one day of immersion, food debris and alveolar bone remnants were scraped from the teeth. They were then decalcified for a period of 14 days in frequently changed EDTA solution, and thoroughly rinsed for two days in refrigerated distilled water. Decalcification with EDTA also devitalized the teeth and destroyed residual oral bacteria. Decalcified reference molars which were not implanted, were prepared for histological section and stained with hematoxylin and eosin. The method of von Kossa was used to determine the completeness of decalcification.

Implantation and histological examination. Implantation of treated molars was accomplished under ether anesthesia. The shaved hindquarters of the donor rat were painted with dilute tincture of iodine and a small incision was made. The molar was placed aseptically under the skin.

Molars were implanted for periods ranging from one hour to 140 days. At the end of the appropriate period the animals were sacrificed with chloroform, the implants recovered in their connective tissue capsules and immediately fixed in neutral formal-saline solution for 24 hours. The devitalized molar implants were then decalcified in formic acid-sodium citrate solution, and prepared for histological section. The decalcified molar implants occasionally exhibited recalcification af[ter] implantation and required redecalcifi[ca]tion in formic acid-sodium citrate soluti[on] for 24 hours prior to sectioning. The maj[or]ity of decalcified molar implants, howe[ver,] were imbedded after fixation. Stai[ning] was done with hematoxylin and eosin.

A majority of decalcified molars w[ere] implanted without further treatment, a[nd] were processed afterward as previous[ly] described. Three other groups of decal[ci]fied molars were treated by different tec[h]niques which will now be explained.

Molars treated with 1-fluoro-2,4-dinitr[o]benzine (FDNB). In order to make hi[sto]logical comparisons with FDNB-treate[d] bone implants (Irving and Migliore, '65) a group of decalcified rat molars wer[e] treated with FDNB before implantatio[n] (Sanger, '45). Treatment was restricted t[o] decalcified molars since the reaction is no[t] complete in undecalcified specimens.

All FDNB-treated molars were imp[lanted] for 14 days, the period of maximum gia[nt] cell formation in bone implants. Af[ter re]covery, implants were fixed in neutra[l] formol-saline solution, prepared for histological section, and stained with hematoxylin and eosin.

A summary of the treatment of the tw[o] groups of rats is given in table [1].

Acid phosphatase determination. Molars in this group were extracted, decalcified, and implanted for periods of 4-[...] days. After recovery, they were fixed i[n] refrigerated Baker's formol-calcium solution pH 7.2 for 24 hours followed by at least 24 hours in a refrigerated solution of 0.88 M sucrose and 1% gum acacia (Holt, '59). Cryostat sections 8 μ thick were made at −20°C and air dried. The acid phosphatase method of Barka and Anderson ('62) was then used. Air dried cryostat sections were incubated in the test solution at 25°C for periods of time ranging from ten minutes to two hours. Sixty minutes at 25°C was considered to produce optimal color. Decalcified reference molars, boiled reference molars, and liver sections in solutions, with and without substrate, were processed simultaneously as controls.

ırptive activity of the giant cells in ars was considerably less than that erved in bone.

he invasion of decalcified and italized molar implants by basophilic nonuclear cells was of considerable ıificance, in that these cells were ociated with resorption of dentin and dentin. Similar cells were also observed various tissue implants of Irving and ndelman ('62) and Irving and Migliore 5), but were not related to resorption cause of the presence of multinucleated ınt cells and the irregular structure of implanted tissues. The virtual absence giant cells and the original smoothness d regularity of decalcified dentin and edentin surfaces allowed the formation resorption lacunae associated with the sophilic mononuclear cells to be clearly monstrated. Predentin resorption in calcified and devitalized molars was of terest in that this tissue has been garded as immune to resorption (Cabrini, aisto and Manfredi, '57).

Although resorption of dentin by ıononuclear cells was observed in all ıolars (except those treated with FDNB) it 'as most extensive in decalcified molars nd was often related to penetration of the entinal tubules by these cells. As with the receding leukocytes, penetration of the entinal tubules by the mononuclear cells ppeared to be an active process. This was uggested by the extension of long acuolated cytoplasmic processes that ften appeared to seek out dentinal ubules, which the processes entered efore the nuclei.

The mechanism of dentin resorption ould not be determined. However, extra-ellular enzyme secretion or phagocytosis vere given serious consideration in view of the presence of acid phosphatase in discrete "lysosome-like" cytoplasmic bodies in the basophilic mononuclear cells. Acid phosphatase is a hydrolase which has been associated with lysosomes by DeDuve ('59). According to Barka ('62), phagocytic, secretory, and absorptive cells display the greatest acid phosphatase activity. This enzyme is found in high concentrations in multinucleated giant cells (Cabrini et al., '62) and has been associated with bone resorption (Suzi, Goldhaber, and Jennings, '66). The presence of high concentrations of acid phosphatase, localized in cytoplasmic particles, would seem to be evidence of the resorptive capability of these cells.

The results of this study showed that the cellular responses to decalcified molars closely resembled that to implanted collagen (Irving and Migliore, '65) while the response to devitalized molar implants resembled that to bone implants. Although resorption of molars by mononuclear cells and multinucleated giant cells was significant, resorption of similar bone implants was more extensive and occurred more rapidly.

The cellular responses observed in this study closely resembled those reported by Langeland ('57) and Orban ('40) in studies of pulpal inflammation and dentin resorption in human teeth. For this reason the use of subcutaneous autogenous molar implants is suggested as a model for the experimental study of tooth resorption.

ACKNOWLEDGEMENTS

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appeared as long, thin basophilic lines in the dentinal tubules (fig. 7).

The basophilic mononuclear cells were associated with considerable resorption of predentin and dentin after 28 days of implantation. Those which remained on the surface were associated with surface resorption, while the penetrative cells were found in large "pockets" of dentin resorption (fig. 8). Giant cells were usually not observed in these 28 day molar implants.

After 56 days of implantation, all decalcified molars exhibited a "moth-eaten" appearance caused by coalescence of dentinal tubules by resorption associated with the mononuclear cells (fig. 9).

Infrequently a few multinucleated giant cells were associated with mild dentin resorption after 84–140 days of implantation.

FDNB treated decalcified molar implants. Large numbers of multinucleated giant cells were observed around 14 day FDNB-treated molar implants. Most of the giant cells aggregated around the outer root surfaces, where they appeared to engulf particles of cementum and periodontal remnants (fig. 10). Other giant cells were found in the pulp chambers, and occasionally within well-defined resorption lacunae in the coronal dentin (fig. 11). Penetrative mononuclear cells were rarely observed in the dentinal tubules and were markedly reduced in numbers in the pulp cavities and capsules.

Acid phosphatase determination. The leukocytes observed during the first week of implantation gave a moderately positive test for acid phosphatase. The penetrative mononuclear cells were, on the other hand, strongly positive, and the enzyme seemed to be concentrated within discrete bodies in the cytoplasm, possible lysosomes (fig. 12).

DISCUSSION

The appearance of numerous leukocytes in the primary response to all molar implants seemed to be a localized acute inflammatory reaction unrelated to the specific implanted tissue. A similar response was also stimulated by implanted autogenous bone (Irving and *Handelman*, '63), and FDNB-treated collagen (Irving and Migliore, '65). Infiltration of the pulp cavities and penetration of the [illegible] tubules by the leukocytes appeared to [illegible] active process. Orban ('40) called [illegible] to the ability of leukocytes to change sh[illegible] as they pass through capillary walls, [illegible] concluded, as do the present authors, th[illegible] these cells actively elongate and [illegible] the dentinal tubules. The fate of t[illegible] leukocytes was presumably degenerati[illegible] Although lytic enzymes may have be[illegible] released from these cells, there was [illegible] evidence of resultant dentin [illegible] after their disappearance.

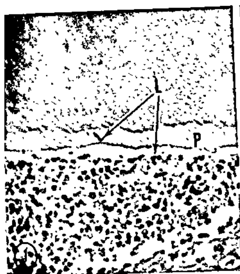
The secondary cellular response whi[illegible] appeared after 14 days also contained [illegible] "non-specific" component. The appearan[illegible] of basophilic mononuclear cells, fibrobla[illegible] and capillary networks was noted in [illegible] molar implants, as well as implants [illegible] bone (Irving and Handelman, '63) (Ir[illegible] and Migliore, '65) and collagen (Irving a[illegible] Migliore, '65).

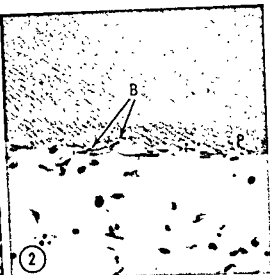
On the other hand, the appearance [illegible] giant cells in the secondary respon[illegible] seemed to be mediated by the nature of th[illegible] implanted tissue. *Giant cells* were rarel[illegible] seen around decalcified molar implant[illegible] whereas decalcified bone implants w[illegible] rapidly attacked by giant cells. Apparent[illegible] a giant cell-stimulating factor present [illegible] decalcified bone matrix is absent, [illegible] present in reduced amounts, in [illegible] dent[illegible] matrix.

As with bone implants, treatment wi[illegible] FDNB resulted in the appearance of lar[illegible] numbers of resorptive multinucleate[illegible] giant cells around decalcified molar[illegible] FDNB reacts with the epsilon amin[illegible] groups of lysine or hydroxylysine o[illegible] collagen. The presence of nitro groups [illegible] the para position seems to be the chemi[illegible] configuration responsible for giant ce[illegible] stimulation when this reagent comb[illegible] with collagen (Irving and Migliore, '65).

Unlike the decalcified molars, devitalized molars did eventually attract giant cells, but they were fewer in numbers and appeared more slowly. At first the giant cells were attracted only to fragmented roots, a "tropic effect" observed with the sharp ends of bone implants (Irving and Handelman, '63). After most dentin fragments were resorbed, giant cells were associated with gradual resorption of intact root and crown dentin (after 56 days). The

LANTATION OF RAT MOLARS
s T. Irving and James A. Bond









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PLATE 1

EXPLANATION OF FIGURES

1 Cellular response around devitalized molar, removed after seven days of implantation. Massive invasion of leukocytes (L) in the pulp chamber with minimal penetration of tubules of the predentin (P) layer. × 440.

2 Cellular response around devitalized molar, removed after 14 days of implantation. Basophilic, ovoid mononuclear cells (B) have replaced the leukocytes in the pulp chamber and are associated with resorption lacunae in the predentin (P). × 440.

3 Deposition of an "osteoid-like" material (a) in the pulp horn of a devitalized molar, removed after 28 days of implantation. Formative cell layer (b). × 270.

4 Area of resorption (R) of root dentin (D) in devitalized molar, removed after 56 days of implantation. Multinucleated giant cells (G). Connective tissue capsule (C). × 440.

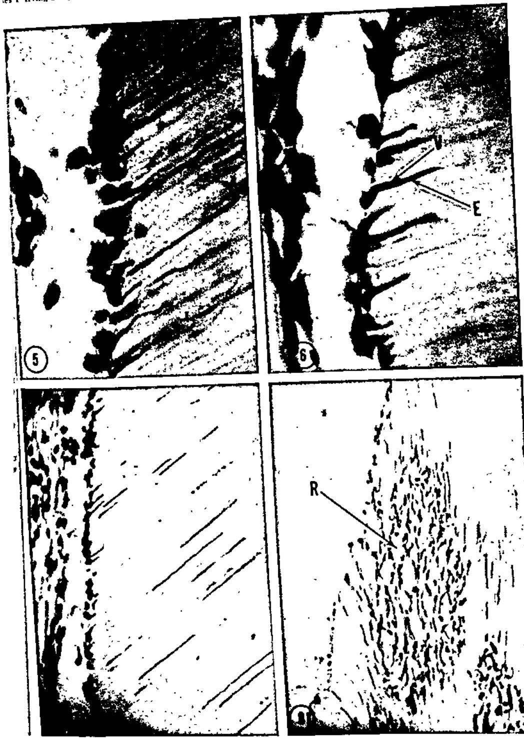

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PLATE 2

EXPLANATION OF FIGURES

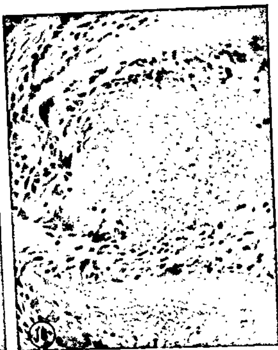
5 Aggregation of leukocytes along the pulpal dentin wall of a decalcified molar removed after 18 hours of implantation. Note the extensive cellular penetration of the dentinal tubules. ×970.

6 Penetration of dentinal tubules by basophilic mononuclear cells in decalcified molar removed after 14 days of implantation. Cytoplasmic extension (E) containing a vacuole (V). ×970.

7 Deep penetration of dentinal tubules by basophilic mononuclear cells in decalcified molar, removed after 14 days of implantation. ×440.

8 Centralized area of dentin resorption (R) in the cusp of a decalcified molar removed after 28 days of implantation. Note the presence of basophilic mononuclear cells in the resorption cavity and in the dentinal tubules. ×270.


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PLATE 3

EXPLANATION OF FIGURES

9 Cross-section view of dentinal tubules in decalcified molar removed after 56 days of implantation. "Moth-eaten" appearance produced by resorption of dentin between adjacent dentinal tubules containing basophilic mononuclear cells. × 440.

10 Aggregation of numerous multinucleated giant cells around the root apex of an FDNB treated decalcified molar removed after 14 days of implantation. × 440.

11 Cellular response in pulp chamber of FDNB treated decalcified molar removed after 14 days of implantation. Note the presence of large numbers of multinucleated giant cells and area of dentin resorption (R). × 270.

12 Positive acid phosphatase reaction (Barka and Anderson, '62) in two basophilic mononuclear cells. Cryostat section of decalcified molar removed after 14 days of implantation. Numerous acid phosphatase positive "lysosome-like" bodies are present within the cytoplasmic extensions in the dentinal tubules. × 970.


R
11
12

›velopment of the Gonads in the Female ›panese Quail[1]

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ABSTRACT The germ cells in the Japanese quail, appear in the gonadal area between two to two and one half days incubation with a preferential distribution favoring the left gonad. Proliferation of the primary or medullary cords from the germinal epithelium was observed between four and five and one half days. The secondary or cortical cords were found to proliferate in the left gonad on the sixth day. The right gonad revealed neither cortical cords nor cortex in any of the cases studied. It began to involute around eighth day and reached its minimum size by seven to ten days after hatching. In the adult it was composed of loose, or occasionally cord-like, epithelial cells, sparsely scattered "fat laden" cells, masses of lymphocytes and fibrous connective tissue, the latter predominating in older stages. Germ cells either disappeared or could no longer be recognized in the right gonad by four days after hatching. In the left ovary oogonia began to show meiotic changes by the eleventh day and primary follicles were present on the day of hatching. Ovulation was observed as early as 45 days after hatching. In female embryos the right Mullerian duct underwent atrophy and became a rudimentary cloacal remnant between eight and 14 days. In males complete atrophy of both left and right Mullerian ducts occurred between seven and 13 days. The mesonephric ducts persisted in females and were demonstrable throughout post-hatch period as thread-like structures.

›lthough the Japanese quail, *Coturnix* ›urnix *japonica* Temminck and Schle- ›, was first introduced into North Amer- › for the purpose of providing game, the ›empts both earlier (Philips, '28) and ›re recently (Cottam and Stanford, '59) ›ve apparently failed to establish it as a ›ccessful fauna in this country. However, Padgett and Ivey ('59) pointed out, ›turnix has become an ideal laboratory ›imal because of its hardiness, ease of ›ndling, precociousness and great laying ›pacity. Our principal interest in this bird ›us far has been to study the effects of ›nistral ovariectomy with particular refer- ›ce to its effects on the rudimentary right ›nad. In order to interpret the changes ›served following sinistral ovariectomy, it ›came necessary to know something of ›e embryonic development of the right ›nad in the female and its subsequent ›volution. Since we found no pertinent ›terature on the subject, we decided to ›vestigate this problem.

MATERIALS AND METHODS

Quail eggs[3] were incubated at 100°F and he embryos sacrificed at one half day ntervals throughout the incubation period ›f 16 to 17 days. A number of specimens for each age group were examined grossly in order to gain some information as to the variability in normal development. The typical embryos of each group were then compared with the stages reported by Padgett and Ivey ('60). Postembryonic birds were sacrificed at one day intervals for the first ten days, at two day intervals for the next ten, then at five day intervals up to maturity (45 to 60 days) and at random thereafter up to three and one half years. The number of specimens studied for each stage is shown in table 1.

Specimens were fixed in Bouin's and Zenker's fluids and in Bensley's acetic-osmic-bichromate mixture which was

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[2] The study was carried out in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Anatomy, Loyola University Stritch School of Medicine, during the tenure of a Graduate Student Teaching Assistantship. The senior author subsequently was supported by a Medical Student Research Fellowship in Anatomy and currently is a Research Associate in the Department of Anatomy, Loyola University Stritch School of Medicine.

[3] The initial stock of eggs and birds were purchased from the Antelope Valley Farm, Lincoln, Nebraska. Gifts of young stock were also received from Dr. A. V. Nalbandov, Department of Animal Physiology, University of Illinois, Urbana, Dr. R. K. Meyer, Department of Zoology, University of Wisconsin, Madison, and young stock and fertile eggs through the courtesy of Dr. J. R. Howes, Department of Poultry Science, Auburn University, Auburn, Alabama. These gifts are gratefully acknowledged. The birds are now reared in our own laboratory

elongated, narrow, light-colored bodies the medial-ventral side of the mesophric ridges extending from behind the phalic end of the Wolffian body to about mm caudally. They are composed of a rminal epithelium of two to three layers d an underlying tissue of loosely ranged mesenchymal cells (fig. 2). A ateral asymmetry as concerns the numr of germ cells and the size of the gonads as already evident at this time. Among e ten embryos examined, seven showed a mewhat larger left (30 to 60 μ in thickss) than right (25 to 55 μ) gonad and the rm cells were distributed in approxiately a 5:2 ratio between left and right. the remaining embryos both gonads ere approximately the same size, wever, distribution of the germ cells owed a 4:3 ratio in favor of the left nad. By the fourth day of incubation d-like projections from the germinal epielium were noticeable, indicating the ginning proliferation of the first or imary set of sex cords. This proliferation as active by four and one half days and ad practically terminated by five and one alf days (fig. 3). The mesenchyme cells ppear to distribute themselves between he cords, to form the connective tissue troma rather than contributing to the formation of the cords themselves. The cords onsist mainly of epithelial cells with a few erm cells embedded in them. When proliferation is complete, a layer of connective issue stroma appears under the epithelium representing the primary tunica lbuginea.

In general, the PGCs up to this time (5 to days) have retained their original size nd shape. A few of them, however, are omewhat smaller and this is apparently he result of mitotic division. One noticeble change at this time is the disappearance of the cytoplasmic fat globules. The size of the germ cells, however, remains he most significant identifying characteristic during this period.

Sex differentiation and development of gonads to time of hatching. The period of development from sex differentiation to the time of hatching may be arbitrarily divided, for convenience of description, into two stages: (1) from six to nine days of incubation when most of the cortical cords are formed and (2) from ten days to hatching when differentiation of the germ cells and involution of the right gonad in females appear to be the most important events.

From six to nine days of incubation. Proliferation of the primary or medullary cords is terminated by five and one half to six days, at which time a definite determination of sex can be made. In addition to the fact that the left gonad in females is larger than the right, the germinal epithelium of the left is also much thicker (3 to 5 cells or 25 to 30 μ). In the right gonad on the other hand, the epithelium has diminished to little more than a single layer of cells (fig. 4). In male embryos, as the first proliferation comes to an end, both right and left gonads acquire an epithelium similar to that of the right in females, i.e., a single layer of cuboidal cells.

Sex differentiation is further evident when the cortical cords, or cords of second proliferation begin to appear as bud-like projections from the epithelium of the left gonad of females starting on the sixth day of incubation. This proliferation becomes very pronounced during the succeeding days, especially between seven and nine days. Although a sporadic division of germ cells was noticed in earlier stages, it is during the height of cortical proliferation that the active phase of mitotic division of these cells was seen. The resulting daughter cells are smaller and show no trace of cytoplasmic fat depositions. Unlike the medullary cords, the cortical cords consist mainly of germ cells and a few somatic cells derived from the germinal epithelium.

In none of the 25 embryos examined between six and nine days of incubation did we find any trace of a cortex or proliferation of cortical cords in the rudimentary right gonad. As mentioned earlier, the epithelium of this gonad is already a thin, single layered, structure by six days of incubation.

The medulla of both left and right gonads in females continued to increase in size during the period of cortical proliferation. At seven days of incubation the medulla of the left had reached a thickness of 170 to 190 and the right 130 to 150 μ.

TABLE 1

Specimens employed in microscopic study of gonads[1]

Embryonic period		Postembryonic period	
Age of embryos in days	Number of specimens examined	Age of birds in days	Number of specimens examined
0-1	4	1-2	2
1-2	6	2-3	4
2-3	12	3-4	4
3-4	10	4-5	3
4-5	10	5-6	1
5-6	9	6-7	2
6-7	8	7-8	2
7-8	8	8-9	2
8-9	9	9-10	4
9-10	5	10-12	3
10-11	2	12-14	2
11-12	6	14-16	4
12-13	2	16-18	3
13-14	7	18-20	2
14-15	3	20-25	6
15-16	2	25-30	2
16-17[2]	5	30-35	3
		35-40	1
		40-50	5
		50-60	3
		60-70	2
		70-80	3
		80-3.5 yr.	10

[1] All specimens studied following sex differentiation on sixth day were females
[2] Day of hatching.

found to be a good fixative for embryos of early stages since it satisfactorily preserved cytoplasmic yolk inclusions in the primordial germ cells. The material was sectioned at 4 to 6 μ and stained in hematoxylin and eosin, iron hematoxylin or Mallory's triple (McFarlane, '44) stain. Specimens fixed in the acetic-osmic-bichromate mixture were stained employing Bensley's anilin-acid fuchsin-Wright's blood stain method (Swift, '15).

An attempt was made to distinguish the germ cells of early quail embryos by fixing them in chilled Gendre fluid and staining for glycogen by the periodic acid-Schiff (PAS) technic and in cold acetone for staining with alkaline phosphatase employing Gomori's ('46) and Burstone's ('58) modifications.

OBSERVATIONS

Development of the undifferentiated gonad. The formation of the gonads may conveniently be considered as beginning with the arrival of the primordial germ cells (PGCs) in the peritoneal epithelium midway between the cranial aspect of the mesonephric ridge and the developing dorsal mesentery. In the Japanese quail this happens at about 48 to 60 hours incubation. Prior to this stage, the germ cells can be seen in the entoderm of the blastoderm, space between ectoderm and mesoderm, blood islets (fig. 1), and the splanchnopleur, revealing the extragonadal origin of these cells.

The PGCs of the quail resemble those described by Swift ('14) in the chick, in size (14 to 16 μ), shape (spherical to oval), prominent nuclei (7 to 10 μ) and above all in a cytoplasm filled with fat globules (fig. 1). When stained for glycogen with the PAS reagent, these cells did not show any significant reaction. It is interesting in this connection that Meyer ('61), Domm and Clawson ('62) and Clawson and Domm ('63) observed that glycogen can be used as a diagnostic characteristic of the early germ cells in the chick. Attempts to trace these cells with alkaline phosphatase stain also failed in this species although it has been found to be highly successful in the case of mouse embryos (Chiquoine, '54; Mintz, '59) and others.

By three to three and one half days of incubation, the genital ridges were visible

:longated, narrow, light-colored bodies the medial-ventral side of the meso-hric ridges extending from behind the halic end of the Wolffian body to about mm caudally. They are composed of a minal epithelium of two to three layers : an underlying tissue of loosely anged mesenchymal cells (fig. 2). A iteral asymmetry as concerns the num- of germ cells and the size of the gonads s already evident at this time. Among ten embryos examined, seven showed a newhat larger left (30 to 60 μ in thick-ss) than right (25 to 55 μ) gonad and the m cells were distributed in approx-ately a 5:2 ratio between left and right. the remaining embryos both gonads re approximately the same size, wever, distribution of the germ cells owed a 4:3 ratio in favor of the left nad. By the fourth day of incubation d-like projections from the germinal epi-elium were noticeable, indicating the ginning proliferation of the first or imary set of sex cords. This proliferation as active by four and one half days and id practically terminated by five and one lf days (fig. 3). The mesenchyme cells pear to distribute themselves between e cords, to form the connective tissue roma rather than contributing to the for-lation of the cords themselves. The cords onsist mainly of epithelial cells with a few erm cells embedded in them. When prolif-ration is complete, a layer of connective ssue stroma appears under the epithe-um representing the primary tunica lbuginea.

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The medulla of both left and right gonads in females continued to increase in size during the period of cortical proliferation. At seven days of incubation the medulla of the left had reached a thickness of 170 to 190 and the right 130 to 150 μ.

This increase is mainly due to the mitotic activity of the cells of the medullary cords. By seven and one half to eight days, the medullary cords of both gonads in female embryos began to show certain atrophic changes. The first indication of this was visible when some of the cords, especially those towards the hilar area, acquired a lumen (fig. 5). These cords gradually became distended with the lining cells becoming cuboidal and later somewhat more flattened. Some of the cords became loosely arranged and broke up into small isolated groups of cells. Some of these cells underwent cytological changes becoming vesicular and vacuolated, resembling the "fat laden" cells described in the chick embryo by Brode ('28), while others underwent necrosis and gradually disappeared.

From ten days of incubation to hatching. The left ovary. By the tenth day, cortical proliferation may cease in the medial ventral areas of the ovary where the cortical cords remain detached and separated from the epithelium by a discontinuous connective tissue stroma which represents the developing secondary or ovarian tunica albuginea. The cessation of proliferation of cortical cords gradually spreads in all directions from the medial ventral region of the ovary, so that at the time of hatching, proliferation of these cords may be seen only in the dorsal lips or folds of the cortex close to the mesovarium.

By the eleventh day, some of the oogonia at the inner aspect of the cortex revealed the early meiotic changes. Their nuclei enlarged and began to show deeply staining chromosomes similar to those seen in typical leptotene stages of meiosis. Furthermore, the chromosomes became polarized, that is to say they were not randomly distributed but lay clumped to one side with a clear space at the opposite pole of the nucleus, thus resembling the synizesis arrangement (fig. 6). This transformation continued and spread over the entire cortex as incubation advanced and hatching time approached. Meanwhile, by 13 to 14 days, the older oocytes, located at the inner aspect of the cortex, underwent further meiotic prophase changes in anticipation of the growth phase. All of the oocytes appeared to enter this growth phase but only a fraction of them continued to grow and acquire a follicular envelope. Th[illegible] underwent involution and [illegible] beginning at 14 days of incubation.

During the period from 10 to 16 days, [illegible] medulla of the left ovary also rev[illegible] some growth. This growth was lar[illegible] confined to the outer sub-cortical [illegible] the medulla and in consequence this [illegible] became very compact and sharply di[illegible]guished from the inner vacuo[illegible] lar zone. The "fat laden" cells are fa[illegible] prevalent in the medulla at this stage ([illegible] 6). While the oogonia of the cortex w[illegible] undergoing meiotic changes, some of [illegible] germ cells located in the medulla show[illegible] similar changes resulting in the syniz[illegible] arrangement of their [illegible] However, these changes in the latter [illegible] not proceed much further, instead, th[illegible] underwent necrosis and gradually dis[illegible]peared from the medulla.

The rudimentary right gonad. T[illegible] rudimentary right gonad of the fem[illegible] ceased to grow between eight and n[illegible] days of incubation. In fact it began to sh[illegible] signs of involution by the eighth day, a[illegible] at the time of hatching it measur[illegible] approximately 1.5 to 2 mm in length a[illegible] 0.5 mm in width while the left ovary h[illegible] an average length of 4 mm and a width [illegible] 1.5 mm. This disparity in size between th[illegible] right and left female gonads was m[illegible] evident in their thickness, the right ru[illegible]ment ranging from 45 to 60 μ as compar[illegible] to 220 to 300 for the left ovary (fig. 8). [illegible] germinal epithelium at this stage is so th[illegible] that it is barely distinguishable from th[illegible] visceral peritoneum. In none of the [illegible] embryos examined, between ten days a[illegible] the time of hatching, did we observe cor[illegible]cal cords or a cortex in the right rud[illegible]tary gonad, nor were any germ cells seen [illegible] the germinal epithelium.

The atrophic changes seen in th[illegible] medulla of the right gonad, beginning [illegible] eight to nine days of incubation, continu[illegible] at an accelerated rate during the rest of t[illegible] incubation period. The lacunae of the l[illegible]nated medullary cords gradually be[illegible] larger and in consequence the lining [illegible]thelium became thinner (fig. 7). Conv[illegible]sion of medullary cells into "fat l[illegible] cells continued but as the embryos [illegible] older, this transformation decreased.

The number of germ cells in the medu[illegible]

the right rudimentary gonad also [de]creased as development advanced. At [the] time of hatching, a cross section of this [gon]ad might reveal one or two and occa[sion]ally none at all. None of the germ cells [obs]erved showed meiotic changes such as [tho]se seen in the left ovary.

Development of gonads in the post-[hat]ch period. The left ovary. A detailed [dis]cussion on the growth and development [of] the left ovary during the post-hatch [per]iod is not intended, hence only certain [rel]evant changes will be noted.

The mitotic division of germ cells seems [to] have ceased in all areas of the cortex by [th]e day after hatching. Even as early as the [da]y of hatching, the growing oocyte may [be] surrounded by a follicular covering (fig. [..]). This was particularly so in embryos [th]at required 17 or more days to hatch. As [no]ted earlier, not all oocytes acquire a fol[lic]ular envelope; only a small percentage [di]d so while those that did not, disap[pe]ared. The formation of ovarian follicles, [as] well as the development of oocytes, was [fir]st seen along the medial ventral area of [th]e ovary and then spread to other areas. [By] the eighth day after hatching, follicles [co]uld be seen in all areas of the ovarian [co]rtex. The older follicles attained full [gr]owth and ruptured between 45 and 75 [da]ys.

Soon after hatching, an increase in num[be]r was noticed in the epithelial cells of [th]e cortex of the left ovary some of which [di]fferentiated into "interstitial" cells. These [ce]lls were especially abundant in the [th]ecal layer of growing follicles.

A medulla is recognizable in the left [o]vary of the adult. The medullary cells of [th]e outer compact zone intermingle with [th]e interstitial elements of the cortex and [th]us lose their identity in the post-hatch [o]vary. The reticular zone of the medulla in [th]e adult ovary is confined to the hilus and [c]onsists mostly of connective tissue infil[t]rated with blood vessels. Loose epithelial [c]ells were seen in the early post-hatch [s]tages but in mature ovaries, they have [e]ither disappeared or have become [s]o modified in such a way that they are no [l]onger distinguishable from other connec[ti]ve tissue cells. The "fat laden" cells of the [m]edulla gradually disappear in the early [p]ost-hatch period, so that in ovaries from birds beyond ten days of age these cells are seldom seen.

Germ cells were as a rule, not seen in the medulla of the left ovary after hatching. However, in one four day old bird a few were found in some of the sections. Scattered cells with fairly large nuclei may be seen in the medulla of adult ovaries, however, their identity as germ cells is doubtful.

The rudimentary right gonad. The rudimentary right gonad of the female continued to regress for some days after hatching and attained its smallest size sometime between seven and ten days. At this time it is visible as an elongated, narrow, flattened bit of tissue lying on the ventral surface of the post caval vein and/or the medial surface of the right mesonephric remnant. A wide individual variation was observed in the size of these gonads which generally ranged from 1 to 2 mm in length and 0.2 to 0.6 in width.

In none of the 53 right gonads examined histologically, from hatching to sexual maturity, was there any evidence of a cortex. The epithelium covering the gonad was invariably thin, flattened and similar to the peritoneal lining of the coelom (figs. 9, 11, 12).

The medulla of the right gonad showed some variations as the birds approached sexual maturity. Usually, it was composed of loose masses of epithelial cells derived from the original medullary cords, a few "fat laden" cells, connective tissue stroma and some blood cells (fig. 11). In a few cases small solid cords of epithelial cells were seen. Distended luminated cords and masses of lymphocytes were also observed in the stroma. Fibrous connective tissue was increasingly prevalent in the older gonads. In a few the bulk of the gonad was composed of fibroblast-like cells with very few epithelial cells in evidence. In such cases, it seems that the epithelial cells have either largely disappeared or have transformed into fibrous connective tissue-like cells. The number of "fat laden" cells also showed considerable individual variation while in some cases none of these cells could be found.

Germ cells were seen in the right gonad as late as three to four days posthatch after which they could no longer be found

This increase is mainly due to the mitotic activity of the cells of the medullary cords. By seven and one half to eight days, the medullary cords of both gonads in female embryos began to show certain atrophic changes. The first indication of this was visible when some of the cords, especially those towards the hilar area, acquired a lumen (fig. 5). These cords gradually became distended with the lining cells becoming cuboidal and later somewhat more flattened. Some of the cords became loosely arranged and broke up into small isolated groups of cells. Some of these cells underwent cytological changes becoming vesicular and vacuolated, resembling the "fat laden" cells described in the chick embryo by Brode ('28), while others underwent necrosis and gradually disappeared.

From ten days of incubation to hatching. The left ovary. By the tenth day, cortical proliferation may cease in the medial ventral areas of the ovary where the cortical cords remain detached and separated from the epithelium by a discontinuous connective tissue stroma which represents the developing secondary or ovarian tunica albuginea. The cessation of proliferation of cortical cords gradually spreads in all directions from the medial ventral region of the ovary, so that at the time of hatching, proliferation of these cords may be seen only in the dorsal lips or folds of the cortex close to the mesovarium.

By the eleventh day, some of the oogonia at the inner aspect of the cortex revealed the early meiotic changes. Their nuclei enlarged and began to show deeply staining chromosomes similar to those seen in typical leptotene stages of meiosis. Furthermore, the chromosomes became polarized, that is to say they were not randomly distributed but lay clumped to one side with a clear space at the opposite pole of the nucleus, thus resembling the synizesis arrangement (fig. 6). This transformation continued and spread over the entire cortex as incubation advanced and hatching time approached. Meanwhile, by 13 to 14 days, the older oocytes, located at the inner aspect of the cortex, underwent further meiotic prophase changes in anticipation of the growth phase. All of the oocytes appeared to enter this growth phase but only a fraction of them continued to grow and acquire a follicular envelope. T[illegible] underwent involution and [illegible] beginning at 14 days of incubation.

During the period from 10 to 16 days [illegible] medulla of the left ovary also rev[illegible] some growth. This growth was [illegible] confined to the outer sub-cortical are[illegible] the medulla and in consequence th[illegible] became very compact and sharply di[illegible]guished from the inner vacuolated [illegible]lar zone. The "fat laden" cells are [illegible] prevalent in the medulla at this stage [illegible] 6). While the oogonia of the cortex w[illegible] undergoing meiotic changes, some of [illegible] germ cells located in the medulla show[illegible] similar changes resulting in the synize[illegible] arrangement of their ch[illegible] However, these changes in the latter [illegible] not proceed much further, instead, t[illegible] underwent necrosis and gradually dis[illegible]peared from the medulla.

The rudimentary right gonad. T[illegible] rudimentary right gonad of the fem[illegible] ceased to grow between eight and n[illegible] days of incubation. In fact it began to sh[illegible] signs of involution by the eighth day, a[illegible] at the time of hatching it meas[illegible] approximately 1.5 to 2 mm in length a[illegible] 0.5 mm in width while the left ovary h[illegible] an average length of 4 mm and a width [illegible] 1.5 mm. This disparity in size between t[illegible] right and left female gonads was m[illegible] evident in their thickness, the right ru[illegible]ment ranging from 45 to 60 μ as compar[illegible] to 220 to 300 for the left ovary (fig. 8). T[illegible] germinal epithelium at this stage is so t[illegible] that it is barely distinguishable from t[illegible] visceral peritoneum. In none of the [illegible] embryos examined, between ten days a[illegible] the time of hatching, did we observe [illegible]cal cords or a cortex in the right rudime[illegible]tary gonad, nor were any germ cells seen [illegible] the germinal epithelium.

The atrophic changes seen in [illegible] medulla of the right gonad, beginning [illegible] eight to nine days of incubation, contin[illegible] at an accelerated rate during the rest of [illegible] incubation period. The lacunae of the lu[illegible]nated medullary cords gradually beca[illegible] larger and in consequence the lining [illegible]thelium became thinner (fig. 7). [illegible]sion of medullary cells into "fat [illegible] cells continued but as the embryos [illegible] older, this transformation decreased.

The number of germ cells in the medul[illegible]

r and cortical cords as well as in other or events of later development.

, in the chick, an extragonadal origin of PGCs became apparent from our les in the quail, for these cells were id in the entoderm of the blastoderm, space between the ectoderm and meso-n, in the blood stream, and in the nchnopleur prior to their arrival in the adal area at two to two and one half s of incubation. Firket ('14), although eeing with the concept of an extra-adal origin of the germ cells in the k, does not believe that the definitive ytes arise from these cells. Instead, he eves that these cells undergo degenera- beginning at 15 days of incubation and t they are subsequently replaced by new cells arising from the germinal epithe-n. Such a view was previously reported Dantschakoff ('08) and more recently by enberg and Garwacki ('38). Swift ('15) l Goldsmith ('28) on the other hand, intain that it is the PGCs that later give to the definitive oocytes. In the quail were able to follow the course of the Cs from the time of their arrival in the nad, through mitotic divisions, and later, ough the transformation of oogonia into cytes, some of which became enveloped follicles. However, we observed that in quail many of the PGCs undergo generation and disappear, beginning at days of incubation, but it is the surviv- that develop into the definitive oocytes. Firket ('13, '14), Swift ('15), Blocker 3), Stanley and Witsch ('40), Clawson d Domm ('63) and others, have observed at in chick embryos the PGCs collect eferentially in the left gonad, sometimes two to five times greater numbers than the right, as early as 96 hours of incuba-n. Prior to two and one half to three days e thickness of the germinal epithelium d the number of incorporated germ cells ere more or less equal on left and right des. A similar phenomenon was observed our quail embryos as early as three days incubation. The germinal epithelium of e left gonad was thicker than that of the ght and the germ cells showed a 5:2 ratio favor of the left. However, this disparity as not so pronounced in any of the youn-er embryos examined. Some of these howed a near uniformity in the size of left and right gonads and in these the ratio was approximately 4:3. As is now generally known, in the female domestic fowl and the quail, as well as most other birds, only the left ovary becomes functional while the right ceases to develop at an early age and persists throughout life as a rudimentary gonad. In the male on the other hand, both left and right gonads become functional. In general, we may therefore assume that those embryos which show a pronounced asymmetry of the gonads are genetically determined females. However, a more extensive study on the distribution of the germ cells in the quail is necessary before a fixed ratio can be established.

Contrary to the prevailing view that in the domestic fowl, the medullary cords proliferate from the germinal epithelium, (Hoffman, 1892, D'Hollander, '04, Firket, '14, Swift, '15, '16, Benoit, '24, '51, Goldsmith, '28, Brode, '28, Essenberg and Garwacki, '38, Willier, '39, and others), Witschi ('35, '60) reports that in birds these cords are derived from the mesenchymal blastema of the indifferent gonad. Blocker ('33) in his studies on the English sparrow *Passer domesticus* earlier came to the same conclusion.

Bud-like projections were seen to arise from the germinal epithelium of quail embryos of four to four and one half days of incubation which by further proliferation appear to give rise to the medullary cords. Our study did not permit us to determine whether any cells of the mesenchymal blastema contributed to the formation of the medullary cords.

From the studies of Domm ('27), Brode ('28) and Willier ('39) it is known that the medullary tissue does not completely disappear in the female chick although Firket ('20) believed that it does. Brode and Willier both reported that it persists in the left ovary as well as in the rudimentary right gonad after transforming into one of the following structures: distended tubules which may temporarily contain germ cells, clusters of cords of clear or "fat laden" cells and isolated medullary cord cells. In the quail we found a similar transformation of the medulla of the left and right gonads. The difference between the domestic fowl and the quail seems to lie chiefly in the degree of atrophy that occurs and

although occasional large nuclei lacking distinct cell walls were seen in the medulla. The identity of the latter as germ cells was questionable.

The rudimentary right gonad in older birds. The right gonad sites of ten older females ranging in age from 230 days to three and one half years were studied histologically. A rudimentary gonad was found in all of these cases (fig. 12). The epithelial cells were fewer in number but there was more fibrous connective tissue when compared with the younger stages. The so-called "fat laden" cells were rarely observed in any of these gonads but masses of lymphocytes were frequently seen.

The development of Mullerian and Wolffian ducts. Mullerian ducts. The tubal ridges, that later contribute to the formation of the Mullerian ducts were usually visible during the third day of incubation as thickened lines of peritoneum along the dorso-lateral side of the mesonephros, lying immediately lateral to the mesonephric duct. By three and one half days, the cranial end of each ridge began to invaginate and to form a groove which soon transformed into a tube, the Mullerian duct. The tube was open at the cranial end but closed caudally. The part thus formed was relatively short. The major portion of the duct was formed by a posterior elongation which reached the cloaca at about six to six and one half days incubation.

In female embryos, development of the Mullerian ducts was uniform on either side up to eight days at which time both ducts showed a length of 6 to 6.5 mm. At this time the right Mullerian duct ceased to grow and showed signs of atrophy. This began at the cranial end and by the fourteenth day of incubation the tube had disappeared except for a short cloacal stump measuring about 2.5 to 3 mm in length. In adult females this rudimentary cloacal remnant was found attached to the right side of the cloaca as reported in the brown Leghorn fowl (Domm, '27). It was found to be somewhat larger in laying than in non-laying females presumably because of its responsiveness to estrogenic hormones.

The left oviduct on the other hand continued to grow throughout the entire embryonic as well as early postembryonic period. In the period between 35 to 45 days, a tremendous increase in size and coiling took place in preparation for ovulation and laying. A noticeable increase in vascularization was also observed at this period. At the time of ovulation the oviduct was large, approximately 15 to 30 cm in length, highly convoluted, and took up most of the space in the body cavity between the ovary and the cloaca.

It is of interest to note in this connection that in males regression of the Mullerian ducts occurred simultaneously in both left and right ducts and that it began ... and one half days of incubation. Unlike the female, atrophy of these ducts in the male begins at the caudal end and progresses cranially, rather rapidly, since by the 11th day they have practically disappeared except for a small thread-like ... which had usually disappeared by the ...teenth day.

Wolffian ducts. The mesonephric duct persists in the adult female as a minute straight, thread-like ridge, lying in close association with and parallel to, the ureter on the ventral surface of the metanephros. Anteriorly, it is connected to the mesonephric remnant, consisting of a few convoluted tubules, the epoophoron, and posteriorly to the cloaca.

DISCUSSION

In comparing the development of the gonads of the quail, and those of the chick which is well known to students of avian embryology, it is clear that, despite the difference in incubation periods viz., 16–17 days for the quail and 21 for the chick, the degree of development of the gonads at the time of hatching is about the same. Consequently, embryonic development requiring 21 days for the chick is completed in a shorter period in the quail. Hence, there is evidently a speeding up of development in the case of the quail. For example, in the chick, Swift ('14, '15) and others have found that the PGCs appear in the presumptive germinal epithelium by three and one half days of incubation whereas in the quail we found that this occurred as early as two to two and one half days. Such an acceleration was also observed in the proliferation of the medul...

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consequently in the amount of medullary tissue that persists, especially in the right gonad. In the chicken, cords of clear or unmodified medullary cells and a considerable number of "fat laden" cells are quite common in the rudimentary right gonad of adults. In the quail on the other hand, medullary cells, aggregated in a cord-like fashion, were observed only in exceptional cases; they usually occurred as scattered isolated cells. The "fat laden" cells were relatively rare and sometimes even completely absent. Our observations indicate that atrophy of the right gonad is more severe in the quail than in the chick which would seem to be supported by the earlier disappearance of the germ cells from the rudimentary right gonad. In the chick, Brode ('28) observed germ cells in the medulla of both the left ovary and the right rudimentary gonad up to three to four weeks after hatching while in our studies on the quail these cells were either absent or could no longer be recognized beyond four days after hatching.

One of the important and interesting observations on the rudimentary right gonad of the quail was the absence of cortical tissue. According to Brode ('28) in the domestic fowl, 39% of the embryonic right gonads and 20% of the adult revealed either a cortex or cortical cords. The presence of cortical tissue has also been reported in the developing right gonad of the English sparrow (Blocker, '33) and the Pekin duck (Lewis, '46). The absence of this tissue in the right gonad of the quail is of particular importance in studies on the hypertrophied rudimentary right gonad following sinistral ovariectomy. Finlay ('25) and Domm ('27) postulated that the rudimentary right gonad of the brown Leghorn fowl may develop into an ovotestis or an ovary only when cortical elements are present in the rudimentary right gonad at the time of ovariectomy, and Domm (loc. cit.) maintained that the presence of a normal sized right ovary in poulards was contingent on the amount of cortical tissue present in the right gonad at the time the left ovary was removed. Our observations on the hypertrophied rudimentary right gonad of the quail are in agreement with this view, namely, that cortex will not develop, *de novo*, since in a total of 50 ovariectomies (Kannankeril and Dom[illegible] '63, '64) not a single case showed ovar[illegible] tissue in this gonad. If the hypothesis [illegible] Finlay and Domm (loc. cit.) is valid [illegible] this result would be expected since [illegible] cortical elements were found in any of [illegible] quail embryos studied. The reason for [illegible] apparent complete absence of cortex in [illegible] rudimentary right gonads of the quail [illegible] contrast to the chick and some [illegible] is not known. We may tentatively concl[illegible] that in the quail, the genetic and/or h[illegible] monal inhibitory control of cortical dev[illegible] opment in the right gonad is compl[illegible] whereas in those forms where [illegible] occurs this control is either incomplet[illegible] lacking.

ACKNOWLEDGEMENTS

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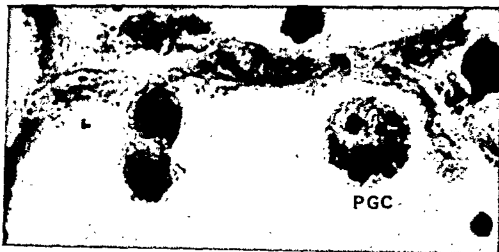
PLATE 1

EXPLANATION OF FIGURES

1 Photomicrograph of a primordial germ cell (PGC) located in a blood islet of an embryo of two and one half days incubation. Note the large size of the nucleus and the lipid granules in the cytoplasm. × 1500.

2 Photomicrograph of the left (L) and right (R) gonads of a three and one half day embryo. Note the difference in distribution of germ cells between left and right gonads. × 600.

Cross section of the left gonad of a four and one half day embryo showing the ingrowth from the germinal epithelium of the primary or medullary sex cords (MC). × 600.


PGC


L
R

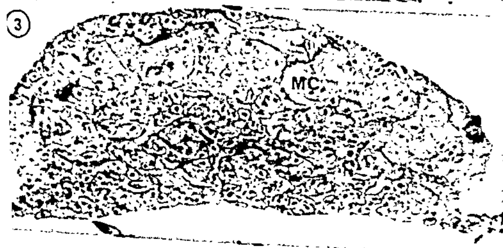

3
MC

PLATE 2

EXPLANATION OF FIGURES

4 Cross section through the peripheral portion of the left and right gonads of a six day female embryo. Note the thin, single layered, germinal epithelium (GE) of the right gonad and the thick multilayered epithelium of the left gonad. The latter also shows small bud-like projections indicating the beginning of proliferation of the secondary or cortical cords (CC). MC, medullary cords. × 530.

5 Cross section of the rudimentary right gonad of an eight day female embryo. Note the absence of any cortex or cortical cords, also the loosely arranged medullary cords and the lacunae (L) in certain of the cords. × 260.

6 Cross section of the left ovary of an 11 day female embryo showing a portion of the cortex and the underlying medullary tissue. Note the oocytes in synizesis (O) at the inner aspect of the cortex and islets of "fat laden" cells (FLC) in the medulla. × 650.

7 Cross section of the rudimentary right gonad of an 11 day female embryo. Note the lone germ cell (GC) in the degenerating medullary tissue and the "fat laden" cells (FLC). × 475.

DAL DEVELOPMENT IN FEMALE JAPANESE QUAIL
annankeril and L V Domm

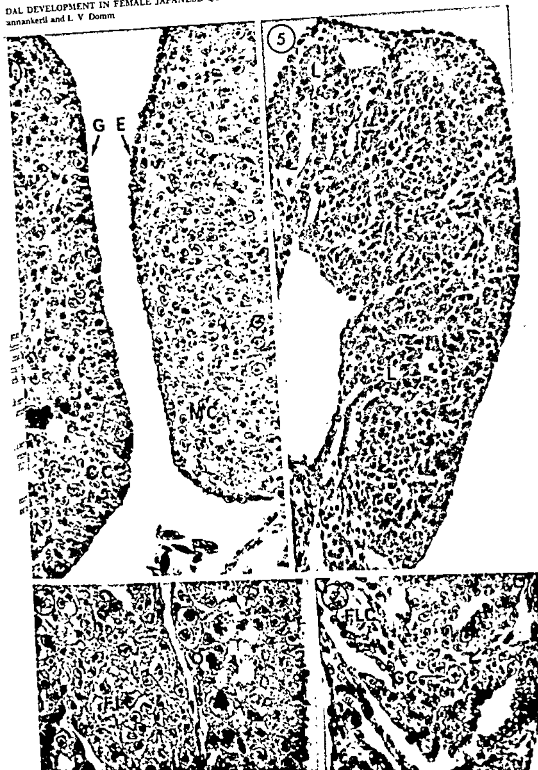


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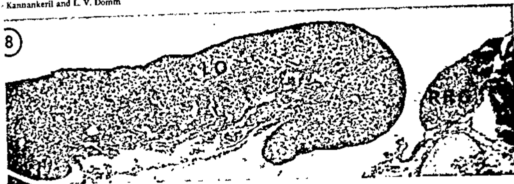
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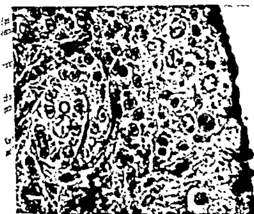
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8
LO
RRO


9
PC
FLC




FLC
PC


12

PLATE 3

EXPLANATION OF FIGURES

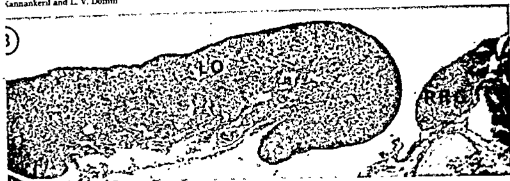
8 Cross section of the rudimentary right gonad and the left ovary of a 14 day embryo. Note the wide disparity in size between the two gonads. LO, left ovary; RRG, rudimentary right gonad. × 135.

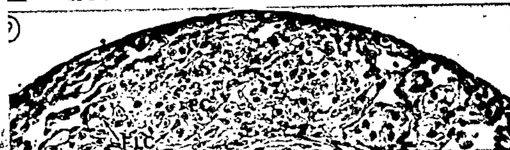
9 Cross section of the rudimentary right gonad of a newly hatched quail (17 days of incubation). Note the "fat laden" cells (FLC) and other loose epithelial cells (EC). × 450.

10 A part of the cortex of the left ovary of the newly hatched quail shown in figure 9. Note the oocyte (O) being enveloped by follicle cells (FC). × 600.

11 Cross section of the rudimentary right gonad of seven day post-hatch quail. Note the groups of "fat laden" cells (FLC) and loosely arranged epithelial cells (EC). × 500.

12 Cross section of the rudimentary right gonad of a two year old quail. Note the increase in fibrous connective tissue cells. Compare figures 5, 9, and 11. × 200.


LO
RRO


FLC




FLC


12

·servations on Epidermal Differentiation :he Fetal Rat[1]

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ABSTRACT Study of fetal rat epidermis has revealed the presence of a single layer of non-keratinizing cells, the periderm, that covers the free surface of the skin during embryonic life. The periderm resembles structurally the epithelium lining the amniotic cavity, and the two are believed to be similar in function and embryonic origin.

Observations of keratinization have shown that keratohyalin granules formed initially during fetal life may differ in structure from those formed during later fetal life and in the adult. Those formed first contain lipidlike globules embedded in a dense granular matrix. Those formed next contain dense patches of finely granular material embedded in a less dense matrix. Granules formed late in fetal life have a homogeneous structure and resemble those of adult skin.

As in adult epidermis, tonofibrils, abundant in epidermal cells prior to cornification, probably contribute to formation of the keratohyalin granules. Also, as in adult tissue, ribosomes lie close to the surfaces of developing keratohyalin granules and seem to be involved in their formation.

lectron microscopic examination of nan fetal skin (Bonneville, '64; athnach and Wyllie, '65; Hashimoto et '66) has revealed that the free surface the epidermis is covered by a layer of ı-cornified cells, beneath which lie the ny cells of the thin stratum corneum. ıs layer has been called the periderm, 1 it is believed to share its origin and ıction with the epithelial lining of the nniotic cavity (Bonneville, '64). In order confirm that the presence of a periderm a general feature in development of : mammalian epidermis, investigation rat skin at various gestational ages ıs undertaken. Fine structural details its development and evidence for its rsistence during the fetal life of the oino rat are presented in this study.

Preliminary observations of the fetal rat in revealed that following primary :ation with glutaraldehyde (Sabatini et . '63) and postfixation with osmium :atives, the keratohyalin granules in the ·st five or so cell layers that undergo ·rnification exhibit marked structural fferences from granules previously ›served in the epidermis of embryo mice ·lenefee,'57), of newborn rats (Matoltsy, 2) and of mature mammalian skin, as :viewed by Brody ('64), Odland ('64) and oth and Clark ('64). In some instances they display morphological variability similar to that described by Farbman ('66a) in rat tongue. This study therefore contains a detailed description of the unusual keratohyalin granules found in fetal rat skin in an attempt to evaluate the significance of these images and to add to the general understanding of the cornification process.

METHODS AND MATERIALS

Pregnant female albino rats were obtained from the Charles River Breeding Laboratories, Wilmington, Massachusetts. Samples of skin were taken from the scapular region of fetuses at various ages of gestation ranging from 10-19 days. In this study, the animals described are referred to both by crown-rump (C-R) length and by their approximate gestational age.

The morphological observations of skin development recounted in this paper were made on skin fixed in 6.25% glutaraldehyde (Sabatini et al., '63) in Sorensen's buffer (pH 7.4) for two hours. After thor-

[1] This investigation was supported in part by four grants from the United States Public Health Service: Research Grant 5-RO1-GM 10299 and Training Grant 5T-01-AM 05220 to Tufts University School of Medicine (Dr Walter F. Lever, Principal Investigator), Research Grant GM 1591 to New England Medical Center Hospitals (Dr George F. Wilgram, Principal Investigator) and Research Grant CA-00592 to Brown University (Dr. Hermann B. Chase, Principal Investigator).

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ough rinsing in buffer, the tissues were postfixed in 1% OsO_4 in Sorensen's buffer (pH 7.4) for two hours. All specimens were dehydrated in ethanol and embedded in Epon (Luft, '61). Sections for both light and electron microscopy were made on a Porter-Blum MT-1 ultramicrotome. For light microscopy, one to two μ thick sections were mounted on glass slides and stained with toluidine blue. Those for the electron microscope were cut with a diamond knife and stained with aqueous uranyl acetate followed by lead citrate (Reynolds, '63). Observations were made on RCA electron microscopes, Models 3F and 3G, and on the Siemens Elmiskop I.

OBSERVATIONS

The periderm

In the youngest fetus examined (crown-rump length, C-R, 15 mm; ca ten days gestation) the skin from the scapular area consists generally of only two cell layers (fig. 1). The layer of flattened cells bordering the free surface of the epithelium is continuous with the lining of the amniotic cavity. During the whole of fetal life this outermost epidermal layer, referred to here as periderm, never undergoes cornification. Its fate during differentiation of the epidermis may be followed in light and electron micrographs.

Light microscopy. Before any fully cornified cells have formed (fig. 2), the young epidermis (C-R, 20 mm; ca 12 days gestation) has differentiated into three layers that correspond to layers found in mature tissue. The basal layer, the stratum germinativum, consists of a single row of low columnar cells. These underlie approximately two layers of rounded cells, the stratum spinosum. They, in turn, are covered by two layers of flattened cells, the stratum granulosum, that contain small dark-staining keratohyalin granules. The most superficial cell layer, the periderm, is made up of cells with rounded nuclei and cytoplasm devoid of granules.

As development proceeds (C-R, 28 mm; ca 15–16 days), the number of cell layers in the granulosum increases (fig. 3). In certain regions of the most superficial granular layer large individual keratohyalin granules may be identified, while in other areas the intergranular spa[illegible] filled with dark-staining material. At t[illegible] stage the periderm layer is thin, but m[illegible] of its flattened cells are occasionally s[illegible] bulging into the amniotic cavity. Still l[illegible] during gestation (C-R, 35 mm; ca 1[illegible] days), when cornification of a second [illegible] layer has begun (fig. 4), the perider[illegible] difficult to detect in light micrographs [illegible] is best examined with the electron mi[illegible]scope.

Electron microscopy. The perid[illegible] has been found in all specimens of f[illegible] skin examined so far. Usually the perid[illegible] appears as a thin layer with short p[illegible]trusions on its free surface (fig. 5[illegible] certain instances, however, the free s[illegible]face is expanded by villuslike struct[illegible] that may be branched (fig. 7). It is in t[illegible] condition that the periderm most rese[illegible]bles the amniotic epithelium (fig. 8) w[illegible] which it is continuous. When the perid[illegible] rests on cornified cells, it is usually fla[illegible] than at earlier stages (fig. 9).

During most of fetal life the f[illegible] structure of the periderm is that of [illegible] healthy functioning tissue. Its cytopl[illegible] contains many fine fibrils, some mi[illegible]tubules, free ribosomes and mitochond[illegible] (figs. 5, 7, 9). The nuclei appear viable a[illegible] do not undergo degenerative changes co[illegible]parable to the striking nuclear alterati[illegible] that occur in cells undergoing cornifi[illegible]tion (Farbman, '66a). During the last t[illegible] days of fetal life its cytoplasm appears l[illegible] well organized (fig. 12) and may unde[illegible] some degenerative changes.

Early in development the periderm [illegible] attached to the underlying granular la[illegible] by desmosomes alone (fig. 5), but fr[illegible] about the twelfth day of gestation [illegible] lamellar material becomes abundant [illegible] the intercellular space (figs. 7, 9). T[illegible] material is believed to be derived fr[illegible] membrane-coating granules (MCG's) [illegible] keratinosomes[3] (see following sectio[illegible] Late in development (one or two d[illegible] before birth) the periderm, though s[illegible] present (fig 12), is less firmly attach[illegible] because the intercellular material a[illegible] desmosomes linking it to the cornif[illegible]

[3] Both terms currently in use for these granules tha[illegible] characteristic of differentiating epidermis are adm[illegible] unsatisfactory, since they are not descriptive of structu[illegible] function. These terms are retained, however, because the[illegible] familiar to a number of workers and because I hesita[illegible] suggest another name before their function is known

beneath it have disappeared (at least the material sampled in this study), ably in preparation for shedding of periderm at birth.

Keratohyalin granules

The most superficial layer of the stratum corneum. Keratohyalin granules appear first in the layer immediately beneath periderm (fig. 5) at about the eleventh of gestation. The granules comprise at two kinds of material: dense homogeneous globular elements that resemble and a matrix of fine dense particles which the globules are embedded. Lipoprotein membranes separating the components of the granule from each other or the surrounding cytoplasm have not been detected.

The granules do exhibit, however, certain relationships with other cytoplasmic structures. Tonofibrils frequently approach individual tonofilaments are apparently embedded in the periphery of the granules, especially when the latter are small (fig. 6). The fibrils are not particularly well developed or abundant in the surrounding cytoplasm, and many profiles of the larger, more mature granules do not show this relationship. Ribosomes, on the other hand, are present in large numbers throughout the cytoplasm, and often they appear to blend into the matrix of the granules (fig. 6). It seems likely that the ribosomes are a normal component of the keratohyalin granules as other workers have suggested (Rogers, '64; Rhodin and Reith, '62).

When keratohyalin granules first appear, the surrounding cytoplasm possesses, in addition to the tonofibrils and ribosomes already mentioned, many small bodies known as membrane-coating granules or keratinosomes (fig. 5). Small Golgi regions and mitochondria are also present. The latter often enclose a large granule (fig. 5), the significance of which is unknown. As the cells become flattened (about the 14th day) the enlarged keratohyalin granules occupy relatively more of the cytoplasmic volume (fig. 7). Recognizable structures in the surrounding cytoplasm consist largely of ribosomes. The contents of the membrane coating granules or keratinosomes are believed to contribute to the intercellular lamellar material, as has been postulated in a number of fine structural studies of epidermis (Farbman, '64; Frithiof and Wersäll, '65; Matoltsy and Parakkal, '65; Matoltsy, '66; Bonneville et al., '68).

About the sixteenth day of gestation, the profile of the large heterogeneous granules may still be seen (fig. 9). The intergranular cytoplasm consists of a homogeneous material, the density of which is nearly equal to that of the matrix of the granule. As in normal and psoriatic epidermis the plasma membrane of such cells has become "thickened" (Matoltsy and Parakkal, '65; Farbman, '66b; Bonneville et al., '68) and appears as a dense outline around the cell. Beneath it is a thin layer of fine, dense granular material.

During the last days of fetal life the intergranular areas of these cells appear as a flattened scale. The "thickened" membrane encloses a homogeneous material almost as dense as itself (fig. 12). The original heterogeneous keratohyalin granules may still be seen, however, giving this layer, when observed in light micrographs, a beaded appearance (fig. 4). This layer has also been observed at the surface of the epidermis of the newborn rat. There is, therefore, no shedding of horny layers until after birth.

The second through fourth layers of the stratum corneum. The next deeper layer (the second beneath the periderm) and the two to three below it become cornified, each lagging behind the one above it. In none of the succeeding layers, however, do the keratohyalin granules appear as in the first layer described above. None contain the lipidlike globules but rather consist of a matrix of fine granular material with irregular patches (figs. 7, 9, 10). Often a nucleoid, apparently of matrix material, is free of the denser granular component (fig. 10). Ribosomes are closely associated with the surface of these keratohyalin granules at all stages of their development, but tonofibrils are closely related to them only during the early ones (fig. 11). At this time the dense patches are apparently confined to the periphery of the granules, and tonofibrils are usually embedded in their surfaces (fig. 11). In spite of the heterogeneous nature of these granules, no

ough rinsing in buffer, the tissues were postfixed in 1% OsO_4 in Sorensen's buffer (pH 7.4) for two hours. All specimens were dehydrated in ethanol and embedded in Epon (Luft, '61). Sections for both light and electron microscopy were made on a Porter-Blum MT-1 ultramicrotome. For light microscopy, one to two μ thick sections were mounted on glass slides and stained with toluidine blue. Those for the electron microscope were cut with a diamond knife and stained with aqueous uranyl acetate followed by lead citrate (Reynolds, '63). Observations were made on RCA electron microscopes, Models 3F and 3G, and on the Siemens Elmiskop I.

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Dense peripheral patches occur in large keratohyalin granules of the fifth cell layer undergoing cornification (fig. 13). It is not known whether these patches are of similar origin and nature to others described above, but it is tentatively assumed that the keratohyalin granules of the fifth cornified layer may form in a manner generally like those in preceding layers.

Small keratohyalin granules forming late in fetal life are apparently homogeneous (fig. 14). Although tonofibrils are closely associated with their outer surfaces, no patches of material can be detected in the peripheral regions of the granules. This may be due to an increased concentration of matrix material that would prevent detection of any patches present. Alternatively, changes in the mode of formation of the granules and in the composition of the granules could be occurring. The specific factors that might influence these changes are, of course, unknown.

Just as in the present study of fetal rat skin, the close association of ribosomes with keratohyalin granules has been noted generally in fine structural studies of epidermis, and their probable role in granule formation has been proposed (Rogers, '64; Rhodin and Reith, '62). It is not apparent, however, whether ribosomes (or their derivatives) occur within the

trace of a similar heterogeneity can be observed in the flattened plates that make up the second, third and fourth cell layers when fully cornified (fig. 12).

The fifth through seventh layer of the stratum corneum. Once the stratum corneum achieves a thickness of four layers, cornification has already been initiated and continues, of course, in the underlying layers. The keratohyalin granules in these layers differ, however, from those previously described. In the fifth layer that undergoes keratinization extremely dense homogeneous deposits are found at the periphery of the granules (fig. 13), the structure of which is made up primarily of fine, dense granular matrix. The deposits are not uniform in shape and project from the granule in such a way as to give it an irregular polygonal profile. After osmium fixation alone the deposits have not been observed, and the granules appear polygonal. Both tonofibrils and ribosomes are closely associated with the surface of such keratohyalin granules.

When the sixth and seventh cell layers beneath the periderm have begun to cornify and have formed small-to-medium sized keratohyalin granules, no internal heterogeneity can be detected (fig. 14). The round-to-oval profiles of the granules that develop late in fetal life are made up of homogeneous dense material. As in the case of all types of granules studied, ribosomes and tonofilaments seem to blend into the material at the surface of the granule (fig. 14).

DISCUSSION

The periderm

This study has shown that throughout fetal life the epidermis of the rat has on its free surface a single layer of cells that do not undergo keratinization. A similar layer has been observed in human fetal skin (Bonneville, '64; Breathnach and Wyllie, '65; Hashimoto et al., '66). The author designated it as the periderm[4] in the belief that it corresponded to the most superficial non-keratinizing layer of human fetal epidermis described by light microscopists (Arey, '54). In regard to fine structure the human periderm cells resemble one cell type found in the epithelial lining of the amniotic cavity (Bourne, '62; Sch- '63; and Thomas, '65). It was poss- that the periderm is similar in orig and may share at least some of the f tions of the amniotic epithelium. In fetal life, before cornification occurs periderm may perhaps effect exch between the amniotic cavity and fet sues. These interpretations may re ably be extended to the rat perid which, though less highly developed that of the human, nevertheless dis structural properties of an absor epithelium and resembles the epit lining of the amniotic cavity. In the clear evidence of secretory activity i amniotic epithelium of the periderm not been seen as it has in human ti (Thomas, '65; Breathnach and Wylie Secretory cells may be present in numbers but were not observed i samples examined in this study.

Two to three days before birth, periderm apparently becomes deta from the underlying epidermis, du breakdown of desmosomes and o intercellular materials. The mechan effecting these changes are unknown.

Keratohyalin granules

It should perhaps be stressed at outset that the appearance of keratohyalin granules, as described in t paper, resulted from the double fixatio glutaraldehyde followed by osmium. C with this procedure was the loss of outer three to four granular la prevented, for in spite of cautious hand these layers were sloughed off a osmium fixation alone, thus it impossible to compare the structure of developing granules after different p parative procedures.

The initial formation of keratohya granules occurs in the cell la immediately beneath the periderm, an has been demonstrated in this study t these granules differ not only from th observed in adult epidermis but also f

[4] The term periderm has also been used to designate a la embryonic chick epidermis (Fitton-Jackson, quoted by Fe In the chick the periderm is eight-to-ten cells in thickne contains droplets of keratin-like material. It therefore do seem to correspond strictly to the periderm described study. At the time I applied this term to fetal epidermis unaware of its use in regard to chick tissues.

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Small keratohyalin granules forming late in fetal life are apparently homogeneous (fig. 14). Although tonofibrils are closely associated with their outer surfaces, no patches of material can be detected in the peripheral regions of the granules. This may be due to an increased concentration of matrix material that would prevent detection of any patches present. Alternatively, changes in the mode of formation of the granules and in the composition of the granules could be occurring. The specific factors that might influence these changes are, of course, unknown.

Just as in the present study of fetal rat skin, the close association of ribosomes with keratohyalin granules has been noted generally in fine structural studies of epidermis, and their probable role in granule formation has been proposed (Rogers, '64; Rhodin and Reith, '62). It is not apparent, however, whether ribosomes (or their derivatives) occur within the

granules, surrounded by the dense material that is produced, presumably, under their influence. Histochemical studies of epidermis (Leuctenberger and Lund, '51; Szodoray and Nagy-Vezekéni, '64) have indicated that the nucleoprotein may be included within the keratohyalin granules as a densely staining homogeneous matrix material is forming. The latter probably corresponds to the granules isolated by Matoltsy ('62) that are resistant to the action of trypsin and to urea in 1 to 3M concentrations. The spatial relationship between the ribosomes and the matrix during granule formation, however, has not been determined in more detail. It seems likely that a large number of ribosomes would form a functional unit that gives rise to a keratohyalin granule. As yet there is no morphological evidence that a focus of synthetic activity has been formed until its product can be detected.

ACKNOWLEDGMENTS

I am indebted to a number of investigators who have a long-standing interest in skin and who each kindly provided in part the laboratory facilities used during this investigation. My sincerest thanks are due to Dr. Walter F. Lever, Tufts University School of Medicine; Dr. Hermann B. Chase and Dr. J. Walter Wilson, Brown University; and Dr. George F. Wilgram, New England Center Hospital. In addition I wish to express my gratitude to Miss Jo Ann Hammons, who assisted in preparation of material for microscopy, and to Miss Janice Glenn for her help in preparing the manuscript and illustrations.

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PLATE 1

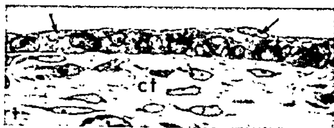
EXPLANATION OF FIGURES

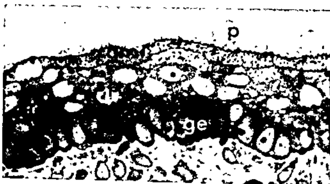
1 Light micrograph of fetal rat epidermis, crown-rump length (C-R), 15 mm; ten days gestation. Two cell layers are present, resting on fetal connective tissue (ct). The flattened superficial layer (arrows), bordering the amniotic cavity, will form the periderm. The basal layer will give rise to the four multicellular layers of mature skin. × 630.

2 Light micrograph of fetal rat epidermis, C-R, 20 mm; 11-12 days gestation. The stratum germinativum (ge), a layer of low columnar cells, lies beneath two layers of rounded cells, the stratum spinosum (sp). The stratum granulosum (gr) is represented by a single layer of cells immediately beneath the periderm (p). The latter is devoid of cytoplasmic granules. × 630.

3 Light micrograph of fetal rat epidermis, C-R, 28 mm; 14 days gestation. Evidence of the presence of a periderm (p) may be seen. The stratum corneum consists of a single cell layer. The keratohyalin granules peculiar to this layer give it a beaded appearance (co). In certain areas, cornification is not complete and individual granules may be seen (arrows). Both the stratum granulosum (gr) and the stratum spinosum (sp) are three to four layers thick. × 630.

4 Light micrograph of fetal rat epidermis, C-R, 35 mm; 18-19 days gestation. Periderm, though present, is thin and difficult to detect in light micrographs. The beaded appearance of the most superficial layer of the stratum corneum is still apparent (arrows). Large keratohyalin granules are present in five to six layers of the stratum granulosum (gr). × 630.


cf


p
ge


gr
p
co
sp
3


4

PLATE 1

EXPLANATION OF FIGURES

1 Light micrograph of fetal rat epidermis, crown-rump length (C-R), 15 mm; ten days gestation. Two cell layers are present, resting on fetal connective tissue (ct). The flattened superficial layer (arrows), bordering the amniotic cavity, will form the periderm. The basal layer will give rise to the four multicellular layers of mature skin. × 630.

2 Light micrograph of fetal rat epidermis, C-R, 20 mm; 11-12 days gestation. The stratum germinativum (ge), a layer of low columnar cells, lies beneath two layers of rounded cells, the stratum spinosum (sp). The stratum granulosum (gr) is represented by a single layer of cells immediately beneath the periderm (p). The latter is devoid of cytoplasmic granules. × 630.

3 Light micrograph of fetal rat epidermis, C-R, 28 mm; 14 days gestation. Evidence of the presence of a periderm (p) may be seen. The stratum corneum consists of a single cell layer. The keratohyalin granules peculiar to this layer give it a beaded appearance (co). In certain areas, cornification is not complete and individual granules may be seen (arrows). Both the stratum granulosum (gr) and the stratum spinosum (sp) are three to four layers thick. × 630.

4 Light micrograph of fetal rat epidermis, C-R, 35 mm; 18-19 days gestation. Periderm, though present, is thin and difficult to detect in light micrographs. The beaded appearance of the most superficial layer of the stratum corneum is still apparent (arrows). Large keratohyalin granules are present in five to six layers of the stratum granulosum (gr). × 630.

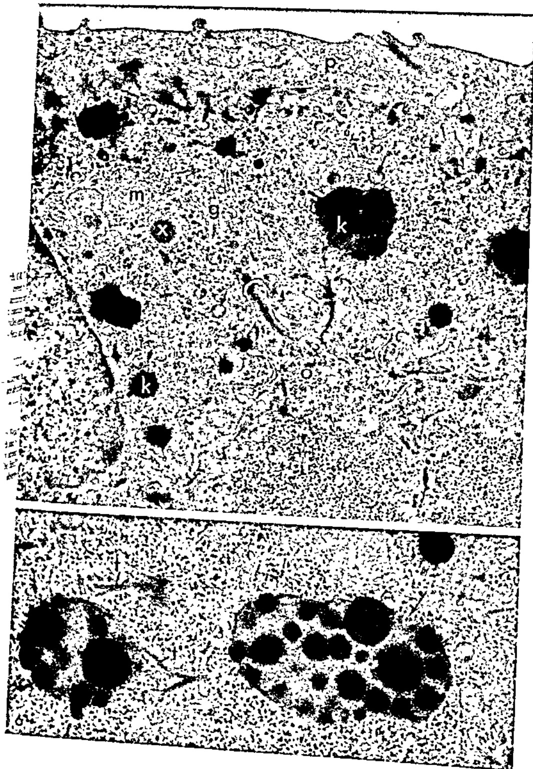

p
m
x
g
k
k
o
5
6

PLATE 2

EXPLANATION OF FIGURES

5 Cross section of epidermis from fetal rat (crown-rump length, C-R, 25 mm; 13–14 days gestation). A thin epithelial layer, the periderm (p), covers the free surface of the epidermis and forms part of the lining of the amniotic cavity. It is attached by desmosomes to the underlying layer, which contains small keratohyalin granules (k). The granules comprise lipidlike globules embedded in a dense matrix. Such granules are unique to this layer of the fetal skin and, as far as the author is aware, have not been observed in mature mammalian epidermis. In early stages of cornification, the cytoplasm surrounding the keratohyalin granules contains Golgi membranes (g), ribosomes, and mitochondria (m). The latter often enclose a large granule (x) of unknown significance. Keratinosomes or membrane-coating granules (arrows) are abundant. Glycogen deposits (gl), typical of cells in the stratum spinosum of fetal epidermis, may also be identified. × 20,800.

6 Keratohyalin granules in cell layer immediately beneath periderm. Fetal rat, C-R, 25 mm; 13–14 days gestation. Tonofibrils (t) frequently approach and blend into the surface of the keratohyalin granules. Ribosomes also may appear to be embedded in the peripheral regions of the granule (arrows). The lipidlike droplets and the dense granular matrix of the granules may be observed. × 39,600.

k
n
k'
k'
k

n
bm
cf
8

PLATE 3

EXPLANATION OF FIGURES

7 Periderm and stratum granulosum of fetal rat epidermis (crown-rump length, 28 mm; 14 days gestation). The periderm cell has an irregular free surface. Such villuslike projections serve to expand the cell surface. A portion of the nucleus (n) of the periderm cell is present. The cell underlying the periderm is flattened so that the large keratohyalin granules (k) form a series of bulges in the cell's profile. Note that the lipidlike globules may be observed within the granules and that the surrounding cytoplasm is rich in ribosomes. Smaller keratohyalin granules (k') occur in the cell lying deep to the first granular layer. Their internal structure is heterogeneous, consisting of a granular matrix, within which denser patches of material are encompassed. Fine lamellar plates, believed to be derived from the keratinosomes, lie in the intercellular spaces (arrows). × 17,500.

8 Epithelial cell of amnion of fetal rat (C-R, 35 mm; 18-19 days gestation). Microvilli project into amniotic cavity. No evidences of cornification are seen in the cytoplasm surrounding the flattened nucleus (n). A thin basement membrane (bm) separates the epithelium from the delicate collagen network of the underlying connective tissue (ct). Compare epithelial cell with periderm shown in figure 7. × 19,200.

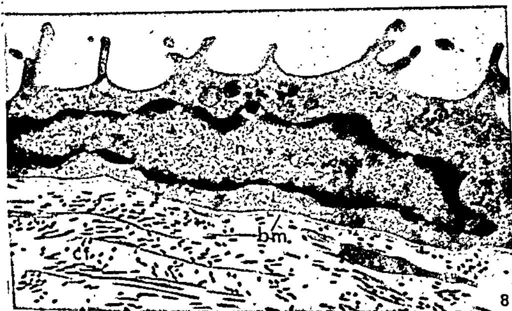
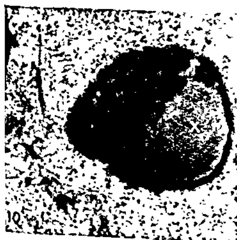

n
b.m.
c.f.
8

PLATE 3

EXPLANATION OF FIGURES

7 Periderm and stratum granulosum of fetal rat epidermis (crown-rump length, 28 mm; 14 days gestation). The periderm cell has an irregular free surface. Such villuslike projections serve to expand the cell surface. A portion of the nucleus (n) of the periderm cell is present. The cell underlying the periderm is flattened so that the large keratohyalin granules (k) form a series of bulges in the cell's profile. Note that the lipidlike globules may be observed within the granules and that the surrounding cytoplasm is rich in ribosomes. Smaller keratohyalin granules (k') occur in the cell lying deep to the first granular layer. Their internal structure is heterogeneous, consisting of a granular matrix, within which denser patches of material are encompassed. Fine lamellar plates, believed to be derived from the keratinosomes, lie in the intercellular spaces (arrows). × 17,500.

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10


t
t

PLATE 4

EXPLANATION OF FIGURES

9 Periderm (p) overlies single layer of cornified cells in the epidermis of fetal rat (C-R, 30 mm; 16 days gestation). Plasma membranes of adjacent periderm cells interdigitate. Desmosomes (d) and close junctions of adjacent cell membranes (j) apparently aid in formation of a tightly sealed covering for the differentiating stratum corneum. The large heterogeneous keratohyalin granules (k) that occur in the cornified cell layer at an earlier stage in development (compare figs. 5, 7) may still be seen, but they are surrounded by additional horny material. A thin layer of dense granular material (arrows) separates the granule as well as the intergranular deposits from the "thickened" plasma membrane. Beneath the corneum, cells of the stratum granulosum contain large keratohyalin granules (k'). Note the dense patches of material that characterize the granules. The cytoplasm that surrounds them is usually devoid of tonofilaments. Fine lamellar plates, believed to be derived from keratinosomes, may be seen intercellularly between the two layers of keratinizing cells (*). × 34,000.

10 Keratohyalin granule within fetal rat epidermis (C-R, 30 mm; ca 16 days gestation). Granule seen here is forming within the second or third layer of cells beneath the periderm. Dense patches of material are embedded in a less dense matrix. In nucleoid-like area (x) no patches occur. Few tonofibrils are associated with granules at this stage of development, but ribosomes usually adhere closely to the surface of the granule. × 42,000.

11 Keratohyalin granule within fetal rat epidermis (C-R, 30 mm). This granule lies in the cell layer immediately beneath that containing the granule seen in figure 10. Note the frequent association between tonofibrils (t) and the surface of the granule. A few dense patches (arrows) of material lie at the periphery of the granule. It is postulated that the patches may arise as fibrillar material is incorporated into the granule. × 42,000.


h
10


t

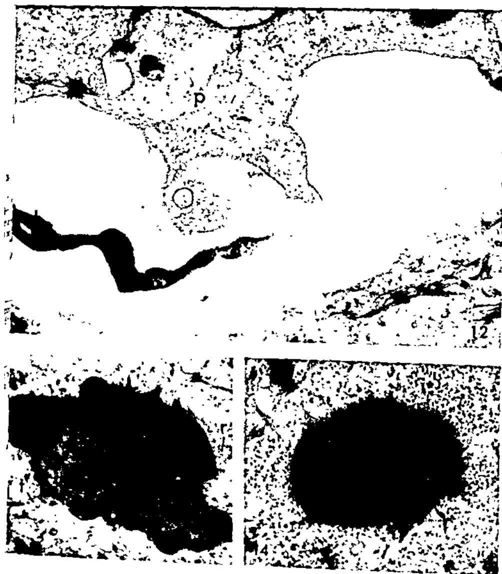
PLATE 5

EXPLANATION OF FIGURES

12 Periderm and stratum corneum of fetal rat skin (C-R, 35 mm; 18-19 days gestation). The periderm layer (p) is not firmly attached to the underlying horny cells as it is at earlier stages of development. In the cornified layer immediately below the periderm aggregates of large lipidlike droplets are found in the keratohyalin granules (k) as they are at an earlier stage of development (compare figs. 5, 7). Succeeding layers (co), however, do not form similar granules. Spaces between horny cells are preparation artifacts. ×26,400.

13 Keratohyalin granule in fifth cell layer beneath the periderm (C-R, 35 mm). Large irregular dense bodies of homogeneous material are embedded in the periphery of the granule, which consists in the main of a deeply staining granular matrix. Tonofibrils (t) seem to blend into the surface of the granule. Ribosomes (r) also lie near the granule, and it seems feasible that some of them may be embedded in the dense granular matrix. ×23,200.

14 Keratohyalin granule forming in the seventh cell layer beneath the periderm (C-R, 35 mm). Note that the tonofibrils and the ribosomes are both closely associated with the surface of the granule. In this granule and those that form later in fetal life and after birth, no internal heterogeneity can be detected. ×32,400.


p
12
14

Electron Microscopic Study of Uterine ›ries During Pregnancy

ERNEST N. ALBERT[2] AND DANIEL C. PEASE[1]

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ABSTRACT The mesometrial arteries of pregnant guinea pigs are not mere blood conducting channels. Instead they are dynamic in nature. The cells forming the intima and media exhibited significant structural modifications during pregnancy. The endothelial cells became hypertrophied and attenuated. Their cytoplasm showed an increase in the amount of ribosomes, rough endoplasmic reticulum and lysosome-like bodies during pregnancy. Dense bodies that were often closely associated with the Golgi apparatus were seen in pregnant animals. These bodies exhibited a graduation in size within each cell and were thought to be the precursors of the lysosome-like bodies. Similar structures were also seen in smooth muscle cells of the mesometrial arteries of pregnant animals.

The endothelium of these arteries was often discontinuous in full term pregnant animals. In that case, the underlying smooth muscl4 cells provided the lining surface. Irrespective of which type of cells formed the lumen of these arteries, they became coated with a "fuzzy" layer which specifically stained intensely with phosphotungstic acid, suggesting that this layer might be a mucoprotein. The smooth muscle cell of the arterial media also became hypertrophied during pregnancy. The cytoplasm exhibited a gradual increase in free ribosomes and rough endoplasmic reticulum, mitochondria and microtubules throughout pregnancy. During the latter half of pregnancy many smooth muscle cells developed extensive amounts of smooth endoplasmic reticulum whose vesicles often were filled with an electron opaque secretory material. It is suggested that the development of unusually large quantities of smooth endoplasmic reticulum might be associated with steroid metabolism. Unusually thick basement membranes were formed around the modified smooth muscle cells of full term pregnant animals and the surfaces of these cells often exhibited large numbers of microvilli. The microvilli seemed to disrupt the continuous patterns of the basement membrane.

electron microscopic studies of the :ular system, the efforts of most investors have been directed principally ards the large arteries and the capillary s. As a result of this, only a few electron roscopic reports are available on the phology and function of the smaller tributing" arteries (Moore and Ruska, Movat and Fernando, '63; Bunce, '65; ›din, '62, '67). The scarcity of the exist- literature on this subject also may be to the general impression that all aller arteries have essentially the same rphology and that they are merely blood tributing channels. However the mesotrial arteries of the pregnant guinea pig not fit into this simple category. It has 'n shown that the endothelial and ooth muscle cells of these arteries are pable of carrying out a wide variety functions (Albert, '67a,b; Albert and ›ussry, '67). Their initimal and medial ils demonstrated cyclic alterations dur- ı pregnancy and post-partum involution. ›e cells of the intima and media were thought to be involved in the degradation and synthesis of elastic fibers during pregnancy and post-partum involution respectively. Although our histochemical stains supported the view that cells of the intima and media resembled endothelial and smooth muscle cells, we had but limited morphological criteria upon which to base decisions. It was also reported that the endothelial lining of mesometrial arteries became discontinuous during pregnancy, and was reconstructed during post-partum involution. There was, however, a possibility that this might have been due to an artifact of fixation. Hypertrophy and hyperplasia of intimal cells was also apparent, but it was not clear as to how and what cytoplasmic organelles caused these changes in size and number. These questions prompted us to begin a thorough study of the morphological alterations that occur in mesometrial arteries of pregnant

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:d rather of plates or islands of elastic ıe since its pattern has been seen only vo dimensions.

nooth muscle cells. The three or four r thick media of the mesometrial arte- was formed by a pure population of :al smooth muscle cells. All muscle ; were enveloped by a continuous thin :ment membrane (figs. 1, 4-6). Collage- ; fibers and occasional fragments of tica were dispersed throughout the ia.

ıe smooth muscle cells (figs. 1, 4, 6) : long and spindle-shaped. They con- ed prominent nuclei that conformed to shape of the cell. The main part of the plasm was occupied with longitudi- ·y oriented myofibrils and associated nse bodies," characteristic of smooth scle, which could be detected amongst m (figs. 4-6). There were electron dense ques immediately below the plasma- ıma that resembled the dense bodies of cytoplasm (figs. 4-6). Often continu- s between the dense bodies of the cyto- sm and the dense patches under the cell mbranes were observed (figs. 4, 5), sug- ting that they may be fundamentally same. Most of the cytoplasmic organ- s were located at the nuclear poles ween the bundles of myofilaments. In eral, the cytoplasmic organelles were abundant. Of the organelles, free ribo- ıes and mitochondria were most com- n. Few microtubules and vesicles of gi, or of smooth or rough surfaced endo- ısmic reticulum could be seen.

Group II: Mesometrial arteries from 1-5 week pregnant guinea pigs

Endothelial cells. The first obvious dification in the endothelial cells, as a ult of pregnancy, was a shape change. transverse sections the cells apparently lened greatly, perhaps as a response to reasing size of the arterial lumen, and : corresponding increase in surface area :. 7) The cells were not notably flattened the early stages of pregnancy, however, by implication their volume had reased substantially (figs. 7, 8). The ent of the attenuations was dependent on the stage of pregnancy.

iurfaces of endothelial cells were cov- d by a continuous dark staining "fuzzy" er which followed the membrane con- irs into open pits and vesicles (fig. 7). This is in contrast to the non-pregnant animals which demonstrated a patchy layer on the endothelial surfaces.

The cytoplasm of endothelial cells in one to three week pregnant animals gave indications of increased metabolic activity. This was reflected in an increase in the number of free ribosomes, while the other organelles did not show noticeable change (fig. 7). By the fifth week of pregnancy, however, the endothelial cells did demonstrate a significant increase in the rough endoplasmic reticulum and also the Golgi vesicles (fig. 8). Pinocytotic vesicles and plasma membrane infoldings could be distinguished from other smooth surfaced vesicles by the presence of dark staining surfaces (fig. 7). These were thought to be due to the "fuzzy" coating that lined the luminal surface of endothelial cells.

Lysosome-like bodies became increasingly apparent after the first three or four weeks of pregnancy (fig. 8). These bodies were of various sizes. Filamentous structures and occasional microtubules also were seen in some endothelial cells of pregnant animals. Patches of thicker basement membrane can usually be seen under the endothelium at this stage of pregnancy.

Smooth muscle cells. Like the endothelial cells, smooth muscle cells also showed an hypertrophy which was reflected after conception by an increase in the organelles. The muscle cells became progressively enlarged. However, with the progress of gestation the myofibrillar zones did not seem to increase. Rather, it was those areas of cytoplasm occupied by organelles which remarkably expanded. This became dramatically evident by the fifth week of pregnancy as demonstrated by figure 9 in which it can be seen that there was by now much rough and smooth surfaced endoplasmic reticulum, prominent Golgi zones, quantities of free ribosomes, patches of glycogen, and probably more than the original number of mitochondria. Thus, areas occupied by myofibrils came to be conspicuously delineated at the periphery of the cells and separated from the main cytoplasmic mass, located centrally. There was no obvious change in the number and distribution of microtubules. Nuclei did increase in size, and developed prominent and patterned nucleoli.

During early pregnancy, vesicular structures with electron dense contents were

animals using the higher resolution and better methods of tissue preparation of electron microscopy.

MATERIALS AND METHODS

Pregnant guinea pigs were used for the present investigation. The animals were divided into three groups. Group I consisted of adult non-pregnant female guinea pigs and served as the control group. Groups II and III represented the experimental animals at various stages of pregnancy. One to five week pregnant animals were put into Group II, and six to nine week pregnant animals into Group III. These animals were sacrificed at weekly intervals throughout their pregancy.

All guinea pigs were anesthetized by an intraperitoneal injection of sodium pentobarbital. Their thoracic cavities were opened and the descending part of the aorta cannulated with plastic tubing. The animals were then perfused for five minutes with 5% glutaraldehyde in 0.2 M Collidine buffer (Bennett and Luft, '58). After perfusion, the abdominal cavities were opened and arterial segments of mesometrial arteries closest to the myometrium were dissected out and further fixed in buffered glutaraldehyde for one hour. Following aldehyde fixation, the tissues were washed in two ten minutes changes of collidine buffer, and post-fixed in 1% buffered osmium tetroxide for an additional hour. After osmication, the tissues were washed again in the buffer and dehydrated in ascending series of alcohols. The specimens were then embedded in Vestopal W. After sectioning they were doubly stained with uranyl acetate and lead citrate (Venable and Coggeshall, '65). Some of the sections were also stained with 5% aqueous phosphotungstic acid (Pease, '64).

OBSERVATIONS

General observations

In the adult nulliparous animal, all mesometrial arteries appeared alike to the naked eye. But in the pregnant animal, some of the arteries were conspicuously larger in diameter and exhibited decidedly greater tortuosity. The arteries that directly supplied the placenta were the largest and most coiled. These will be referred to as "placental" arteries, and the others as "peripheral." The placenta[l arte]ries consistently exhibited prono[unced] alterations in their morphology d[uring] pregnancy, while the peripheral ar[teries] demonstrated minimal and inconsi[stent] changes, depending upon the distan[ce of] the peripheral artery from the plac[enta]. The part of all placental vessels close [to] the uterine horn exhibited ma[ximal] changes. Thus, for the sake of consiste[ncy] all our tissues from pregnant animals w[ere] selected from the placental arteries before they entered the myometrium.

The observations which follow on [the] three groups of animals (non-preg[nant,] early and late-pregnant) are descr[ibed] under individual headings.

Group I: Mesometrial arteries of non-pregnant animals

The endothelial cells. The endoth[elial] cells usually were closely packed, ov[al or] cuboidal. They were separated from [the] muscular media by their basement m[em]brane, the internal elastic lamina, and [a] few scattered collagenous fibers (figs 1[, 2]). The endothelial cell surfaces were usu[ally] smooth with some undulations and oc[ca]sional cytoplasmic processes extend[ing] into the lumen. There were hints of [a] specialized, but incomplete surface la[yer] lining the vascular lumen. This amorph[ous] layer became quite prominent on the lu[mi]nal surface of endothelial cells in me[so]metrial arteries at full term as descri[bed] later.

The cytoplasm of the endothelial ce[lls] contained some mitochondria, a f[ew] strands of rough endoplasmic reticul[um] and free ribosomes. The ribosomes we[re] either scattered individually or occurred [in] groups as polysomes. The Golgi zone [of] these cells was usually small and vestig[ial]. Many inpocketings of the plasma mem[brane] along with deeper vesicles, genera[lly] of a pinocytotic type, were observed on [or] below all surfaces—luminal, medial a[nd] intercellular (fig. 2). Some cells contain[ed] fibrillar structures scattered through[out] the cytoplasm (fig. 3). Lysosome-like str[uc]tures were few or absent. The endothel[ial] cells rested upon a thin basement mem[brane].

The internal elastic membrane w[as] regarded as probably a complete fen[es]trated sheet (fig. 2). However it cannot [be] ruled out that it at times may have c[...]

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Group II: Mesometrial arteries from 1-5 week pregnant guinea pigs

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The cytoplasm of endothelial cells in one to three week pregnant animals gave indications of increased metabolic activity. This was reflected in an increase in the number of free ribosomes, while the other organelles did not show noticeable change (fig. 7). By the fifth week of pregnancy, however, the endothelial cells did demonstrate a significant increase in the rough endoplasmic reticulum and also the Golgi vesicles (fig. 8). Pinocytotic vesicles and plasma membrane infoldings could be distinguished from other smooth surfaced vesicles by the presence of dark staining surfaces (fig. 7). These were thought to be due to the "fuzzy" coating that lined the luminal surface of endothelial cells.

Lysosome-like bodies became increasingly apparent after the first three or four weeks of pregnancy (fig. 8). These bodies were of various sizes. Filamentous structures and occasional microtubules also were seen in some endothelial cells of pregnant animals. Patches of thicker basement membrane can usually be seen under the endothelium at this stage of pregnancy.

Smooth muscle cells. Like the endothelial cells, smooth muscle cells also showed an hypertrophy which was reflected after conception by an increase in the organelles. The muscle cells became progressively enlarged. However, with the progress of gestation the myofibrillar zones did not seem to increase. Rather, it was those areas of cytoplasm occupied by organelles which remarkably expanded. This became dramatically evident by the fifth week of pregnancy as demonstrated by figure 9 in which it can be seen that there was by now much rough and smooth surfaced endoplasmic reticulum, prominent Golgi zones, quantities of free ribosomes, patches of glycogen, and probably more than the original number of mitochondria. Thus, areas occupied by myofibrils came to be conspicuously delineated at the periphery of the cells and separated from the main cytoplasmic mass, located centrally. There was no obvious change in the number and distribution of microtubules. Nuclei did increase in size, and developed prominent and patterned nucleoli.

During early pregnancy, vesicular structures with electron dense contents were

detected, often closely associated with the Golgi complex. Some of these vesicles had a homogeneous matrix while others had translucent centers (figs. 9, 18). In general the smaller ones which were closest to the Golgi zones, had translucent matrices, while the larger ones were homogeneous. This became more apparent in later stages of pregnancy.

The dense patches that were seen in smooth muscle of arteries from non-pregnant animals, located immediately below the plasma membrane were also observed in muscle cells of pregnant animals. They appeared to be larger in size and number and seemed to alternate with cell surface protrusions associated with pinocytotic vesicles. This alternating pattern was more prevalent in smooth muscle of mesometrial arteries from pregnant than non-pregnant animals. The basement membranes surrounding the smooth muscle cells of arteries from pregnant animals become conspicuously thicker than those seen around muscle cells in arteries of non-pregnant guinea pigs (compare figs. 6 and 9).

Group III: Mesometrial arteries from 6–9 week pregnant guinea pigs

The endothelial cells. The pattern of the endothelium and its cells presented a peculiar problem in 6–9 week pregnant animals. In the non-pregnant animals it was generally observed that endothelial cells overlapped each other, but this was not commonly the case in near term guinea pigs. Instead, the endothelial cells in this stage of pregnancy often did not even form a continuous lining around the vascular lumina leaving denuded areas of underlying smooth muscle cells. This could partly be attributed to the increase in the size of the lumen of these vessels during pregnancy. At times cells became so attenuated that their cytoplasms seemed to be at the point of breaking or pinching off (fig. 10), and in the event that this would happen the underlying cells would be without an endothelial covering. Under these circumstances, cells of the media would come in direct contact with the blood as shown in figure 11, and apparently serve the same functions as the original endothelial cells.

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The most prominent feature of modified ooth muscle cells of pregnancy was the earance of an extensive amount of ooth endoplasmic reticulum. In some ls most of the smooth endoplasmic icular vesicles were filled with an elec- n dense material (figs. 15, 19), while se of others were empty (fig. 16). It peared that the rough endoplasmic iculum first became prominent during rly pregnancy. The agranular reticulum peared in substantial quantities later in egnancy, and continued to increase with e progress of gestation. We were unable decide whether the increase in the nooth endoplasmic reticulum was at the pense of the rough or whether it was an dependent formation. Large numbers of itochondria (figs. 10, 15) and microtu- les (fig. 21) also were seen distributed roughout their cytoplasm.

The modified smooth muscle cells elaborated an unusually large amount of a dense material which formed the basement membrane. The thickness of this layer varied considerably from cell to cell, and from animal to animal, and indeed was often discontinuous. Thus dimensions became meaningless (compare figs. 19–22). Nevertheless the thinnest basement membrane was significantly thicker than the ordinary layers seen in non-pregnant animals. Figures 21 and 22 demonstrate continuous basement membranes. Figure 19 illustrates patches of basement membrane separated by microvilli, while figure 20 shows a thick basal lamina, formed of several interrupted layers. This layered appearance gave the impression that such lamina may have formed intermittently. In the absence of basement membranes or when incomplete membranes were present, unusual quantities of microvilli ordinarily seemed to be present (fig. 19). These microvilli were often extraordinarily long and numerous (fig. 19). They gave the impression that they were responsible for the discontinuity of the basement membrane.

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During pregnancy both smooth muscle and endothelial cells showed striking alterations in their shapes and numbers and the arrangement of their cytoplasmic components. By term the smooth muscle cells had changed so much that they bore very litte resemblance to their original morphology, and were hardly recognizable any longer as such.

It was evident from the present study that smooth muscle developed a substantial capacity for synthetic activity as pregnancy progressed. Muscle cells of

detected, often closely associated with the Golgi complex. Some of these vesicles had a homogeneous matrix while others had translucent centers (figs. 9, 18). In general the smaller ones which were closest to the Golgi zones, had translucent matrices, while the larger ones were homogeneous. This became more apparent in later stages of pregnancy.

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mesometrial arteries from non-pregnant animals contained very little rough endoplasmic reticulum, ribosomes or Golgi vesicles. But during pregnancy there was an extensive development of all these organelles. Additionally, the smooth surfaced endoplasmic reticulum expanded in a truly remarkable manner, discussed below.

Although in general smooth muscle cells have not been noted for appreciable synthetic activity, certain recent reports have shown this to be the case in special situations. Thus, uterine smooth muscle has been shown to develop an extensive synthetic apparatus during pregnancy (Laguens and Lagrutta, '64) and under estrogen stimulation (Ross and Klebanoff, '67; Laguens, '64). Gorski and Nelson ('65) also showed an increase in ribosomes in uterine muscle in estrogen stimulated rats. These authors suggested that uterine smooth muscle was probably participating in the secretion of collagen as well as non-collagenous proteins. Parker and Odland ('66) have implicated smooth muscle as manufacturing collagen in atherosclerotic plaques. Pease et al. ('60) have emphasized that the presence of only smooth muscle and endothelial cells in the intima and media of pial and aortic vessels demonstrated that these cells must have been responsible for the synthesis of elastica and collagen, as well as the basement membranes found in those arteries.

One of the most unexpected observations in the smooth muscle of mesometrial arteries of full term pregnant guinea pigs was the development of an extensive agranular reticulum, and the presence of electron dense material in its vesicles. This, coupled with the presence of large numbers of microvilli on cell surfaces, gave the smooth muscle the appearance of secretory cells.

Our electron microscopic studies have not given any specific clues as to the nature of the electron opaque material found in the smooth endoplasmic vesicles. However, the occurrence of large amounts of smooth endoplasmic reticulum often has been associated with steroid secreting cells; interstitial cells of testes (Christenen, '65; Fawcett and Burgos, '60); ovarian cells (Muta, '58); adrenal cortex (Enders and Lyons, '64; Sheridan and Belt, '64; Sabatini et al., '62) and others. Jones and Fawcett ('65) have ... generous doses of progesterone caus... significant increase in the agra... reticulum of liver cells. Finally, the id... tification of specific enzymes necessar... the biosynthesis of androgens has b... located biochemically in the ... components of the agranular reticul... testicular cells by Lynn and Brown (... Based upon these reports, the idea t... smooth muscle under certain condit... may be linked to steroid metabolism see... reasonable.

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m red, Luft, ('65) suggested that it ;ht contain mucopolysaccharides. In present study it was shown that the ninal surfaces of mesometrial arteries n pregnant animals stained specifically h phosphotungstic acid, while none of other cytomembranes were stained. osphophotungstic acid has been shown pirically by Pease ('66) also to be a fairly cific stain for polysaccharides.

t may be well to point out that the omplete endothelium seen in full term gnant animals was not due to improper hniques of fixation, dehydration or bedment. The reality of this observation n further be substantiated by the fact t the fuzzy layer on denuded smooth scle must have formed while these cells re in contact with the normal blood flow. breaks in the endothelial lining had been e to the techniques of tissue preparation, n this layer would not have been sent on the smooth muscle surfaces. her reasons for the actual existence of posed muscle cells have already been orporated in the "Observations."

During pregnancy some of the smooth uscle cells elaborated unusually thick sement membranes and/or irregular asses of similar material. The thickness d pattern of these varied from cell to cell d animal to animal. This observation is no means unique as increases in thick-ss of basement membranes have been ported as a result of aging and species fferences (Donahue and Pappas, '61; oom et al., '59; Stehbens and Silver, '65) s well as in relation to the site of the ves-l (Pease, '60). Pathological thickenings basement membranes also have been scribed often (Zacks et al., '62; Lannigan al., '64). However, in the present obser-ations it should be emphasized that the dothelial basement membrane thick-ned only when the underlying smooth muscle showed morphological alterations. urthermore, the most pronounced thick-ning of the basement membranes was en only around extensively modified mooth muscle cells. To the best of our nowledge, this sort of excessive formation ssociated with smooth muscle cells has ot been described before.

Aside from the modifications that take place in individual cells during pregnancy, the basic arterial skeleton undergoes profound alterations. The internal elastic membrane was always absent in vessels in late pregnancy. Very few collagenous fibers were seen in the intima and media of these vessels by the end of pregnancy. Instead, unusually thick basement membranes and microvilli occupied the intercellular spaces.

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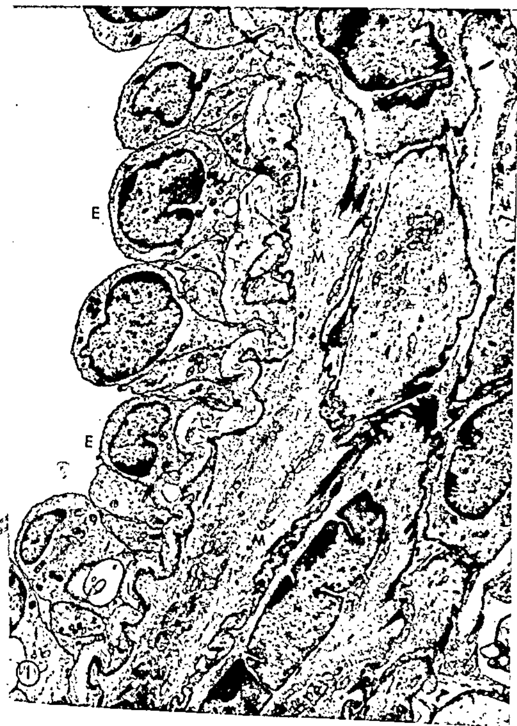
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PLATE 1

EXPLANATION OF FIGURE

1 Cross section of the mesometrial artery from a non-pregnant guinea pig. Endothelial cells (E), internal elastic membrane (I), and smooth muscle cells (M). × 5,500.

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PLATE 2

EXPLANATION OF FIGURES

2 Higher power micrograph of an endothelial cell from the mesometrial artery of a non-pregnant animal. Notice the amount and distribution of the organelles. There are hints of a dark staining surface layer (S). Pinocytotic vesicles are present on all surfaces of the cell in the center. Fenestrated internal elastic membrane (I). × 19,300.

3 Another higher power micrograph of a similar cell as in figure 2. Notice the presence of fibrillar structures in the cytoplasm. × 41,300.

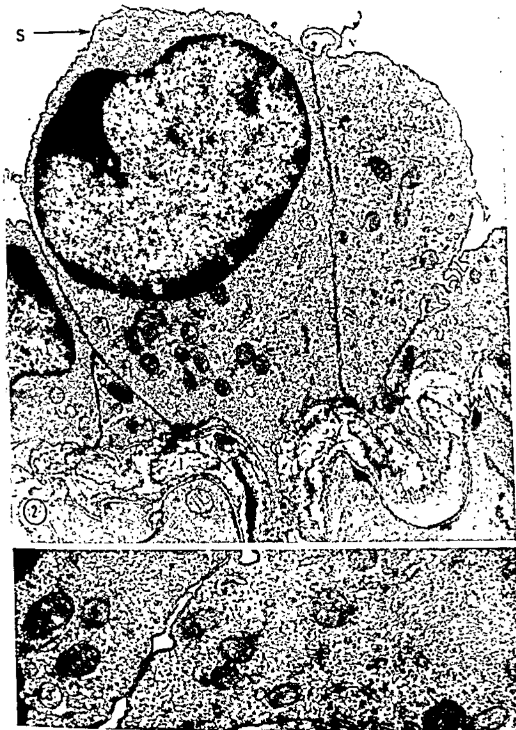

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PLATE 3

EXPLANATION OF FIGURES

4 Smooth muscle cell from mesometrial artery of a non-pregnant animal. The cytoplasm is filled with myofibrils. Dense bodies (D) are seen associated with the myofibrils. Some of them are attached to the dark plaques below the plasmalemma (arrows). ×42,000.

5 Shows the basement membrane associated with smooth muscle of non-pregnant animals. Notice the continuity between the dense body and the dark plaques below the plasma membrane (arrow). ×43,100.

6 Also shows the thin continuous basement membrane (arrow), myofibrils and dense bodies. ×43,100.

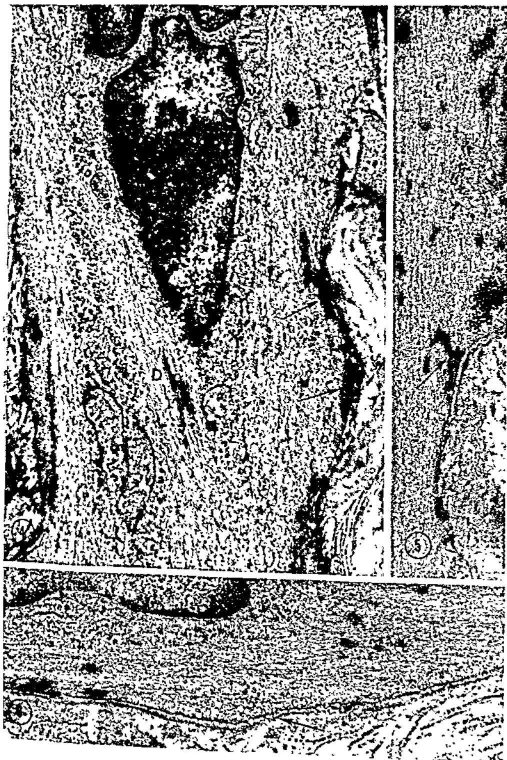


PLATE 3

EXPLANATION OF FIGURES

4 Smooth muscle cell from mesometrial artery of a non-pregnant animal. The cytoplasm is filled with myofibrils. Dense bodies (D) are seen associated with the myofibrils. Some of them are attached to the dark plaques below the plasmalemma (arrows). ×42,000.

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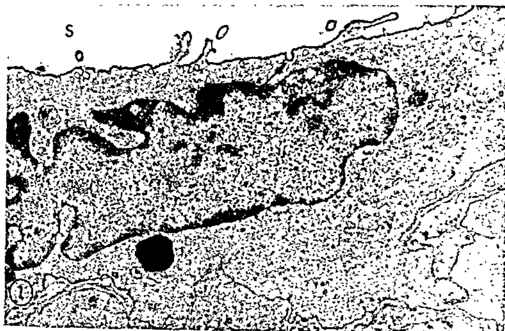

S
L

PLATE 4

EXPLANATION OF FIGURES

7 Endothelial cell from a three week pregnant guinea pig. Notice the presence of a "fuzzy" dark staining surface layer (S). This can also be seen covering the pits and vesicles associated with the plasma membrane (arrow). Free ribosomes are present either individually or in groups as polysomes. × 21,000.

8 Endothelial cell from a five week pregnant animal. Notice the increase in the cytoplasmic organelles over those seen in figure 2, large and small lysosome-like bodies and patches of thick basement membrane (arrow). × 13,500.

S

1

PLATE 5

EXPLANATION OF FIGURE

9 Smooth muscle cell from a five week pregnant guinea pig. Bulk of the cytoplasm is filled with large quantities of organelles, while the myofibrils (M) are pushed toward the periphery. Notice the presence of vesicular structures around the Golgi zone (arrows). These are shown at higher magnification in figure 18. During pregnancy thicker basement membranes also are associated with muscle cells. × 32,700.

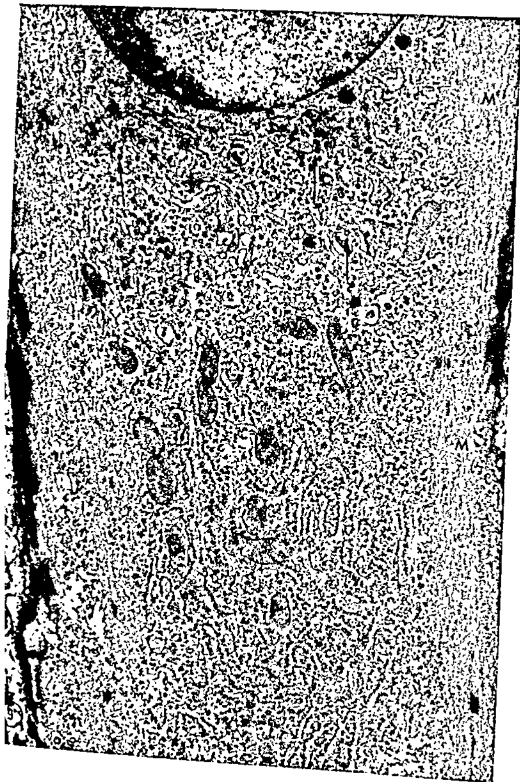

M
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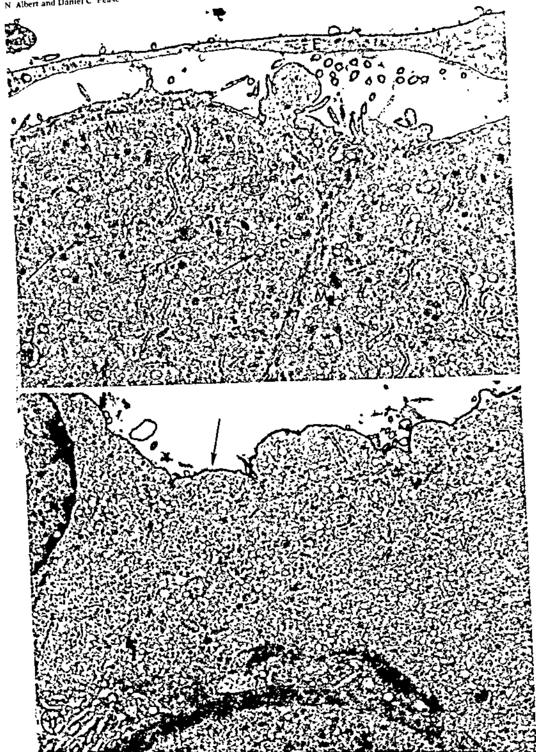

E

PLATE 6

EXPLANATION OF FIGURES

10 Taken from an eight week pregnant guinea pig. The endothelial cell (E) is lining the vascular lumen by a thin attenuated process. The surface layer is coated by the "fuzzy" dark staining layer, but the underlying muscle cell surfaces do not have this layer. Smooth muscle cell processes and microvilli are seen invading the subendothelial zone. The endothelial basement membrane and the internal elastic membrane are absent. The cytoplasm of muscle cells is packed with rough and smooth endoplasmic reticulum. Some of the agranular vesicles contain electron dense material while others are empty (arrows). A thin band of myofibrils (M) is seen around the periphery and also projecting into the cytoplasm. × 19,300.

11 Same stage of pregnancy as figure 10. The endothelium is absent. Smooth muscle surface is coated with the "fuzzy" layer (arrow). × 18,000.



PLATE 7

EXPLANATION OF FIGURES

12 Endothelial cell from a nine week pregnant animal. Surface of the cell is coated with the dark "fuzzy" layer. The cytoplasm is filled with organelles. The Golgi is quite prominent. Notice the presence of vesicular structures closely associated with the Golgi zones (arrow). Some of them have a homogeneously dense matrix. Many of these vesicles are budding off from the membranous stacks of Golgi. This is more clearly shown in figures 13 and 14. Smaller homogeneous dense bodies appear to be increasing in size and are thought to be forming the lysosome-like bodies (L). × 12,250.

13 Higher magnification of the Golgi in figure 12. Some vesicular structures in close association with the Golgi (arrow). × 25,200.

14 Another Golgi zone at higher magnification. Notice the vesicle that appears to be pinching off from the main membranous cistern (arrow). On the opposite side some dense material is seen inside the Golgi vesicle. × 48,700.




14

PLATE 8

EXPLANATION OF FIGURES

15 Modified smooth muscle cell from a nine week pregnant animal. Notice that the cytoplasm contains rough and smooth endoplasmic reticulum. Some of the agranular reticulum vesicles are filled with an electron dense material (arrow). The myofibrils are in the periphery. Dense bodies (D) associated with the myofibrils can be easily identified. × 43,100.

16 Same stage as in figure 15. The cell in the center can hardly be recognized as a smooth muscle cell. Almost all of the agranular reticulum vesicles are empty. The cell in the lower right corner has hardly altered during pregnancy. × 43,100.

PLATE 8

15

16

PLATE 8

EXPLANATION OF FIGURES

15 Modified smooth muscle cell from a nine week pregnant animal. Notice that the cytoplasm contains rough and smooth endoplasmic reticulum. Some of the agranular reticulum vesicles are filled with an electron dense material (arrow). The myofibrils are in the periphery. Dense bodies (D) associated with the myofibrils can be easily identified. × 43,100.

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PLATE 9

EXPLANATION OF FIGURES

17 Part of the media of a mesometrial artery taken from a nine week pregnant animal. The endothelium is absent. The entire media is formed of modified smooth muscle cells. ×4,500.

18 Part of smooth muscle cell from five week pregnant animal. Notice the rounded structures with homogeneous matrices. Some of them have translucent centers (arrows). These are thought to be identical to those described in figures 9 and 12-14. × 45,000.

PLATE 10

EXPLANATION OF FIGURES

19 Represents one of the several patterns of basement membranes observed in late (6-9 weeks) pregnancy. Note the patches of thick basement membrane (arrows) alternating with well developed microvilli. Dense bodies (D). × 53,200.

20 This also represents one of the late stages of pregnancy. Notice the different pattern of the basement membrane. × 22,800.

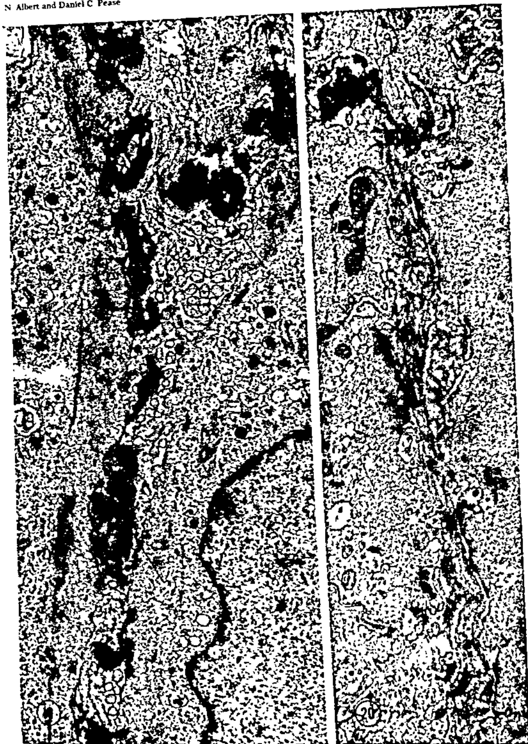

18
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23

PLATE 11

EXPLANATION OF FIGURES

21–22 Show a continuous and homogeneous basement membrane. Although the magnifications are not the same, the basement membrane in figure 21 is proportionally thicker than that in figure 22. Also note the presence of microtubules (arrows) in figure 21. Figure 21, ×49,000; figure 22, ×21,500.

23 Denuded smooth muscle surface as seen after staining with phosphotungstic acid. Only the surface layer is stained. None of the other membranous components show any affinity for this stain. ×71,500.


21
22
23

PLATE 11

EXPLANATION OF FIGURES

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nal Morphology of the English Sole
ırophrys vetulus)

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ABSTRACT The nephron of the English sole, *Parophrys vetulus*, is composed of the following regions: a renal corpuscle with few capillary loops and an extensive mesangial area with mesangial cell processes insinuated between endothelium and podocyte basement membrane; a ciliated neck segment with mucous-containing cells; the first major brush border segment with a broad vacuolated apical zone most closely resembling the mammalian proximal convoluted tubule, a second major brush border segment with the tallest most eosinophilic cells; a gradual transition to a third major brush border segment with lower, more basophilic cells; no distal convolution and a collecting tubule which empties into a collecting duct system. Granulated cells in the arteriolar media appeared to be modified smooth muscle cells.

nique features of fish kidneys have ned the basis for theories concerning ıl physiology and make these organs ıable in correlative studies of structure function (see reviews of Marshall, '34; ıth, '51; Forster, '61).

ıolated flounder kidney tubules are ıl for physiologic studies of transport cesses *in vitro* because of the ease with ıch they may be isolated and mainıed for several hours in oxygenated anced salt solutions (Forster, '48; For- and Taggart, '50; Taggart and Forster, ; Forster and Hong, '58; Hong and For- r, '59). Active transport may be assessed the rate and degree of intraluminal dye umulation.

/e have used flounder tubules in a mulıceted investigation of cell injury ump and Bulger, '65, '67). To establish mal controls, a detailed study was made the structure of the kidney of a local cies of flounder, the English sole, ophrys vetulus, and forms the basis of ; report.

MATERIALS AND METHODS

pproximately 50 representatives of a al species of flounder, the English sole, ophrys vetulus, were used for this dy. The majority of fish were obtained trawl from Puget Sound and were fixed nediately or maintained in recirculat- salt water aquaria for short periods ore use; a few fish were caught by hook l line and fixed immediately.

The fish were stunned by a blow on the head and the body cavity was quickly opened. Kidneys were fixed by perfusion via the renal portal system or by immersion of pieces in fixative solutions. In addition, small pieces were rapidly frozen in isopentane and subsequently freeze-dried and fixed in formaldehyde vapor. The fixative solutions used included the following: 1. 4% phosphate-buffered formaldehyde (Gridley, '60); 2. 6.25% glutaraldehyde buffered in 0.1 M cacodylate with and without post-fixation with 2% osmium tetroxide buffered with *s*-collidine (Sabatini et al., '63); 3. Helly's solution (Gridley, '60); 4. 2% osmium tetroxide buffered with s-collidine (Bennett and Luft, '59); 5. Millonig's solution (Millonig, '62); 6. 10% acrolein buffered in phosphate (Luft, '59); 7. 0.6% or 3% potassium permanganate in water or buffered in cacodylate or veronal acetate (Luft, '56); or 8. a combined fixative containing 6.25% glutaraldehyde and 2% osmium tetroxide buffered in *s*-collidine (Trump and Bulger, '66). Tissues fixed in the first three solutions were dehydrated in ethanol and embedded in a mixture of paraffin and Piccolyte while tissues fixed with methods 2, 4-8 were dehydrated in ethanol and embedded in Epon epoxy resin (Luft, '61).

Paraffin sections were occasionally examined unstained or were stained with hematoxylin and eosin (H&E), the periodic-acid-Schiff technique (PAS), or Gomori's trichrome (Gridley, '60). Serial sections were made and nephrons were

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hat the "membrane" was located er on the pedicel. The basement meme was reflected along the podocytic and did not completely encircle the lary. Coated caveoli were frequently on the plasmalemma adjacent to the ment membrane and desmosomes seen between adjacent processes. A of narrow, short filaments (or fuzz) ated into Bowman's space from the r surface of the plasmalemma of the ocyte. The mitochondria were elond or irregular in shape with a pale rix and a few short cristae. A moderate ber of cisternae of rough-surfaced oplasmic reticulum were present, espey in the cell body region. Abundant oth-surfaced vesicles and free RNP icles were identified. The Golgi apparafrequently was extensive, consisting of tiple cisternae and vesicles on both aces. Multivesicular bodies were racterized by a few internal vesicles a condensation of material in one on on the cytoplasmic side of the limitmembrane. Cytosomes (as defined by sson and Trump, '64) with a pale or lerate matrical density contained a ety of materials including amorphous sities, filaments, membranes or occaial dense rhomboidal crystals. Bundles ilaments traversed the cell bodies and e especially concentrated in the larger processes. The nuclei showed frent indentations and had homogeneous omatin after fixation with osmium teide.

Bowman's space. Bowman's space tained a flocculent material. There , however, a much higher concentraof flocculent material in the capillary nens.

Endothelium. Although the endolium lining the afferent arteriole was k and filled with vesicles, the endothen of the capillary loops was of the thin iety containing both vesicles and pores. capillaries were surrounded by a flocent material which was indistinguishafrom the mesangial matrix. In some ions this material formed a thin layer der the endothelium but in general no tinuous layer of condensed material s found. In some regions endothelial ocesses extended from the basal surface the endothelial cells, penetrating the mesangial matrix to lie adjacent to the basement membrane of the podocyte. In a few regions, the endothelium lay adjacent to the podocytic basement membrane without an intervening layer of mesangium. The endothelium often was closely associated with mesangial processes and desmosomes were seen between the two cell types. The endothelial pores appeared open and were not bridged by a distinct diaphragm.

E. *Mesangial region.* The mesangial region was extensive being composed of many mesangial cells and an abundant mesangial matrix. The mesangial cells were found not only in the axial region of the capillary loops, as seen in normal mammalian glomeruli, but frequently had long processes insinuated between the podocytic basement membrane and the capillary endothelium, thus encircling the capillary loops. Rounded mesangial processes also extended into depressions in the endothelial cytoplasm and occasionally into the capillary lumens. The mesangial cells were characterized by extremely irregular nuclei and a cytoplasm filled with fine filaments. The extensive mesangial matrix which surrounded the cells consisted mainly of a fine flocculent material of low density permeated by fine filaments. The filaments often ran singly but large aggregates were seen. Areas showing a moderate density also could be identified. The mesangial matrix blended into the basement membrane.

II. *Neck* (*figs. 1, 6*)

Near the urinary pole of the renal corpuscle, the cells of the parietal layer became more cuboidal and finally columnar and formed a funnel-like structure marking the beginning of the neck segment. The neck segment varied in length but was usually short. The cells of the neck segment were narrow and columnar with basal nuclei which were elongate or irregular in shape. The cytoplasm was scanty and basophilic containing apical PAS-positive granules and cilia. The diameter of this segment was small with respect to the rest of the nephron. Electron micrographs demonstrated cells of irregular shape showing some lateral interlocking. In addition to the junctional complexes, tight junction regions were seen between adja-

followed to determine the sequence of the morphologic regions. Epon sections were stained with toluidine blue (Trump et al., '61).

Acid phosphatase was demonstrated on frozen sections of formaldehyde-fixed tissue using the technique of Barka and Anderson ('63).

OBSERVATIONS

The kidney of the English sole, *Parophrys vetulus*, lay in a retroperitoneal position on the dorsal and caudal surfaces of the body cavity. Along the dorsal aspect of the body cavity, the kidney was bounded by the backbone and the ribs, and along the caudal aspect, it abutted upon the fused transverse processes of the first caudal vertebra. The kidney formed a single midline structure except in the most anterior region (the head kidney) where the kidney tissue split at approximately the level of the heart and two small projections continued anteriorly.

The kidney was composed of nephrons embedded in hematopoietic tissue. The nephrons consisted of a renal corpuscle, a neck, three major segments with brush borders and a collecting tubule which connected with the system of collecting ducts. This division of the nephron was based on the study of numerous light and electron microscopic preparations as well as reconstructions from serial sections by light microscopy. Some minor variations in the last regions of the nephron, such as dual collecting ducts, could occasionally be found. Scattered throughout the hematopoietic tissue were islands of cells which contained abundant pigment deposits. Few nephrons were found in the head kidney. Located in the hematopoietic tissue of this region, interrenal and suprarenal tissues were identified. Along the dorsal surface of the kidney, the spherular corpuscles of Stannius could be found. Vessels were seen throughout the kidney substance. Nerves and associated ganglion cells coursed along the larger arteries. Also, in long stretches of the media of arterioles, rounded cells filled with PAS-positive granules were seen. These granules contained high acid phosphatase activity.

I. Renal corpuscle (figs. 1, 5)

The renal corpuscle consisted of the following structures: a parietal layer of Bowman's capsule generally composed of squamous cells; a visceral layer consisting of irregular shaped branching cells; Bowman's space between these two cap[sular] layers; a system of capillary loops; and abundant mesangial area.

A. Parietal layer. The parietal layer was lined by squamous cells containing flattened nuclei slightly irregular in shape. Occasional cilia were seen emerging from depressions in the cytoplasm. The mitochondria were large and elongated, contained a pale matrix and few cristae and often appeared as circular profiles and therefore at least some appeared to be tubular. The endoplasmic reticulum consisted of a moderate number of rough surfaced cisternae and occasional circular profiles, and small irregular cisternae of smooth-surfaced elements. A moderate number of free ribonucleoprotein (RNP) particles were present in the cytoplasm. The Golgi apparatus, seen adjacent to the nucleus, was inconspicuous being composed of only a few short cisternae and vesicles. Single membrane-bound bodies consisted of multivesicular bodies and a second type characterized by a more matrix of intermediate density. The cytoplasmic matrix was filled with a large number of filaments. Filaments of small dimensions were seen forming a thick band around the entire cell adjacent to the plasmalemma while a network of larger filaments permeated the remaining cytoplasm. The parietal cells rested on a relatively thin basement membrane. Near the urinary pole the cells increased in height and frequently cells of differing morphology were seen lying between the cells of the visceral layer. These cells were interpreted as wandering cells.

B. Visceral layer. The visceral layer was lined by cells (podocytes) with extremely complex shape similar to that demonstrated by Zimmermann ('15) in mammalian visceral epithelial cells. They consisted of a rounded cell body with several large processes extending peripherally. The large processes gave off small processes (pedicels) which interdigitated with other small processes forming a layer adjacent to the basement membrane. The pedicels were connected by a filtration "membrane" similar to that seen in mammalian renal corpuscles but differ

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cent lateral plasmalemmas in a more basal position.

Cilia emerged in groups of approximately twelve from some apical cell profiles. The cilia were of the usual structure. The basal bodies were connected with an extensive system of striated rootlets. A few apical microvilli extended into the lumen. The majority of the cells contained apical mucous granules (which contained a flocculent material of low concentration). The membranes of the granules often were incomplete. The mitochondria were few in number, irregular in shape and large in size. The rough-surfaced endoplasmic reticulum was well-developed consisting mainly of branching cisternae; free ribosomes were abundant. In fortuitous sections, the Golgi apparatus was extensive consisting not only of vesicles and cisternae, but also of pale vacuoles. The nuclei were irregular having deep cytoplasmic indentations. Wandering cells were occasionally seen in the neck region. The basement membrane varied in thickness but frequently was irregular in contour. Smooth muscle cells sometimes surrounded the tubule, but a complete muscular coat was lacking.

III. *First major segment with a brush border* (*figs.* 1, 2, 7)

A. Light microscopy. The first major segment was composed of tall columnar cells with occasional long cilia projecting into the lumen. The cells possessed a tall PAS-positive brush border. There was a large apical zone which contained vacuoles but lacked mitochondria. A diffuse PAS-positivity was seen in this zone as well. The nucleus lay in the mid to basal region of the cell and appeared circular although irregularities in shape were frequent. The nuclei sometimes appeared rounder, lighter and slightly larger than those of the second major segment. The cytoplasm below the apical zone was eosinophilic although a slight basophilic cast could be seen. After applying Gomori's trichrome procedure with chromotrope 2R staining, this zone was seen to be filled with large elongate or circular mitochondria; PAS-positive granules were seen in the cytoplasm as well. Small acid phosphatase-positive granules were found throu- out the cytoplasm. Occasional cells w intensely stained nuclei were s between the tubular cells which w interpreted as wandering cells of blood gin. Similar cells were seen through rest of the nephron. The PAS-positive b ment membrane was thick, and a sma muscle layer lay external to it.

B. Electron microscopy. The cells c prising the first major segment were and columnar. The apical surface was c ered by long, tightly packed microv After osmium tetroxide fixation, plasmalemmas covering adjacent mi villi often appeared as a row of vesicl this appearance was not noted a fixation with glutaraldehyde or perman nate and was interpreted as artifact. membrane rearrangement. Bundles of f filaments from the interior of the micro swept down into the cell. Similar f filaments from the apical cytoplasm w concentrated at the lateral plasmalem in the region of the intermediate junct Tight junctions were seen along the late plasmalemma in a more basal position addition to the junctional complexes single basal body, associated centriole a striated rootlet fibers were seen in the a cal cytoplasm.

A broad zone of apical cytoplasm und lying the microvillus border contained sicular, tubular and vacuolar profiles id tical with those seen in the mammal proximal tubule.

An extensive system of smooth-surfac membranes occupied the basal cytopla most of which represented plasma me branes when the tissue was well fix After potassium permanganate fixat procedures, two types of membranes w observed in the basal cytoplasm (fig. 1 few were similar in thickness and conf ration to the membranes of the endop mic reticulum while the majority w similar to and confluent with the b plasmalemmas. After fixation with os um tetroxide, only a few of the membr connected with the basal plasmalem Instead they formed large cisternae o ented perpendicular to the basement me brane or occasionally formed circ whorls. Near the basal plasmalemma cisternae appeared to convert into tub

10). After primary fixation with glu- ldehyde-containing fixatives, the mem- es were more frequently seen in conti- y with the basal plasmalemma (fig. 11). e mitochondria were large and elon- containing an abundant number of ae, a moderately dense matrix sub- ce and some matrical granules. Mito- dria were in apposition to the smooth- aced membranes of the basal cyto- m.

e rough-surfaced component of the oplasmic reticulum was fairly abun- t throughout the cytoplasm and quently oriented along the lateral cell nbranes. The smooth-surfaced compo- t consisted of vesicles or irregularly- ped cisternae. The Golgi apparatus was nd adjacent to the nucleus.

ne class of single membrane-bounded ies were small in size, of round, elon- ed or irregular structures with a matrix sity similar to that of the mitochondria. ir close spatial relationship to the mbranes of both smooth- and rough- faced endoplasmic reticulum, their con- tions with the smooth-surfaced mem- nes and their presence in this cell type, make it likely that these represent crobodies.

ytosomes were especially abundant d demonstrated marked variation in fine ucture. The type most frequently seen ntained a pale to moderately dense floc- lent material. A second population of tosomes seen in this region contained arse and fine filamentous material. This ter type of cytosome is found only in this gment. In the flounder, *Paralichthys hostigma*, acid phosphatase activity has en demonstrated in bodies of similar rphology (Trump, Green, and Hickman, published observations).

The basement membrane showed consid- able variation in thickness and homoge- ity. It was generally several times icker than that seen covering rat kidney bules. Although it commonly appeared a single thick layer, a few tubules had a sement membrane coating showing a finite reticulated network. A thin layer of llagen fibrils surrounded the basement embrane, and beyond this a layer of mooth muscle cells formed a peripheral vestment. These cells were filled with fine filaments and dense plaque-like structures.

IV. Second and third major segments with a brush border (figs. 2, 3, 8, 9)

A. Light microscopy. The second major segment was composed of columnar cells that were frequently even taller than those comprising the first major segment. The cytoplasm was intensely acidophilic. A prominent brush border demonstrating a slight PAS-positivity covered the luminal surface and occasional cilia projected into the lumen. Although the small apical region contained no mitochondria or vacuoles, the remaining cytoplasm was filled with large elongate mitochondria (fig. 8). The mitochondria were longer and more numerous than those seen in the first major segment which accounted for the extreme acidophilia and chromotrope 2R staining. The mitochondria were generally oriented perpendicular to the basement membrane of the tubule and frequently extended the entire height of the cell. A few cells in the early part of this segment contained extremely large mitochondria, especially near the base. Oval nuclei usually occupied a midposition in the cells although sometimes they lay slightly basally. PAS-positive droplets were seen in the cytoplasm as well and were especially concentrated in the apical region. Acid phosphatase-positive granules were concentrated in the apex. A thick PAS-positive basement membrane, which showed some variation in thickness, and which was embraced by a layer of smooth muscle cells, surrounded the tubule. At the transition zone between the first and second major segments some intermingling of cell types could be seen.

The second and third major segments had many morphological features in common. In fact there appeared to be a gradual transition between them. Typical cells of the third major segment, however, were lower being cuboidal to low columnar and the brush border was much less conspicuous. This segment had a diameter which was smaller than that of the first or second major segments. An extremely small apical zone was present which lacked mitochondria and vacuoles; however, mitochondria filled the remaining cytoplasm.

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Cilia emerged in groups of approximately twelve from some apical cell profiles. The cilia were of the usual structure. The basal bodies were connected with an extensive system of striated rootlets. A few apical microvilli extended into the lumen. The majority of the cells contained apical mucous granules (which contained a flocculent material of low concentration). The membranes of the granules often were incomplete. The mitochondria were few in number, irregular in shape and large in size. The rough-surfaced endoplasmic reticulum was well-developed consisting mainly of branching cisternae; free ribosomes were abundant. In fortuitous sections, the Golgi apparatus was extensive consisting not only of vesicles and cisternae, but also of pale vacuoles. The nuclei were irregular having deep cytoplasmic indentations. Wandering cells were occasionally seen in the neck region. The basement membrane varied in thickness but frequently was irregular in contour. Smooth muscle cells sometimes surrounded the tubule, but a complete muscular coat was lacking.

III. First major segment with a brush border (figs. 1, 2, 7)

A. Light microscopy. The first major segment was composed of tall columnar cells with occasional long cilia projecting into the lumen. The cells possessed a tall PAS-positive brush border. There was a large apical zone which contained vacuoles but lacked mitochondria. A diffuse PAS-positivity was seen in this zone as well. The nucleus lay in the mid to basal region of the cell and appeared circular although irregularities in shape were frequent. The nuclei sometimes appeared rounder, lighter and slightly larger than those of the second major segment. The cytoplasm below the apical zone was eosinophilic although a slight basophilic cast could be seen. After applying Gomori's trichrome procedure with chromotrope 2R staining, this zone was seen to be filled with large elongate or circular mitochondria; PAS-positive granules were seen in the cytoplasm as well. Small acid phosphatase-positive granules were found thr out the cytoplasm. Occasional cells intensely stained nuclei were between the tubular cells which interpreted as wandering cells of blood gin. Similar cells were seen through rest of the nephron. The PAS-positive b ment membrane was thick, and a sm muscle layer lay external to it.

B. Electron microscopy. The cells prising the first major segment were and columnar. The apical surface was ered by long, tightly packed After osmium tetroxide fixation, plasmalemmas covering adjacent m villi often appeared as a row of vesi this appearance was not noted a fixation with glutaraldehyde or perma nate and was interpreted as artifac membrane rearrangement. Bundles of filaments from the interior of the m swept down into the cell. Similar filaments from the apical cytoplasm concentrated at the lateral plasmal in the region of the intermediate junct Tight junctions were seen along the lat plasmalemma in a more basal positio addition to the junctional complexes single basal body, associated centr striated rootlet fibers were seen in the cal cytoplasm.

A broad zone of apical cytoplasm und lying the microvillus border contained sicular, tubular and vacuolar profiles id tical with those seen in the mamma proximal tubule.

An extensive system of smooth-surfa membranes occupied the basal cytopl most of which represented plasma me branes when the tissue was well fix After potassium permanganate fixa procedures, two types of membranes w observed in the basal cytoplasm (fig. 1 few were similar in thickness and ration to the membranes of the endopl mic reticulum while the majority w similar to and confluent with the ba plasmalemmas. After fixation with os um tetroxide, only a few of the membra connected with the basal plasmalemm Instead they formed large cisternae ented perpendicular to the basement me brane or occasionally formed circu whorls. Near the basal plasmalemma, cisternae appeared to convert into tubu

ι collecting tubule. The cytoplasm was philic and the mitochondria were gen- y inconspicuous. The cell apices uently contained PAS-positive gran- and occasional acid phosphatase- ive droplets. The oval nuclei were gate along the axis of the tubule. A -positive basement membrane and a oth muscle coat surrounded the tubule.

Electron microscopy. The lumen of collecting tubule was widely dilated contained profiles of cilia. Occasional rovilli were seen on the apical surface. majority of the mucous granules seen he apex of the cell lay apical to a layer ilaments associated with the desmo- tes which formed a thick layer across cell apex although occasional mucous plets were seen basal to it. A single al body was occasionally found in the cal cytoplasm.

Numerous small elongate mitochondrial files containing abundant cristae were n throughout the remaining cytoplasm. Cisternae of rough-surfaced endoplas- c reticulum were extremely abundant oughout the cytoplasm and many of the ternae had dilated lumens. The smooth- rfaced endoplasmic reticulum was tensive as well. The Golgi apparatus was tremely large. Elements of the rough- rfaced endoplasmic reticulum were equently seen adjacent to the external ce of the Golgi apparatus. Several Golgi sternae were seen lying in a closely paral- l fashion and the internal face of the olgi consisted of numerous vesicles and me enlarged vacuoles containing a floc- ulent material of low density.

VI. Collecting duct system (fig. 14)

A. Light microscopy. The collecting ibule emptied into a collecting duct ystem in which the ducts converged to orm larger channels finally emptying into mesonephric duct. The epithelial cells ning the collecting duct system started as ow columnar, but increased in height in he larger channels. They possessed a well- lefined brush border. The lumen was pa- ent, large and contained long cilia from he uniciliated epithelial cells; the termi- nal bars were distinct. Immediately below he luminal border was a layer of numerous small darkly PAS-positive granules. The apical zone beneath this layer contained scattered PAS-positive granules. Larger PAS-positive granules were found just above the nucleus. The nuclei were elongate perpendicular to the axis of the tubule and slight irregularities in shape were noted. Cells presumed to be wandering cells were extremely abundant in this region and often stained intensely with the PAS technique. Besides a thick basement membrane, the ducts were surrounded by many layers of smooth muscle, connective tissue elements and capillaries.

B. Electron microscopy. The cells lining the larger collecting ducts of the kidney were similar in morphology to those lining the collecting tubules previously described although they were taller. A layer of microvilli was seen on the luminal border; the apical cytoplasm contained large numbers of mucous granules; and the remaining cytoplasm was filled with small mitochondria containing numerous cristae and a pale matrix. Many of the mitochondrial profiles were elongate and, in the basal cytoplasm, lay adjacent to the smooth-surfaced membrane. A network of elements of the rough-surfaced endoplasmic reticulum lay between the numerous mitochondria. The Golgi apparatus, lying at the level of the nucleus, consisted of cisternae, a large number of small vesicles and large vacuoles of intermediate density. The nucleus tended to lay below the midlevel of the cell. The epithelium comprising the larger collecting ducts was surrounded by several layers of smooth muscle cells and connective tissue elements. Wandering cells were frequently seen between the lateral cell borders in this region in a manner similar to that seen in collecting tubules.

VII. Wandering cells

A variety of cells were seen lying between the lateral cell borders of the kidney tubular cells. The cell most frequently seen contained numerous ribosomes, a few vesicles, an irregular nucleus, an occasional mitochondria, a small amount of rough-surfaced endoplasmic reticulum. Cells containing cylindrical bodies of complex structure were frequently identified. Other cell profiles contained numerous vesicles of varying density, small mito-

The mitochondria were smaller in size and number than those seen in the second major segment and smaller but more numerous than those in the first major segment. A marked basophilic component was also noted in the entire cytoplasm of this latter segment after H&E staining. The apex contained PAS- and acid phosphatase-positive granules. The oval nuclei lay in a basal position. The basement membrane was thick and stained with the PAS technique. A layer of smooth muscle cells was seen surrounding the tubule.

B. Electron microscopy. The apical surface of each region was covered by tall microvilli which were not as tightly packed as those found in the first major segment. The appearance of the apical cytoplasm in the second and third major segments was similar in morphology although the region was much broader in the second major segment. This apical zone contained numerous smooth-surfaced vesicles and/or irregular-shaped units which varied in size and density. Following glutaraldehyde fixation, the apical vesicles were not as prominent. Irregular-shaped units were more frequently seen and the apical zone contained numerous filaments, microtubules and a background matrix of considerable density. A single basal body was occasionally seen in this apical region. Occasional large invaginations and large apical vacuoles were seen in the apical cytoplasm of these two regions which appeared to correspond to the large apical dye-containing structures seen in isolated tubules (Trump and Bulger, '67).

An extensive system of smooth-surfaced membranes was seen in the basal cytoplasm similar to those described for the first major segment. These membranes sometimes formed whorls surrounding various organelles. Tight junction regions were seen along adjacent lateral cell membranes in both segments beneath the level of the junctional complex.

Large numbers of elongated mitochondria filled the cytoplasm of these two segments. The mitochondrial matrix was dense and contained a few short cristae. Matrical granules were also identified. A few cells of the second major segment had extremely large mitochondria in the cell base.

Scattered units of the rough-sur- endoplasmic reticulum were seen thr- out the cytoplasm but were esp- concentrated in the apical region. T- ternae were frequently parallel wi- lateral cell borders. The rough-sur- endoplasmic reticulum was espe- prominent in the third major seg- which most likely accounts for pronounced basophilia of this segment extensive network of irregular-shaped ments of the smooth-surfaced endo- reticulum were seen throughou- plasm.

The Golgi apparatus was most frequ- seen lateral to the nucleus and cons- of several parallel cisternae and an as- ated group of vesicles. Dilated lu- seen in the cisternae most lateral to nucleus.

Cytosomes, cytosegresomes, multives- ular bodies and microbodies were in both of these two segments. Cytos- were present with both pale matrix dark matrix and contained a variety of terials including myelin figures and ments. Cytosegresomes, containing rec- nizable organelles, were present. A few the cytosegresomes were surrounded sac formed from two membranes wh- contained dense material.

Another class of single membr- bounded bodies were seen in these se- ments. They were frequently associ- with the endoplasmic reticulum and c- tained a central core area and theref- were presumed to be microbodies. In cells of the second major segment, th- bodies appeared to be small and freque- seen around the nucleus. In the th- major segment, the microbodies w- considerably larger in size and widely tributed throughout the cytoplasm.

V. The collecting tubule (figs. 4, 13

A. Light microscopy. The collect- tubule was composed of low cuboidal c- and its diameter was generally sma- than that of the third major segment. luminal border of the cell was rectilin- with the long axis of the cell parallel to axis of the tubule (fig. 3). The length of segment was variable and could extremely long. In some instances m- than one major segment emptied in-

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DISCUSSION

Teleost nephrons have been reported to show an extensive diversity in morphology (Huot, '02; Audige, '10; Verne, '22; Edwards, '28, '29-'30, '33, '35; Marshall and Grafflin, '28; Marshall, '29, '34; Grafflin, '29, '31, '33, '37a,b,c; Nash, '31; Defrise, '32; Guyton, '35; Smith, '51; Forster, '53, '61; Black, '57; Newstead and Ford, '60; Bulger, '65), and teleosts live in widely varied environments with profoundly differing needs with respect to salt and water balance. These observations stimulated comparative studies of kidney physiology and anatomy which have advanced the understanding of renal physiology (Marshall, '34). For example, the existence of aglomerular kidneys gave credibility to the proposed mechanism of tubular secretion (see review of Marshall, '34).

Many light microscopy studies were done between 1920 and 1937 of a wide variety of fish kidneys including Edwards' extensive investigations ('28, '29-30, '33, '35). Edwards ('35) listed several types of renal morphology in fish. Marine teleosts were divided into the following groups: 1. Unisegmental aglomerular tubules lacking a distal convolution; a. Entirely aglomerular as exemplified by *Sygnathidae*; b. Partly aglomerular as exemplified by *Muraenidae* (*Muraena helena*) and *Lophiidae* (*Lophius piscatorius* and *Lophius budegassa*); c. Glomerular with slightly patent or nonpatent necks as exemplified by *Gadidae* (*Merluccius vulgaris*); 2. Plurisegmental glomerular tubules lacking a distal convolution; a. Glomerular but lacking an intermediate segment as exemplified by *Trachinidae* (*Trachinus vipers*), *Scorpaenidae* (*Scorpaena*) *scrofa* and *Myoxocephalus octodicimspinus* and others; b. Glomerular with an intermediate segment resembling some of the tubules of *Muraenidae* (*Muraena helena*). This classification does not include the collecting tubule system.

Forster ('61) was under the impres[sion] that the nephron of the common flou[nder] was uniform enough to be consid[ered] consisting solely of tubules resem[bling] mammalian proximal tubules. Our res[ults] indicate, however, that the kidney of [the] flounder, *Parophrys vetulus*, is simil[ar in] morphology to that of other glomer[ular] marine fish in Edwards' ('35) class 2a [with] a plurisegmental glomerular tubule, la[ck]ing distal convolutions and lacking [an] intermediate segment. Only the cells of [the] first major segment resembled the m[am]malian proximal tubule. Although an i[nter]mediate segment and a distal convolu[tion] are lacking in this fish, it seems to us t[hat] applying the term "proximal convolut[ion]" for the entire nephron will lead to con[fu]sion; it should be noted that most of [the] nephron (segments 2 and 3) is quite d[iffer]ent both structurally and functionally f[rom] any part of the mammalian tubule.

Glomerular development is not extens[ive] in marine teleosts (Nash, '31) and, in g[en]eral, marine teleosts have low glomer[ular] filtration rates (Grafflin, '37a; Forster, '5[3]). In fact, certain marine species are a[glo]merular (Huot, '02). It is not surpris[ing] that the English sole renal corpuscl[es], when compared with the well-stud[ied] mammalian species, are characterized [by] several morphologic features which se[em] to reflect the low glomerular filtration ra[te]. These features include: a paucity of cap[il]lary loops; a relatively thick-walled ca[pil]lary with only a few pores; an extens[ive] system of mesangial cells with process[es] which often encircle the capillary; and [an] abundant mesangial matrix. In this re[gard] Hickman (personal communication) h[as] shown that some individuals of the flou[n]der species, *Paralichthys lethostig[ma]* may be functionally aglomerular, posses[s]ing glomeruli of similar morphology [to] those described here.

Two morphologic features of the flou[n]der renal glomerulus indicate a close re[la]tionship between the podocyte and t[he] glomerular basement membrane. First, [the] basement membrane lies adjacent to [the] pedicel border and does not completely s[ur]round the capillary. Instead, the basem[ent] membrane follows the podocytes at infl[ec]tion points leaving a broad zone of ca[pil]lary to abut only on the mesangi[um]. Second, as the mesangial cells send th[eir]

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Cells of the first major segment are haracterized by an extensive brush border d a broad apical zone containing numer- s tubular and vesicular profiles and ge vacuoles similar to those found in the oximal tubules of a variety of mam- alian species (Rhodin, '62, '63; Ericsson, l, '65), fresh water teleosts (Gritzka, '63; lger, unpublished observations), and odeles (Himmelhoch and Karnovsky, 1). These structures have been asso- ated with the uptake of filtered proteins traus, '57, '62, '64; Miller, '60; Miller and lade, '64; Ericsson, '64, '65) or other acromolecules (Trump, '61). Unpub- shed observations in our laboratory have dicated that these apical tubules partici- ate in the endocytosis of ferritin or other olloidal particles injected into the tubular mens. This is in marked contrast to the handling of similar particles in other regions of the nephron. Also this region has extremely large cytosomes which show acid phosphatase activity and thus contains all of the structures usually associated with highly endocytic cells. Cytosegresomes, most of which probably represent foci of autophagy, were only occasionally seen in this or other regions of the normal nephron. This is in marked contrast to tubules incubated *in vitro* where numerous foci of autophagy occurred (Trump and Bulger, '67).

Aglomerular marine toadfish have no filtered protein and also are the only vertebrate studied which lack the apical zone of tubules and vacuoles (Bulger, '65; Trump and Bulger, '67). It seems conceivable, therefore, that the apical structures that have been associated with endocytosis in mammals are present only in teleosts which possess glomerular filtration; in aglomerular fish the luminal surface of cells lining the tubule is not exposed to particles of colloidal size since these cannot enter through the peritubular capillaries and basement membranes. It therefore seems no accident that this segment specialized for endocytosis is not present in aglomerular fish so far studied (Bulger, '65). There would seem to be at least two ways in which this might be controlled. The first is that these structures may actually be induced by the presence of material such as protein in the tubular lumen. The other possibility might be that these segments degenerate during embryogenesis along with the glomeruli because of some common factor such as blood supply. This last hypothesis seems to be favored by the fact that aglomerular fish also lack a neck region which would seem to be rather useful in these fish.

Cells of the second and third major segment also have a specialized brush border though less so than in the first but lack the apical tubules and vacuoles. The apical zone contains some smooth-surfaced membranes whose configuration depends on the fixative employed. This zone is similar to that found in the aglomerular toadfish (Bulger, '65) and probably functions in a similar manner.

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system of smooth-surfaced membranes in the basal cytoplasm, which most likely appears to be plasma membranes. Differences were seen in the configuration of these smooth-surfaced membranes after primary fixation with osmium tetroxide, glutaraldehyde or potassium permanganate. Such a system was therefore described as constituting part of the smooth-surfaced endoplasmic reticulum in *Fundulus heteroclitus* (Gritzka, '63). However, following glutaraldehyde fixation, connections of these membranes with the basal plasmalemma were frequently seen by us. Two types of membranes were distinguished following permanganate fixation, the most abundant of which invariably connected with the basal plasmalemma forming long narrow interdigitating processes in the basal cytoplasm. Although it is presently impossible to distinguish which one represents the *in vivo* state, it seems more likely that membrane breakdown is occurring following osmium tetroxide fixation. A similar system of elaborate basal plasma membranes was noted in the basal cytoplasm of the aglomerular teleost, *Opsanus tau* (Bulger, '65).

The plasma membranes lie adjacent to the numerous mitochondria found in the cytoplasm of all three major segments. A similar association of mitochondria and membranes is frequently found in other ion transporting tissues (see Bulger, '65 for review). This association correlates well with the physiologic role of these cells in active transport. It has been shown that all of the three major segments are capable of transporting chlorphenol red (Trump and Bulger, '67).

The presence of a class of single membrane limited bodies, containing a moderate matrix density and an occasional central crystalline core area, and lying adjacent to units of endoplasmic reticulum, was noted. These bodies probably correspond with the microbodies found in the liver and kidney of other vertebrate species.

Large flask-shaped structures were sometimes found in the apical cytoplasm of all three major segments which occasionally were seen to be confluent with the lumen. Similar structures formed with great rapidity and in large numbers when colloidal particles were injected retrograde into the nephron (Bulger & Trump, un- lished observations).

The urine of marine teleosts, in ge- and of the southern flounder, *Paralic- lethostigma*, which has recently b- studied by Hickman (personal commu- tion), is composed primarily of magn- sulfate with smaller amounts of sod- potassium, calcium, and other anions. magnesium, which represents the m- constituent, is derived from sea water. is probably excreted almost entirely tubular secretion, even when filtr- rates of urine are at maximal levels *Paralichthys lethostigma* not more t- 3% of the magnesium or sulfate excret- filtered (Hickman, personal commun- tion). The sodium concentration is s- what variable; however, it appears t- virtually all sodium and most of t- chloride are reabsorbed actively again- concentration gradient. Even when f- tion stops, as it appears to do in some in- viduals, sodium and chloride, that leak- the lumen, must still be reabsorbed (H- man, personal communication). T- presence of sodium and chloride secre- by a leak-type of mechanism is sugges- by the presence of these ions in rat- large amounts in aglomerular species d- ing laboratory diuresis at which time t- reabsorptive mechanism is evide- blocked (Forster and Berglund, '56).

There are no data presently available these various secretory and resor- mechanisms in the English sole neph- Our ultrastructure studies together w- previously reported studies on aglomer- nephrons (Bulger, '65) can lead to so- tentative interpretations. Since the a- merular fish has a region resembling t- second and third brush border segme- and since calcium and magnesium tory mechanisms are present in t- species, it would seem that divalent i- secretory systems might be found in these regions. Such an interpretation supported by the finding of microcrys- line deposits within the endoplas- reticulum of the second and third br- border regions of *Paralichthys let- stigma*. Similar microcrystalline depo- can be seen in thin sections of urine se- ment by electron microscopy and x- diffraction of such urine sediments reve- the presence of calcium monohydr-

hate ($CaHPO_4 \cdot 2H_2O$) (Trump and ıan, unpublished observations). The of sodium reabsorption is more diffi- o assign. In comparison with other orting tissues, it would seem that all brush border segments, and to a degree the collecting ducts, have the of mitochondrial-plasmalemma asso- n that seem to be important in this d.

region morphologically similar to the tubule of the mammal (Rhodin, '62, or the goldfish (Bulger, unpublished vations) was found in this nephron. e was no juxtaglomerular apparatus e although large granulated cells were running for considerable distances in nedia of arterioles which appeared present juxtaglomerular cells. The ulated cells appeared to be modified oth muscle cells. In this regard, Con- and Kaley ('64) demonstrated the ence of renin in the kidneys of the ıder, *Paralichthys*.

ie cells of the collecting tubules and er collecting ducts were characterized apical mucous granules. The abun- t small mitochondria and moderate ounts of smooth-surfaced membranes nd in these cells indicate that these s play an active role in the functioning ney instead of being merely a passive tem of conduits.

mooth muscle cells were seen sur- nding the entire flounder renal tubule. vnsley and Scott ('63) noted systaltic scular action in the renal tubules of the ınder. We have also observed this in lated tubules. Such muscular action ght play a role in the propulsion of the rate down the nephron. The beating of iated cells in the neck region and the gle long cilia from cells of the remain- g tubular regions is likely to play a role in id movement as well. Marshall ('34) has ted that ciliated neck and intermediate gments disappear coincidently with the quisition of a relatively high blood pres- re as seen in birds and mamals.

ACKNOWLEDGMENTS

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system of smooth-surfaced membranes in the basal cytoplasm, which most likely appears to be plasma membranes. Differences were seen in the configuration of these smooth-surfaced membranes after primary fixation with osmium tetroxide, glutaraldehyde or potassium permanganate. Such a system was therefore described as constituting part of the smooth-surfaced endoplasmic reticulum in *Fundulus heteroclitus* (Gritzka, '63). However, following glutaraldehyde fixation, connections of these membranes with the basal plasmalemma were frequently seen by us. Two types of membranes were distinguished following permanganate fixation, the most abundant of which invariably connected with the basal plasmalemma forming long narrow interdigitating processes in the basal cytoplasm. Although it is presently impossible to distinguish which one represents the *in vivo* state, it seems more likely that membrane breakdown is occurring following osmium tetroxide fixation. A similar system of elaborate basal plasma membranes was noted in the basal cytoplasm of the aglomerular teleost, *Opsanus tau* (Bulger, '65).

The plasma membranes lie adjacent to the numerous mitochondria found in the cytoplasm of all three major segments. A similar association of mitochondria and membranes is frequently found in other ion transporting tissues (see Bulger, '65 for review). This association correlates well with the physiologic role of these cells in active transport. It has been shown that all of the three major segments are capable of transporting chlorphenol red (Trump and Bulger, '67).

The presence of a class of single membrane limited bodies, containing a moderate matrix density and an occasional central crystalline core area, and lying adjacent to units of endoplasmic reticulum, was noted. These bodies probably correspond with the microbodies found in the liver and kidney of other vertebrate species.

Large flask-shaped structures were sometimes found in the apical cytoplasm of all three major segments which occasionally were seen to be confluent with the lumen. Similar structures formed with great rapidity and in large numbers when colloidal particles were injected retrograde into the nephron (Bulger & Trump, un- lished observations).

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1
Cap
I
II
2
I
II
II
3
III
CT
4

Abbreviations

I, First major segment
II, Second major segment
III, Third major segment
BB, Brush border
BM, Basement membrane
C, Cytosomes
Cap, Capillaries
Ci, Cilia
CT, Collecting tubule
ERG, Endoplasmic reticulum, granular
F, Filaments
Go, Golgi apparatus
I, Intercellular space
LD, Lipid droplets
M, Mitochondria
Mb, Microbodies
MC, Mesangial cells
MG, Mucous granules
MM, Mesangial matrix
Mv, Microvilli
N, Neck region
P, Podocyte
SM, Smooth muscles
Ve, Vesicles

PLATE 1

EXPLANATION OF FIGURES

Figures 1-4 were taken of kidney embedded in a mixture of paraffin and Piccolyte. The tissue for picture 4 was stained with the PAS technique and counterstained with hematoxylin. The others were taken from tissues stained according to the Gomori trichrome technique.

1 Portions of a renal corpuscle showing abundant mesangium and peripheral small capillaries (Cap). Neck segment (N), first major segment (I) and second major segment (II) are seen embedded in the abundant hematopoietic tissue. ×475.

2 Two transition regions from the first (I) to the second (II) major segments. Cells of the second major segment are characterized by large, densely staining mitochondria. Certain cells have extremely large mitochondria (arrows). Note intermingling of cell types in junctional regions, especially marked in transition on left. ×520.

3 A transition zone between a third major segment (III) and a collecting tubule (CT). Note the shape of the luminal border of the collecting tubule cells (arrow). ×630.

4 A collecting duct with tall columnar cells containing apical PAS-positive mucous granules (arrow), smaller PAS-positive granules (double arrow), and a thick basement membrane. Numerous densely-staining nuclei of wandering cells can be identified in the tubule wall. The tubule is surrounded by a thick layer containing collagen and smooth muscle cells. ×510.


BM
Cap
MC
MM
MC

PLATE 2

EXPLANATION OF FIGURE

5 Electron micrograph showing a portion of a renal corpuscle. The podocytes (P) have small processes, called "pedicels," which rest on the basement membrane (BM) and surround the capillaries (Cap). The mesangial cells (MC) are numerous and have cell processes which extend peripherally and frequently almost encircle the capillary. The mesangial matrix (MM), which surrounds the mesangial cells, is abundant. The insert is a higher magnification picture showing the relationship between the pedicels and the basement membrane (BM). The filtration slit "membrane" (arrows) lies high on the pedicel. Fixed by vascular perfusion in osmium tetroxide buffered with *s*-collidine. ×11,700. Insert ×47,400.

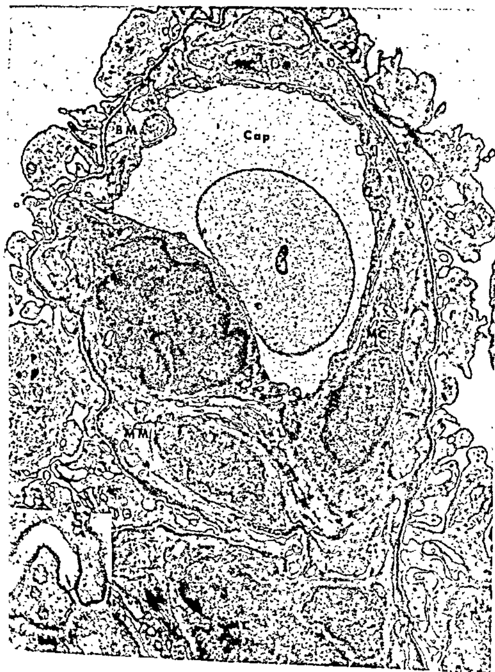

BM
Cap
MC
MM

PLATE 3

EXPLANATION OF FIGURE

6 Electron micrograph of the neck segment showing cellular profiles containing numerous mucous granules (MG) and a cellular profile containing the basal bodies of several cilia (Ci). Cilia are also seen in the lumen of the tubule. Fixed by immersion in osmium tetroxide buffered with *s*-collidine. × 13,900.


MG

PLATE 4

EXPLANATION OF FIGURE

7 Electron micrograph of the first major segment with a brush border. The first segment is characterized by an apical layer of microvilli (BB), abundant cytosomes (C), microbodies (Mb) and mitochondria (M). The apical cytoplasm contains vacuoles, vesicles and dense tubules similar in appearance to those seen in rat proximal convoluted tubule cells. Cilia (Ci) are seen in the tubular lumen. The tubule is surrounded by a thick basement membrane (BM) and a layer of smooth muscle cells (SM). Fixed by vascular perfusion with osmium tetroxide buffered with s-collidine. × 7,400.

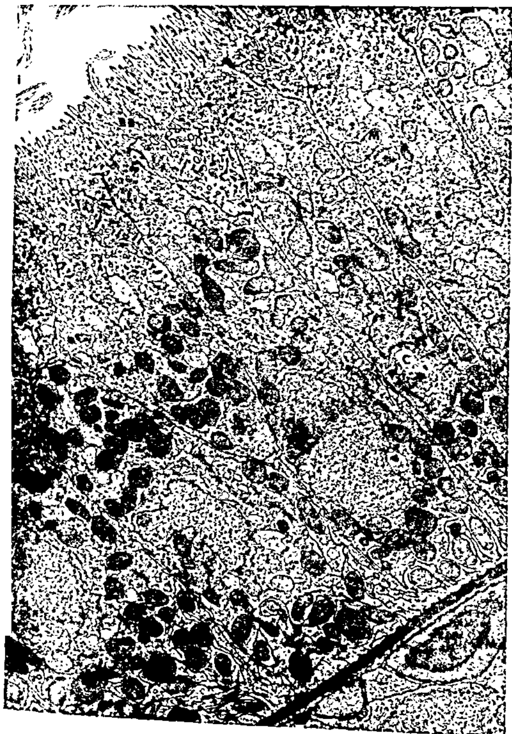


PLATE 4

EXPLANATION OF FIGURE

7 Electron micrograph of the first major segment with a brush border. The fir segment is characterized by an apical layer of microvilli (BB), abundant cyto somes (C), microbodies (Mb) and mitochondria (M). The apical cytoplasm con tains vacuoles, vesicles and dense tubules similar in appearance to those see in rat proximal convoluted tubule cells. Cilia (Ci) are seen in the tubular lumen The tubule is surrounded by a thick basement membrane (BM) and a layer of smooth muscle cells (SM). Fixed by vascular perfusion with osmium tetroxide buffered with s-collidine. × 7,400.

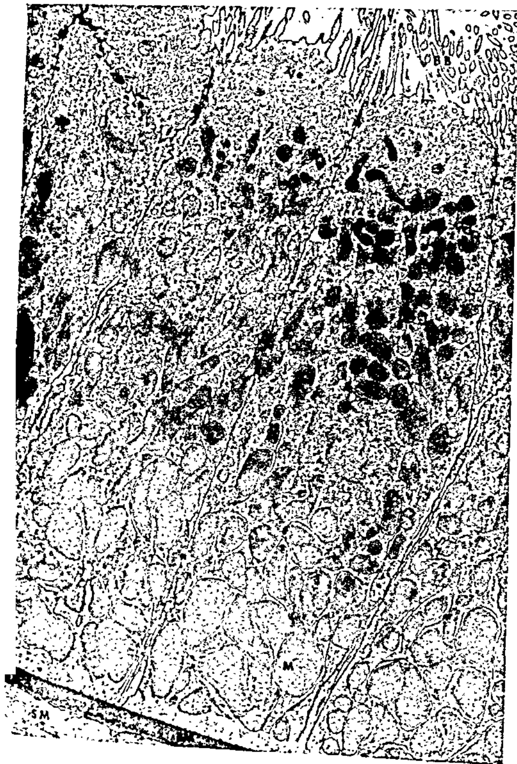

BB
M
SM

PLATE 5

EXPLANATION OF FIGURE

8 An electron micrograph of the second major segment with a brush border. The profiles of the second major segment are characterized by long microvilli (BB), an apical layer of cytoplasm filled with vesicles (Ve), and abundant mitochondria (M) filling the rest of the cytoplasm. The first few cells of this region sometimes have some extremely large mitochondria (M) in the basal cytoplasm. A thick basement membrane (BM) and a layer of smooth muscle cells (SM) surround the tubule. Fixed by perfusion with osmium tetroxide buffered with s-collidine. × 5,900.

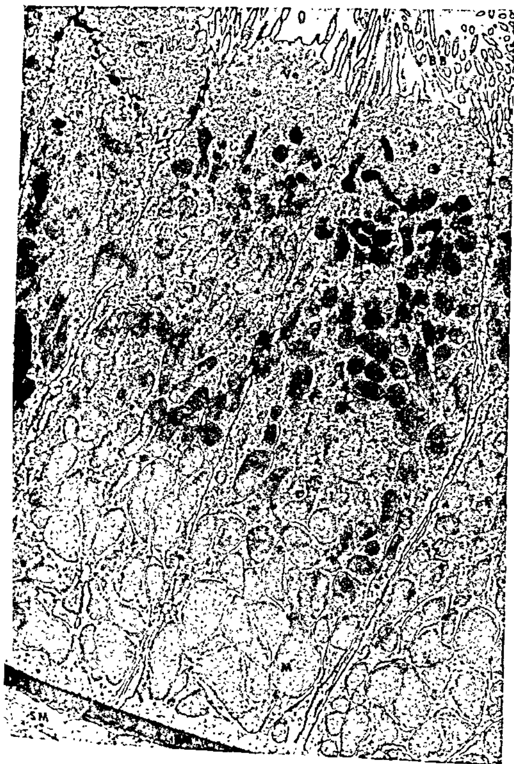

B
M
SM

PLATE 6

EXPLANATION OF FIGURE

9 Electron micrograph of the third major segment with a brush border. The cells of the third major segment are similar in appearance to those of the second segment although they are lower in height, contain more rough-surfaced endoplasmic reticulum (ERG) and have a smaller apical zone of vesicles (Ve). Note slight expansion of lateral intercellular space (I). The tubule is surrounded by a thick basement membrane (BM) and a layer of smooth muscle cells (SM). Fixed by immersion in osmium tetroxide buffered with s-collidine. × 9,200.


SM
L
BB

PLATE 6

EXPLANATION OF FIGURE

9 Electron micrograph of the third major segment with a brush border. The cell of the third major segment are similar in appearance to those of the secon segment although they are lower in height, contain more rough-surface endoplasmic reticulum (ERG) and have a smaller apical zone of vesicles (Ve Note slight expansion of lateral intercellular space (I). The tubule is surrounde by a thick basement membrane (BM) and a layer of smooth muscle cells (SM Fixed by immersion in osmium tetroxide buffered with s-collidine. × 9,200.


11

PLATE 7

EXPLANATION OF FIGURES

10 Electron micrograph showing the basal cytoplasm of a tubular cell fixed in osmium tetroxide. The smooth-surfaced membrane (arrows) seen in the basal cytoplasm do not often connect with the basal plasmalemma. Fixed by vascular perfusion. × 28,400.

11 Electron micrograph showing the basal cytoplasm of the tubular cell after fixation in a mixture of osmium tetroxide and glutaraldehyde. The smooth-surfaced membranes in the basal cytoplasm more frequently connect with the basal plasmalemma (arrows). Fixed by immersion. × 27,700.

12 Electron micrograph showing the basal cytoplasm of a tubular cell after fixation in potassium permanganate. The basal plasmalemma is continuous with the smooth-surfaced membranes of the basal cytoplasm. However, occasional membranes are seen (arrows) which do not connect with the basal plasmalemma and appear to be endoplasmic reticulum. Fixed by immersion. × 69,700.


11

PLATE 8

EXPLANATION OF FIGURE

13 Electron micrograph of a collecting tubule. The collecting tubules are char acterized by apical mucous granules (MG) with a layer of fine filaments (F lying basal to most of the granules. The mitochondria are smaller than thos found in the other regions of the tubule although they are abundant. A well developed Golgi apparatus (Go) and occasional lipid droplets (LD) are seen Fixed by immersion in osmium tetroxide buffered in s-collidine. × 13,000.

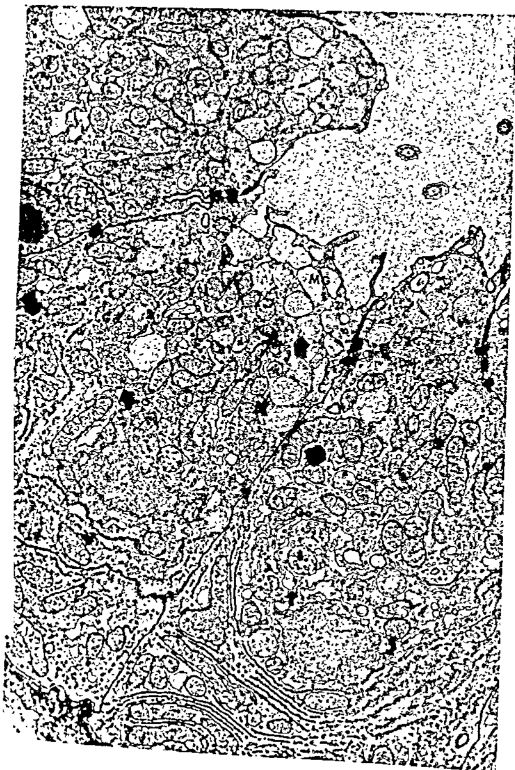

MC

PLATE 8

EXPLANATION OF FIGURE

13 Electron micrograph of a collecting tubule. The collecting tubules are characterized by apical mucous granules (MG) with a layer of fine filaments (F) lying basal to most of the granules. The mitochondria are smaller than those found in the other regions of the tubule although they are abundant. A well-developed Golgi apparatus (Go) and occasional lipid droplets (LD) are seen. Fixed by immersion in osmium tetroxide buffered in s-collidine. × 13,000.


MV
MG
IS
M

PLATE 9

EXPLANATION OF FIGURE

14 Electron micrograph of a large collecting duct. A large collecting duct is characterized by a layer of apical microvilli (Mv), a layer of mucous granules (MG), a large Golgi apparatus (Go), abundant but small mitochondria (M) and a thick coat of smooth muscle cells (SM) surrounding the tubule. Fixed by immersion in osmium tetroxide buffered with s-collidine. $\times$4,100.


MV
MG

; Uptake of Ferritin by Ileal Absorptive Cells in ckling Rats. An Electron Microscope Study [1]

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ABSTRACT An electron microscopic study of the small intestine of rats from birth to 15 days of age has demonstrated a gradient of both structure and function along the length of the small intestine. During this period of development, only the ileum was found to absorb intact exogenous ferritin or colostral proteins. In newborn rats ferritin particles or milk proteins are incorporated by ileal absorptive cells into tubular invaginations of the apical plasmalemma which connect with vacuoles or cisternae in the apical cytoplasm. The ferritin particles are then transported by another vacuolar system to the supranuclear region where several such vacuoles coalesce to form a single supranuclear vacuole. After the first feeding the supranuclear vacuole is established in the ileal cells so that additional ingested proteins, whether ferritin or from milk, are transported from the cisternae directly to the supranuclear vacuole. There was no evidence in the present study to indicate that ferritin is transmitted from the ileal cell to the circulation. The subject of protein transport by the intestinal epithelium is reviewed briefly.

The mucosa of the ileum in neonatal gulates, rodents, and carnivores differs narkably from the adult ileum in its rphological (Clark, '59; Kraehenbuhl, oor and Blanc, '66), cytochemical (Corll and Padykula, '65), and physiological aracteristics (Brambell, '58, '66). Ileal sorptive cells from neonatal animals of ese groups can incorporate ingested proin and transfer certain immune globuas from the absorptive cell into the cirlation. This is an important mechanism r passive immunization of the newborn. laternal antibodies secreted into the costrum and milk can be absorbed and tilized by the suckling animal until active ntibody synthesis is established. The inorporation and transport of protein by the eum of suckling rats ceases about three reeks after birth when there is a transiion to the typical adult type (Halliday, 55a,b; Clark, '59). In some animals from he above species, particularly the rat and log, there is also a prenatal transfer of antibody via the yolk sac and intestine. This is not an important pathway quantitatively in these animals, but in the rabbit and monkey prenatal transfer of proteins via the yolk sac and placenta, respectively, is the principal means of passive immunization.

In those species which absorb intact proteins during the suckling period, the fine structure of the intestinal absorptive cells in the ileum is distinguished from adult absorptive cells by the presence of many small apical cytoplasmic vesicles and a large supranuclear vacuole (Möllendorff, '24; Smith, '25; Comline, Roberts and Titchen, '51b; Comline, Pomeroy and Titchen, '53; Hill and Hardy, '56; Clark, '59; Payne and Marsh, '62; Anderson, '63; Graney, '64; Cornell and Padykula, '65; Kraehenbuhl et al., '66). Clark ('59) studied the uptake of proteins during the early postnatal period in rats and mice by examining the passage of gamma globulins and electron-dense particles across the intestinal mucosa. Three hours after feeding bovine gamma globulins he was able to localize them with a fluorescein-labeled antibody in the supranuclear inclusions of absorptive cells in the midportion of the small intestine and in associated lacteals and mesenteric lymph nodes. Electron microscopic studies with colloidal tracers revealed the uptake of gold and saccharated iron oxide particles into apical cytoplasmic

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Preparation of tissues

animals were anesthetized by an injection of nembutal (0.04 gm body weight) and a laparotomy performed to expose the intestinal t. Small samples of the proximal jeju-n and terminal ileum were excised, cut small blocks and immediately placed ixative.

The fixatives used were 3% glutaralde-le in 0.1 M sodium cacodylate or phos-te buffer (Millonig, '62) and 1% osmi-tetroxide in phosphate buffer (Millonig,) for one to two hours prior to dehydra-n. The tissues were either rapidly de-lrated in graded alcohols or, in some es, were dehydrated in acetone and ined in the block with 0.5–1% po-sium permanganate according to the thod of Parsons ('61). After dehydra-n the tissues were embedded in Epon Araldite (Luft, '61). Thick sections (1–) were cut on an ultramicrotome and amined either by light microscopy after ining with toluidine blue or by electron icroscopy. Thin sections were mounted copper grids with a substrate consisting either formvar or celloidin and a carbon m and were doubly stained with uranyl etate (Watson, '58) and alkaline lead lts (Ryenolds, '63; Venable and Cogges-all, '65).

OBSERVATIONS

Regional variations in the gross morphology of the small intestine

The duodenum and jejunum in the normal suckling animal can be distinguished rom the ileum by their creamy white color n contrast to the brownish-yellow or am-ber color of the ileum (Möllendorff, '24). The distinctive regional coloration of the small intestine corresponds with regional differences in absorptive function of the small intestine that are observed after feeding suckling animals ferritin or colloidal tracers, e.g., carbon or trypan blue.

After young rats are gavaged with a solution of ferritin, there is no change in the creamy color of the jejunum when examined at intervals of 0.5, 1, 2, 4, 6 or 24 hours. When the intestine is opened, the presence of the tracer is revealed by a reddish-brown color in the lumen of the jejunum 30 minutes after feeding, but all of the tracer can be washed away from the mucosal surface by gentle rinsing in saline. In contrast, one hour after feeding ferritin the normal amber color of the upper ileum changes to an intense reddish brown. By two hours after gavage, the ferritin solution completely traverses the small intestine, and the entire ileal mucosa is a reddish-brown color. This is in striking contrast to the pale jejunal mucosa. If other tracers are used, such as trypan blue, hemoglobin or carbon, the amber color of the ileum becomes masked by the color of the tracer. None of the tracers used could be washed from the ileal mucosa with saline rinses, and the pigmentation was still present 24 hours after feeding (the longest time point studied).

These simple macroscopic observations suggest that the jejunum is not important in the uptake of ferritin or particulate tracers. However, light and electron microscopic studies of this area were undertaken in order to verify these conclusions and to compare the cytological features of jejunal and ileal mucosa at various times and under both conditions of fasting and suckling.

Light microscopy

Jejunum. The cytological detail of columnar absorptive cells in the jejunal mucosa of newborn rats (prior to suckling) and fasting young rats is very similar to that of the adult. The apical cytoplasm in all of these cells has a coarse granularity which is probably due to the presence of mitochondria (figs. 1, 3, 5). Particulate inclusions are not prominent. However, in tissues fixed within 30 minutes after young rats are allowed to suckle, the jejunal absorptive cells contain variable amounts of lipid which is distributed in droplets throughout the apical cytoplasm and in the Golgi apparatus (fig. 3). Four hours after suckling, most of the jejunal cells have been cleared of obvious fat droplets (fig. 5).

The initial uptake of lipid by the intestinal mucosa occurs in cells located on the apices of the intestinal villi and later by cells situated along the sides and base of the villi as described previously in the

tubules, membrane-limited vesicles and the supranuclear vacuole. These findings were interpreted as evidence for pinocytotic activity in these cells. It was noted that the colloidal particles appeared to be retained by the lining cells, while some of the gamma globulins were discharged into the lamina propria. It has been shown by others (Comline, Roberts and Titchen, '51a; Hill and Hardy, '56; May and Whaler, '58; Pierce and Johnson, '60) that proteins are usually transported from the lamina propria by lymphatic rather than blood capillaries. More recently, it has been shown that proteins of 70,000 M.W. or less are absorbed via portal blood (Balfour and Comline, '59; Pierce, Risdall and Shaw, '64).

In the current study, ferritin, a naturally occurring electron-dense protein, was introduced into the gastrointestinal tract of suckling rats in order to study further the absorption of intact protein at this stage of development. In particular, it was of interest to determine whether this tracer protein would be transferred from the intestinal lining cells to the lamina propria and thus establish the successive steps in the pathway of protein absorption.

MATERIALS AND METHODS

Animals and experimental grouping

Long-Evans and Charles River rats from newborn to 15 days of age were divided into the following groups:

Controls. (1) Newborn rats separated from the mother prior to suckling, (2) newborn rats separated from the mother 0.5, 1 and 2 hours after suckling, and (3) young rats, 4–15 days of age, separated from the mother 0.5, 1, 2, 3, 4, 6 and 8 hours after suckling. Some of these animals were maintained in a fasting state (no solid or liquid food) for periods up to 24 hours.

Lipid studies. Two 15-day-old rats were separated from the mother and given 1 ml of Mazola corn oil via a gastric tube. The animals were subsequently killed after intervals of one and two hours respectively.

Ferritin studies. Ferritin was administered to newborn rats prior to suckling and to suckling rats up to 15 days of age. A small polyethylene tube (P.E. 10–50) was passed into the esophagus of an [illegible] thetized animal and 0.1 or 0.2 ml of [illegible] lyzed ferritin solution was injected [illegible] tuberculin syringe attached to the t[illegible]

Preparation of gross specimens

Five 5-day-old suckling rats were [illegible] for a special macroscopic study of [illegible] small intestine. One animal was a [illegible] suckling control and the remaining [illegible] each received one of the following [illegible] tions by gastric tube: (1) 0.2 ml of a [illegible] ferritin solution, (2) 0.5 ml of a [illegible] blue solution, (3) 0.2 ml of a 10% he[illegible] globin solution, and (4) 0.3 ml of an [illegible] ink solution. The animals were killed [illegible] hours after gavage and the entire [illegible] intestine from gastroduodenal junction [illegible] caecum was excised and placed in [illegible] saline. A pair of fine scissors was used [illegible] open the intestine and expose the muc[illegible] The specimen was then rinsed in seve[illegible] changes of physiological salt solution [illegible] fixed *in toto* in 10% calcium-formol.

Dialysis procedure for ferritin solutio[illegible]

The ferritin solution used in this st[illegible] was obtained from either of two [illegible] mercial sources (Nutritional Biochem[illegible] Corporation, Cleveland, Ohio; Pentex [illegible] corporated, Kankakee, Illinois). Both [illegible] mercial preparations of ferritin, app[illegible] mately 10 gm% in concentration, [illegible] similarly crystallized from extracts [illegible] horse spleen with cadmium sulfate [illegible] was, therefore, necessary to dialyze [illegible] ferritin solution against a chelating a[illegible] ethylene-di-nitrilo-tetracetic acid (ED[illegible] to remove toxic cadmium contami[illegible] (for details see Farquhar and Palade, [illegible] The ferritin solution was then ren[illegible] isotonic by a subsequent dialysis ag[illegible] Gey's balanced salt solution for 72 [illegible] with three changes (Wissig, '64).

The final dialyzed solution of fe[illegible] was passed through a Millipore [illegible] (0.45 μ) and refrigerated in a sterile [illegible] ber-stoppered serum bottle. Under [illegible] conditions the ferritin solution ma[illegible] stored for at least 6 months withou[illegible] significant deterioration. Small quan[illegible] may be removed as needed with a [illegible] syringe and needle.

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These simple macroscopic observations suggest that the jejunum is not important in the uptake of ferritin or particulate tracers. However, light and electron microscopic studies of this area were undertaken in order to verify these conclusions and to compare the cytological features of jejunal and ileal mucosa at various times and under both conditions of fasting and suckling.

Light microscopy

Jejunum. The cytological detail of columnar absorptive cells in the jejunal mucosa of newborn rats (prior to suckling) and fasting young rats is very similar to that of the adult. The apical cytoplasm in all of these cells has a coarse granularity which is probably due to the presence of mitochondria (figs. 1, 3, 5). Particulate inclusions are not prominent. However, in tissues fixed within 30 minutes after young rats are allowed to suckle, the jejunal absorptive cells contain variable amounts of lipid which is distributed in droplets throughout the apical cytoplasm and in the Golgi apparatus (fig. 3). Four hours after suckling, most of the jejunal cells have been cleared of obvious fat droplets (fig. 5).

The initial uptake of lipid by the intestinal mucosa occurs in cells located on the apices of the intestinal villi and later by cells situated along the sides and base of the villi as described previously in the

adult rat (Hewitt, '54; Palay and Karlin, '59b; Ladman, Padykula and Strauss, '63).

The uptake of milk lipid by the jejunal mucosa in ferritin-fed animals appeared to be the same as in normal control animals. There was no evidence of ferritin uptake by the jejunal absorptive cells, e.g., dense vacuoles; and jejunal tissue from either ferritin-fed or control animals could not be distinguished by light microscopic study.

Ileum. The appearance of ileal absorptive cells in the newborn rat prior to suckling (fig. 2) is similar to the jejunal absorptive cells in the same animal. In some young, the ileal absorptive cells contain a few small vesicles in the apical cytoplasm, deep to the area of the terminal web. Their number was variable, but in no case were there as many vesicles in the newborn as in the suckled or ferritin-fed animals.

After the young have suckled, the appearance of the ileal absorptive cells changes conspicuously, acquiring a unique character which distinguishes them from other absorptive cells in the proximal part of the intestine. A large vacuolar structure appears in the supranuclear area, displacing the nucleus toward the base of the cell (figs. 4, 9). In the suckling pig, a large cytoplasmic vacuole is formed at the base of the absorptive cell and the nucleus assumes an apical position (Payne and Marsh, '62; Mattisson and Karlsson, '66; Sibalin and Björkman, '66; Vodovar and Fléchon, '66). The contents of the vacuoles vary. They may contain either homogeneous dense material, clumps of dense material aggregated about the periphery of the vacuole; or their contents may appear as a meshwork of finely vacuolated, flocculent material. The size of the vacuole varies depending upon whether it is located in an absorptive cell on the apex or base of a villus. The largest vacuoles were usually located in cells on the apex of the villus, with a gradation in size to the smallest located in absorptive cells on the base of the villus. Cells in the crypts of Lieberkühn do not contain supranuclear vacuoles. The apparent local absorption gradient located in an apico-basal direction along the height of the villi is reminiscent of that observed during absorption of fat in the jejunum of suckling young and that reported for the adult jejunum during fat absorption (Hewitt, '54; Palay and Kar '59b). The vacuole persists in the supra nuclear region even when the animal is fasted for periods of up to 24 hours (fig

Fine structure

The fine structure of intestinal epithe lial cells of suckling rats has been de scribed previously (Clark, '59; Kraehe buhl et al., '66; Dunn, '67). However, som structural feautres have not been consid ered heretofore and, in particular, the cy logical differences between the proxim half of the small intestine (duodenum an jejunum) and the distal half (ileum) ha not been emphasized. These differences which are not apparent in the adult, repre sent a longitudinal differentiation of ultr structure in the small intestine of th suckling rat and presumably underlie th specialized absorptive function of the prox imal and distal small intestine.

Jejunum. The fine structure of jejuna absorptive cells from either newborn ra (prior to suckling) or suckling rats (whic are subsequently fasted) is indistinguish able and is comparable to the fine structu of jejunal absorptive cells in fasted ad rats (Palay and Karlin, '59a; Carde Badenhausen and Porter, '67).

When jejunal absorptive cells of youn rats were examined 30 minutes to 2 hours after suckling, osmiophilic droplets presumably lipid, were observed distendin the endoplasmic reticulum of the apic cytoplasm. At later periods, the Gol vesicles and intercellular spaces becom distended with lipid droplets, appearin finally in the lamina propria. The matri of these lipid droplets often appears e tracted and sometimes only a thin sh of dense material remains. This was more consistent finding after glutarald hyde fixation than with osmium fixatio In an attempt to produce better visuali ion of the pathway of lipid through t jejunal cell, Mazola corn oil was given gastric tube to two suckling rats and th jejunums subsequently fixed in osmi (fig. 8). The pathway of the lipid throu the jejunal cells appears to be the sa whether the animal is fed milk or corn In either case, it is not sufficiently diffe from the studie f adult jejunu

d Karlin, '59b; Cardell et al., '67) to rrant discussion here.

the jejunal cells of suckling s absorb lipid in quantity, the present estigation has not demonstrated the capacity of these cells to absorb protein. Ferin was not observed in the jejunal cells, any time interval, after it was administered to either fasting or suckling rats, en though its presence in the jejunal men could be verified by the color of the ntents at the time of autopsy.

There were no obvious differces between jejunal and ileal lining cells newborn rats (prior to suckling) when udied by light microscopy, but an elecon microscopic examination reveals a ndamental difference in their ultrastrucre. The plasmalemma in the intermicrollus spaces of ileal absorptive cells forms vaginations extending for some distance to the apical cytoplasm (figs. 10, 11). hese invaginations often branch and anstomose with others from neighboring ntermicrovillus spaces as they penetrate he cytoplasm. The resulting labyrinth of nterconnected tubules is confined to a and or zone across the apical part of the ell, intermingled with mitochondria and lements of the endoplasmic reticulum figs. 10, 11). Ultimately, the tubules terminate in a vacuolar structure or perhaps better term is cistern, since there are vacuoles of similar size which apparently do not have tubular connections (figs. 10, 11, 14). Within the lumen of these tubules and cisternae there is a fine, flocculent material, but prior to suckling the quantity of this material is not significant. After suckling, however, the dominant feature of these cells is the abundance of this flocculent material throughout the tubular labyrinth and within the cisternae. This substance is similar to that described by Clark ('59) and is a proteinaceous material, presumably derived from the milk and succus entericus. In addition, large vacuoles without obvious tubular connections contain this material also and appear to coalesce, forming the single, large vacuole in the supranuclear region described earlier in this paper. An amorphous material identical to that in the supranuclear vacuole was noted also in the intercellular space (fig. 9). The supranuclear vacuole persists in ileal absorptive cells, at least in those on the upper three-fourths of the villus, even when the animal is fasted for 18–24 hours (fig. 6). With the normal feeding cycle of the young, this means that supranuclear vacuoles are present in the ileum from the first meal until about three weeks after birth when the ileal cells are gradually replaced by cells typical of the mucosa of the adult. After fasting the cells on the basal part of the villus resemble the ileal cells from newborn animals, i.e., they contain a tubular apparatus with its associated cisternae but lack supranuclear vacuoles. The flocculent material within the tubular apparatus is sparse. Presumably, these are differentiating cells which have migrated up the villus during the fasting period.

After ferritin gavage, individual ferritin molecules were identified in the intestinal lumen, intermicrovillus spaces, and within the invaginations of the apical plasmalemma of ileal lining cells (figs. 11–14). Within 1.5 hours after administration of ferritin the tubular apparatus contains the ferritin tracer, as do the associated vacuoles or cisternae (figs. 13, 14). Vacuoles without associated tubules are predominant in the supranuclear area and contain ferritin in high concentration also. It is not clear whether the smooth-surfaced vacuoles bud off from the cisternae or represent cisternae which have lost their tubular connections. In any event, these vacuoles migrate to the supranuclear area where they coalesce, forming a single large vacuole (fig. 14). The progressive increase in size of this vacuole, as successive ferritin-filled vacuoles migrate from the apical area and fuse with it, ultimately results in formation of the definitive supranuclear vacuole. The nucleus is gradually displaced basally to accommodate the expanding vacuole.

On occasion, ferritin particles are seen in crystalline patterns within the smooth-surfaced vacuoles adjacent to the supranuclear vacuole as well as in the supranuclear vacuole itself. More often, the content of the tubular apparatus, apical vacuoles, and supranuclear vacuoles is heterogeneous, consisting of both ferritin particles and a fine, flocculent material

similar to that observed in suckling controls.

The successive steps in the formation of the supranuclear vacuole described above pertain to the newborn rat and the changes which occur after the first meal, whether it consists of milk or ferritin. In older suckling animals, the supranuclear vacuole is already established, so that when additional protein is ingested and taken into the apical tubules and cisternae, the smooth-surfaced vacuoles shuttle the protein directly to the existing supranuclear vacuole. In this study, the supranuclear vacuole appears to represent the terminus of the absorptive pathway for ferritin in the intestinal epithelial cell. It was not observed in association with the Golgi apparatus and there was no evidence to indicate that ferritin was transported from the lining cells to the intercellular space or lamina propria. This is in disagreement with the report of Kraehenbuhl, Gloor and Blanc ('67). A few particles were observed free in the cytoplasm, but these might represent endogenous ferritin or ferritin artifactitiously released from the tubules and vacuoles. The use of glutaraldehyde fixation proved crucial in this regard. Multiple attempts with osmium tetroxide as a primary fixative in combination with a spectrum of buffer systems, consistently resulted in the diffuse distribution of ferritin molecules throughout the cytoplasm of jejunal absorptive cells, the intercellular space, and the lamina propria. Ferritin was retained within membrane-limited systems only when glutaraldehyde was used as the primary fixative.

DISCUSSION

Morphological and functional gradients in the small intestine

The existence of an apico-basal gradient in the structure and function of an adult intestinal villus has been recognized for some time (review by Padykula, '62). The results of the present study have emphasized a longitudinal or proximo-distal gradient in the ultrastructural characteristics of the intestinal lining cells of suckling rats which is related to regional differences in their specific absorptive functions.

The proximal part of the small intes
actively absorbs lipid, but it does not a
sorb intact protein. This interpretation
supported by the observation that neith
ferritin nor aggregates of amorphous p
cipitated material originating from t
milk were identified in lining cells of t
duodenum or jejunum of the young r
Moreover, there is no tubular apparatus
the apical cytoplasm of jejunal lining ce
comparable to that found in the ileum, a
therefore, the jejunum appears to lack t
morphological apparatus necessary for a
sorption of intact proteins.

In two papers reporting ferritin absor
tion by the jejunum of adult rats (Card
et al., '67; Casley-Smith, '67) the expe
ments were done using ligated intesti
segments and a much higher concentrati
of ferritin than the one employed in t
study. The quantity of ferritin taken
under these conditions was very sma
compared with that absorbed by the ileu
of suckling rats; therefore, the ability
the jejunum to absorb significant amoun
of intact protein under normal conditio
remains to be demonstrated. The ileum
the suckling rat absorbs protein primari
but it can absorb lipid also. Cornell a
Padykula ('65) have shown that the ileu
of the suckling rat can absorb an emul
fied lipid when it is injected into the inte
tinal lumen. From the observations in t
present study and those of Cornell a
Padykula ('65), the apical tubular app
ratus and supranuclear vacuole do n
play an obvious role in the absorption
lipid, at least pariteulate lipid. In view
the current concepts of lipid absorptio
(Cardell et al., '67); Strauss and Ito, '65
it is conceivable that free fatty acids
other soluble lipids enter the tubular lab
rinth where they diffuse into the apic
cytoplasm and subsequently become ester
fied in the endoplasmic reticulum. Und
normal physiological conditions, howeve
it appears that only small quantities
lipid reach the distal part of the small i
testine because of the efficiency with whic
lipid is absorbed in the proximal part. I
addition to the differences in structure an
function described above, there are var
ations in the enzymatic activities betwee
the proximal and distal small intestin
which is further evidence of a linea

ctional gradient; both β-glucuronidase (eringova, Jirsova and Koldovsky, '65) d β-galactosidase (Koldovsky and Chytil,) activities are higher in the ileum than the jejunum of suckling animals. Acid osphatase activity is greater in the ileum d has been demonstrated by histochemi- means to be in association with the ranuclear vacuole and adjacent small cuoles (Cornell and Padykula, '65; Sher- y, '66).

Alkaline phosphatase activity, however, greater in the duodenum than in either e jejunum or ileum (Moog, '51, '62; Kol- , Faltova, Hahn and Vacek, '61). It interesting to note that whereas alkaline activity increases after day , β-glucuronidase and β-galactosidase ac- ities decrease. All of the changes in the tivities of the enzymes mentioned above n be delayed by adrenalectomy or in- iced precociously by administration of s cortisone (Moog, '62; Koldovsky nd Chytil, '65; Heringova et al., '65). It hould be emphasized that in addition to gional qualitative differences in the mor- hological, enzymatic, and functional fea- ures which distinguish the jejunum and leum, there are also quantitative changes ccurring within each of these regions luring the course of the suckling period. n the rat, this period is 21 days, but in other animals it may be considerably less. These points have not always been appre- ciated by some investigators, resulting in erroneous generalizations about intestinal histophysiology.

Mechanism of protein uptake by intestinal epithelial cells

Schechtman ('56) has emphasized that there are two distinct steps in the move- ment of proteins across a cellular layer. The first step, in the case of the intestinal epithelial cell, is the uptake of the protein at the surface of the cell and the second is the discharge of protein from the lateral and/or basal surface of the cell. The tacit assumption is that proteins cannot enter the lamina propria by passing directly through the intercellular spaces. There is no evidence at present for the intercellular passage of protein through the intestinal epithelium. There is, however, both physio- logical and immunological data (Brambell, '58, '67; Clark, '59) which show that pro- teins are incorporated nonselectively into intestinal lining cells of certain suckling animals. A variety of proteins and col- loidal particles can be incorporated into the cell, e.g., albumin, gamma globulin, saccharated iron oxide (Clark, '59), fer- ritin (Graney, '64, '65; Kraehenbuhl et al., '67) and peroxidase (Straus, '65), but only specific proteins are discharged from the cells. The mechanism of this discharge will be discussed in the following section.

The rat is born with an antibody titer approximately one-sixteenth that of the maternal serum, but by the second or third day after birth it is equal to or exceeds that in the maternal serum (Halliday, '55b). During the postnatal period, the rat con- tinues to absorb intact antibodies until day 20, although the quantity absorbed be- gins to decrease after day 15 (Culbertson, '39; Halliday, '57; Clark, '59).

The rate of transfer can be quite rapid; measurable quantities of antibody appear in the circulation of the young rat within 30 minutes after feeding immune serum. By three hours the circulating titer has attained a maximum concentration (Halli- day, '55a).

Clark ('59) first suggested that protein was incorporated into the intestinal epi- thelial cells of suckling rats and mice by the process of pinocytosis. This concept has been reiterated for the suckling puppy (Rubin, '66), piglet (Mattisson and Karls- son, '66; Sibalin and Björkman, '66; Vodo- var and Flechon, '66) and rat (Kraehen- buhl et al., '67). The morphological observations of the present study do not seem to be completely consistent with this concept. Ferritin particles enter the apices of the lining cell through a tubular laby- rinth and not by incorporation into vacu- oles resulting from folding of the surface membrane as in the original concept of pinocytosis (Lewis, '31). Similarly, there was no pinocytotic uptake of ferritin *in quania* (Palade, '53) by vesicles of either the smooth-surfaced or coated variety, al- though this phenomenon has been ob- served in several other systems (Wissig, '58; Fawcett, '65).

An alternative viewpoint is that the api- cal tubules and their cisternal terminals are a relatively stable system within the

apical cytoplasm and that the lumen of this system is confluent with the intestinal lumen, i.e., extracellular. This would provide an access for nonspecific diffusion of protein, or other substances for that matter, from the intestinal lumen through the apical tubules and into the cisternae or vacuolar terminals of the tubules. The precise means by which protein moves from the cisternae to the supranuclear vacuole remains elusive. From Clark's studies ('59) and the results obtained with ferritin there is little doubt that the nonselective incorporation of protein proceeds past the point of the cisternae and as far as the supranuclear vacuole. Whether this occurs by a pinching off of small vesicles from the cisternae or from a separate vacuolar system is unknown. There is histochemical evidence which supports to some extent the concept of two separate membrane systems. Cornell and Padykula ('65) have shown that the apical cisternae (apical droplet in their terminology) are rich in esterase, whereas the supranuclear vacuole and adjacent small vacuoles contain both acid phosphatase and esterase activities. Since the supranuclear vacuole is formed in newborn young by the successive coalescence of vacuoles in the supranuclear region, some form of a vesicular transport system must be operating.

Selective discharge of protein from intestinal lining cells

An electron microscopic examination of the ileum from at least ten rats removed at intervals varying from 30 minutes to 24 hours after feeding ferritin has not demonstrated the passage of ferritin beyond the point of the supranuclear vacuole. This disagrees with the results of Kraehenbuhl et al. ('67) who described the movement of ferritin to the basal area of the supranuclear vacuole where some of the particles were incorporated into Golgi vesicles while others appeared to migrate freely through the cytoplasm and lateral cell membrane into the intercellular space. They noted ferritin in blood and lymphatic capillaries within the lamina propria also. The free movement of ferritin through the cytoplasm and lateral cell membrane is open to question on the grounds of inadequate fixation. Their results, however, seemed to be substantiated by immuno-logical data. Using a double-diffusion technique with an Ouchterlony plate they obtained a precipitin line between ferritin antibody and sera from young rats that had received ferritin solution 60–90 minutes previously by intraduodenal injection. In early pilot experiments of the present study, ferritin was administered t young rats by injecting the solution di rectly through the anterior abdominal wal into the stomach, which, in the sucklin rat, was full of milk and readily visualized However, when the abdomen was opene to remove a sample of intestinal tissu it was obvious that a seepage of ferriti had occurred through the needle hole int the peritoneal cavity, even though the i jection was made with a 30-gauge needl To avoid such leakage the gavage metho was subsequently employed. It seems po sible that a similar leakage could occu from the duodenum particularly durin the pumping action of peristaltic mov ments. Therefore, the intestinal uptake o ferritin reported by Kraehenbuhl et a ('67) may actually represent absorption o ferritin from the peritoneal cavity via di phragmatic lymphatics.

Transmission of proteins from intestin lining cells to the circulation of young ra is a highly selective process. Apparentl these cells can distinguish between vario classes of serum proteins (Bangham ar Terry, '57) as well as homologous ar heterologous proteins. When immune se are prepared from a single antigen in d ferent species and fed separately to suc ling rats, the highest titer of circulatin antibody is attained with homolgou serum, while a lower titer is noted wi heterologous sera from more distally r lated species (Halliday, '57). This ar other related evidence suggested that th structure or configuration of a protein important in the process of selective tran mission. It has been demonstrated th fragments of pepsin refined (Brambel Hemmings and Oakley, '59) or papain d gested antibodies (Brambell, Hemming Oakley and Porter, '60) are transmitted t the circulation at quite different rate Fragments I and II, produced by Papai digestion (Porter, '58, '59) contain th antibody reactive sites but are not tran

-ied to the circulation whereas Frag-
-nt III, the antigenic part of the antibody
-ansmitted readily (Brambell et al., '60).
-mbell, Halliday and Morris ('58) have
-tulated that the mechanism of selection
-y be related to receptor sites which act
-a receptor-substrate system recognizing
-pecific part of the protein. Pierce and
-ith ('67a) have noted that interpreta-
-n of some of the studies by Brambell
-al. is complicated by the fact that their
-um assays detected only complete se-
-m agglutinins and did not account for
-omplete antibodies. This is particularly
-nificant in light of recent findings by
-orris ('65) demonstrating incomplete ag-
-tinins in the sera of young rats after
-ding heterologous immune sera. Similar
-ta were obtained by Pierce and Smith
-67a) using newborn pigs. In view of
-se facts, some reassessment of the
-echanism of selective transmission of
-oteins seems necessary; previous experi-
-ents should be repeated using more re-
-ned immunological techniques.

Molecular weights and size of proteins
-ave been discounted in the past as a de-
-rminant of the transmissibility of pro-
-ins (Brambell, '58, '66; Hemmings and
-ones, '62). However, its significance may
-e more important than previously real-
-ed. For example, a very small protein
-uch as bovine insulin, 36,000 M.W. in
-exameric form, is transmitted to the cir-
-ulation of suckling mice (Kelly, '60) and
-ats (Mosinger, Placer and Koldovsky,
-59) in sufficient quantities to induce hypo-
-lycemia. But very large proteins, IgM
-mmunoglobulins, 10^6 M.W. and ferritin,
-50,000 M.W. are not transmitted (Morris,
-65; Graney, '64).

Besides some of the sophisticated im-
-munological and *in vitro* techniques now
-available, recent advances have been made
-n tagging antibodies with electron opaque
-markers. This method should allow one to
-follow the pathway of discharged proteins
-at the ultrastructural level. The evidence
-at present, though purely circumstantial,
-indicates that proteins are discharged at
-the lateral cell membrane, pass through
-the intercellular space and enter the lam-
-ina propria. A similar pathway has been
-described for the yolk sac (Padykula,
-Deren and Wilson, '66). Amorphous precipitated material, identical to the contents of the supranuclear vacuole and blood capillaries of the lamina propria, is often observed in the intercellular spaces of the ileum during suckling, but it is not observed in the jejunum. This material is not observed in the intercellular spaces of the jejunal or ileal epithelium of the adult rat. There is, however, a similar precipitated material in the intercellular spaces of intestinal epithelium in humans who have certain protein losing enteropathies, e.g., intestinal lymphangiectasia (Dobbins, '66).

Digestion and fate of ferritin and colostral protein

There are at least two enzymatic environments which ingested proteins must survive before reaching the circulation, the first is in the lumen of the gastrointestinal tract and the second is in the ileal lining cell itslef. Proteolytic activity in the small intestine of newborn rats in high, and does not differ significantly from the activity in adult intestine (Noack, Koldovsky, Friedrich, Heringova, Jirsova and Schenk, '66). Survival of proteins in the intestinal lumen is apparently related to a low activity of digestive enzymes in the stomach. Mosinger et al. ('59) have shown that the proteolytic activity of the gastric contents is low *in* newborn rats, but increases significantly after day 21. These changes correspond with the morphological development of the parietal and chief cells in the gastric glands (Manville and Lloyd, '32; Kammeraad, '42; Hill, '56).

Acid phosphatase and other hydrolytic enzymes have been localized histochemically in the supranuclear vacuole of ileal lining cells (Cornell and Padykula, '65; Straus, '65; Shervey, '66) which would indicate an active lysosomal activity in these cells. It should be mentioned from a comparative standpoint, an apical tubular apparatus with an esterase positive vacuolar system and a second vacuolar system with both esterase and acid phosphatase activities is found in other cell types besides the ileal lining cell, e.g., cells of the proximal convoluted tubule of the kidney (Straus, '64) and yolk sac endoderm (Padykula, '58; Sorokin and Padykula, '64; Lambson, '66). It is not known whether all proteins

must pass through the supranuclear vacuole prior to discharge, or if transmitted proteins bypass the vacuole. It is known that only about 10% of ingested proteins are actually transmitted across the epithelium (Morris, '65; Bamford, '66; Pierce and Smith, '67a,b); the remaining presumably are degradated by the lining cells.

It has been suggested (Brambell et al., '58; Leissring and Anderson, '61; Anderson, '65; Morris, '64; Brambell, '66) that the protein distributed about the periphery of supranuclear vacuoles, may be adsorbed or bound to the limiting membrane of the vacuole. It was further speculated that the binding of the protein may in some way protect it from the proteolytic activity in the vacuole. In the present study there was no evidence to indicate specific binding of ferritin to either the surface membrane or the limiting membranes of the tubular system or supranuclear vacuole. Using the tetrad structure of the micellar core of ferritin as a criterion of its integrity, there was no evidence of ferritin degradation 18 hours after feeding. This is not a good parameter, however, since all ferritin molecules are not necessarily oriented favorably to visualize the micelles. In addition, Matioli and Baker ('63) have described a ferritinic form of hemosiderin as an intermediate in the degradation of ferritin to nonferritinic hemosiderin. The half-life of intestinal lining cells in the suckling animals is about 20% longer than in the adult (Koldovsky, Sunshine and Kretchmer, '66) so that considerable intracellular digestion of protein may occur before the cells are sloughed into the intestinal lumen.

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Note added in proof: Since this manuscript was submitted for publication, two significant review articles related to this topic have appeared in *The Handbook of Physiology*, Section 6: Alimentary Canal, Volume III. Intestinal Absorption, American Physiological Society, Washington, D. C., 1968. See J. J. Deren, page 1099 and I. G. Morris, page 1491.

PLATE 1

EXPLANATION OF FIGURES

The following six figures are photomicrographs of the small intestine from three different animals: newborn rat prior to suckling, nine-day-old suckling rat and a ten-day-old rat fasted for 18 hours prior to sacrifice. Each pair of photographs represent Epon sections of jejunal and ileal tissue taken from the same animal and stained with toluidine blue. Magnification in figures 1–6 is the same, × 1744.

1 Jejunum of newborn rat prior to suckling. There are no obvious inclusions visible in the cytoplasm.

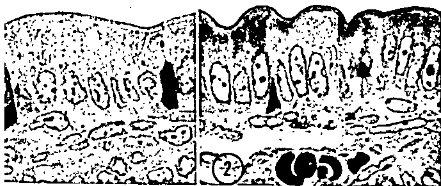
2 Ileum of newborn rat prior to suckling. Morphological pattern of the ileum not significantly different from jejunum (fig. 1).

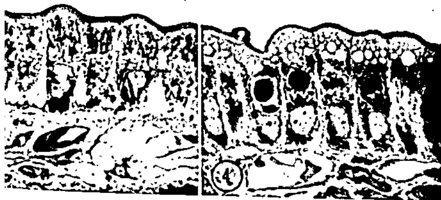
3 Jejunum of nine-day-old suckling rat. Lipid droplets are observed in Golgi apparatus, intercellular spaces and lamina propria.

4 Ileum of nine-day-old suckling rat. Numerous clear vacuoles are noted in the apical cytoplasm. Supranuclear area occupied by large vacuole containing amorphous dense material.

5 Jejunum of ten-day-old suckling rat fasted for 18 hours prior to sacrifice. At this stage, cytoplasm is completely clear of lipid material. Typical fasting state.

6 Ileum of ten-day-old suckling rat fasted for 18 hours prior to sacrifice. Apical vacuoles and a supranuclear vacuole are still observed in the cytoplasm of the lining cells.


2


4

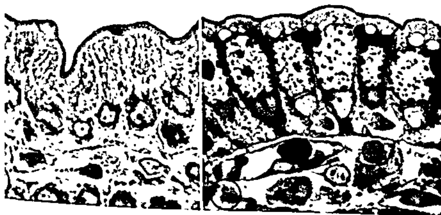


PLATE 2

EXPLANATION OF FIGURES

7 Jejunum of 15-day-old suckling rat fasted for 18 hours prior to sacrifice. Electron micrograph showing mitochondria, membranes of the smooth- and rough-surfaced endoplasmic reticulum. Note absence of any extensive plasmalemmal invaginations. × 29,000.

8 Jejunum of 15-day-old suckling rat, two hours after receiving 1 ml of corn oil. Osmiophilic droplets, presumably lipid, are observed in both smooth- and rough-surfaced endoplasmic reticulum. × 25,000.

TIN UPTAKE BY ILEAL LINING CELLS
O. Graney

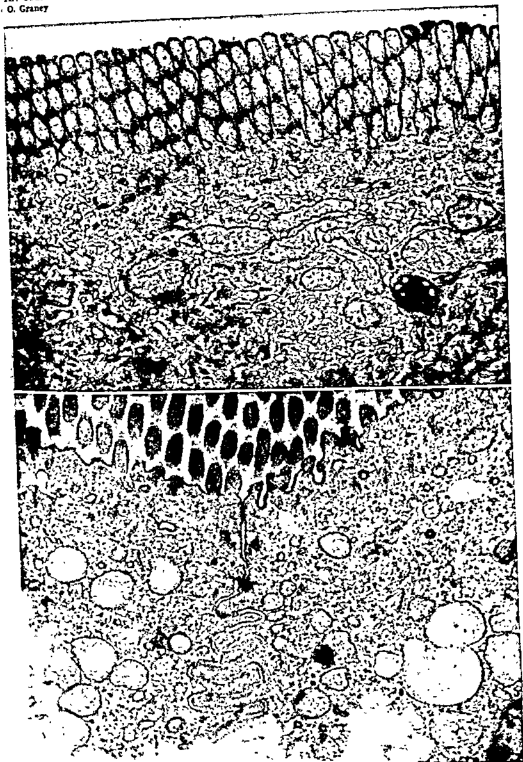


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SNV

e Uptake of Ferritin by Ileal Absorptive Cells in ckling Rats. An Electron Microscope Study [1]

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ABSTRACT An electron microscopic study of the small intestine of rats from birth to 15 days of age has demonstrated a gradient of both structure and function along the length of the small intestine. During this period of development, only the ileum was found to absorb intact exogenous ferritin or colostral proteins. In newborn rats ferritin particles or milk proteins are incorporated by ileal absorptive cells into tubular invaginations of the apical plasmalemma which connect with vacuoles or cisternae in the apical cytoplasm. The ferritin particles are then transported by another vacuolar system to the supranuclear region where several such vacuoles coalesce to form a single supranuclear vacuole. After the first feeding the supranuclear vacuole is established in the ileal cells so that additional ingested proteins, whether ferritin or from milk, are transported from the cisternae directly to the supranuclear vacuole. There was no evidence in the present study to indicate that ferritin is transmitted from the ileal cell to the circulation. The subject of protein transport by the intestinal epithelium is reviewed briefly.

The mucosa of the ileum in neonatal ıgulates, rodents, and carnivores differs markably from the adult ileum in its orphological (Clark, '59; Kraehenbuhl, loor and Blanc, '66), cytochemical (Corell and Padykula, '65), and physiological aracteristics (Brambell, '58, '66). Ileal bsorptive cells from neonatal animals of ıese groups can incorporate ingested proein and transfer certain immune globu- from the absorptive cell into the cir-tion. This is an important mechanism passive immunization of the newborn. laternal antibodies secreted into the co- and milk can be absorbed and tilized by the suckling animal until active ntibody synthesis is established. The in- and transport of protein by the leum of suckling rats ceases about three after birth when there is a transi-on to the typical adult type (Halliday, , Clark, '59). In some animals from above species, particularly the rat and there is also a prenatal transfer of via the yolk sac and intestine. his is not an important pathway quanti-ively in these animals, but in the rabbit monkey prenatal transfer of proteins ia the yolk sac and placenta, respectively, the principal means of passive immunization.

In those species which absorb intact proteins during the suckling period, the fine structure of the intestinal absorptive cells in the ileum is distinguished from adult absorptive cells by the presence of many small apical cytoplasmic vesicles and a large supranuclear vacuole (Möllendorff, '24; Smith, '25; Comline, Roberts and Titchen, '51b; Comline, Pomeroy and Titchen, '53; Hill and Hardy, '56; Clark, '59; Payne and Marsh, '62; Anderson, '63; Graney, '64; Cornell and Padykula, '65; Kraehenbuhl et al., '66). Clark ('59) studied the uptake of proteins during the early postnatal period in rats and mice by examining the passage of gamma globulins and electron-dense particles across the intestinal mucosa. Three hours after feeding bovine gamma globulins he was able to localize them with a fluorescein-labeled antibody in the supranuclear inclusions of absorptive cells in the midportion of the small intestine and in associated lacteals and mesenteric lymph nodes. Electron microscopic studies with colloidal tracers revealed the uptake of gold and saccharated iron oxide particles into apical cytoplasmic

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Preparation of tissues

.he animals were anesthetized by an ... injection of nembutal (0.04 gm body weight) and a laparotomy performed to expose the intestinal t. Small samples of the proximal jeju- and terminal ileum were excised, cut small blocks and immediately placed fixative.

The fixatives used were 3% glutaralde- e in 0.1 M sodium cacodylate or phos- ite buffer (Millonig, '62) and 1% osmi- tetroxide in phosphate buffer (Millonig,) for one to two hours prior to dehydra- n. The tissues were either rapidly de- drated in graded alcohols or, in some es, were dehydrated in acetone and ined in the block with 0.5–1% po- sium permanganate according to the thod of Parsons ('61). After dehydra- n the tissues were embedded in Epon Araldite (Luft, '61). Thick sections (1– μ) were cut on an ultramicrotome and amined either by light microscopy after aining with toluidine blue or by electron icroscopy. Thin sections were mounted n copper grids with a substrate consisting f either formvar or celloidin and a carbon lm and were doubly stained with uranyl cetate (Watson, '58) and alkaline lead alts (Ryenolds, '63; Venable and Cogges- all, '65).

OBSERVATIONS

Regional variations in the gross morphology of the small intestine

The duodenum and jejunum in the nor- suckling animal can be distinguished from the ileum by their creamy white color contrast to the brownish-yellow or am- er color of the ileum (Möllendorff, '24). he distinctive regional coloration of the l intestine corresponds with regional ferences in absorptive function of the intestine that are observed after ing suckling animals ferritin or col- tracers, e.g., carbon or trypan blue. After young rats are gavaged with a so- of ferritin, there is no change in the color of the jejunum when exam- ined at intervals of 0.5, 1, 2, 4, 6 or 24 hours. When the intestine is opened, the presence of the tracer is revealed by a reddish-brown color in the lumen of the jejunum 30 minutes after feeding, but all of the tracer can be washed away from the mucosal surface by gentle rinsing in saline. In contrast, one hour after feeding ferritin the normal amber color of the upper ileum changes to an intense reddish brown. By two hours after gavage, the ferritin solution completely traverses the small intestine, and the entire ileal mucosa is a reddish-brown color. This is in striking contrast to the pale jejunal mucosa. If other tracers are used, such as trypan blue, hemoglobin or carbon, the amber color of the ileum becomes masked by the color of the tracer. None of the tracers used could be washed from the ileal mucosa with saline rinses, and the pigmentation was still present 24 hours after feeding (the longest time point studied).

These simple macroscopic observations suggest that the jejunum is not important in the uptake of ferritin or particulate tracers. However, light and electron microscopic studies of this area were undertaken in order to verify these conclusions and to compare the cytological features of jejunal and ileal mucosa at various times and under both conditions of fasting and suckling.

Light microscopy

Jejunum. The cytological detail of columnar absorptive cells in the jejunal mucosa of newborn rats (prior to suckling) and fasting young rats is very similar to that of the adult. The apical cytoplasm in all of these cells has a coarse granularity which is probably due to the presence of mitochondria (figs. 1, 3, 5). Particulate inclusions are not prominent. However, in tissues fixed within 30 minutes after young rats are allowed to suckle, the jejunal absorptive cells contain variable amounts of lipid which is distributed in droplets throughout the apical cytoplasm and in the Golgi apparatus (fig. 3). Four hours after suckling, most of the jejunal cells have been cleared of obvious fat droplets (fig. 5).

The initial uptake of lipid by the intestinal mucosa occurs in cells located on the apices of the intestinal villi and later by cells situated along the sides and base of the villi as described previously in the

tubules, membrane-limited vesicles and the supranuclear vacuole. These findings were interpreted as evidence for pinocytotic activity in these cells. It was noted that the colloidal particles appeared to be retained by the lining cells, while some of the gamma globulins were discharged into the lamina propria. It has been shown by others (Comline, Roberts and Titchen, '51a; Hill and Hardy, '56; May and Whaler, '58; Pierce and Johnson, '60) that proteins are usually transported from the lamina propria by lymphatic rather than blood capillaries. More recently, it has been shown that proteins of 70,000 M.W. or less are absorbed via portal blood (Balfour and Comline, '59; Pierce, Risdall and Shaw, '64).

In the current study, ferritin, a naturally occurring electron-dense protein, was introduced into the gastrointestinal tract of suckling rats in order to study further the absorption of intact protein at this stage of development. In particular, it was of interest to determine whether this tracer protein would be transferred from the intestinal lining cells to the lamina propria and thus establish the successive steps in the pathway of protein absorption.

MATERIALS AND METHODS

Animals and experimental grouping

Long-Evans and Charles River rats from newborn to 15 days of age were divided into the following groups:

Controls. (1) Newborn rats separated from the mother prior to suckling, (2) newborn rats separated from the mother 0.5, 1 and 2 hours after suckling, and (3) young rats, 4–15 days of age, separated from the mother 0.5, 1, 2, 3, 4, 6 and 8 hours after suckling. Some of these animals were maintained in a fasting state (no solid or liquid food) for periods up to 24 hours.

Lipid studies. Two 15-day-old rats were separated from the mother and given 1 ml of Mazola corn oil via a gastric tube. The animals were subsequently killed after intervals of one and two hours respectively.

Ferritin studies. Ferritin was administered to newborn rats prior to suckling and to suckling rats up to 15 days of age. A small polyethylene tube (P.E. 10–50) was passed into the esophagus of an unanes-thetized animal and 0.1 or 0.2 ml of a dia-lyzed ferritin solution was injected from a tuberculin syringe attached to the tubing.

Preparation of gross specimens

Five 5-day-old suckling rats were used for a special macroscopic study of the small intestine. One animal was a normal suckling control and the remaining four each received one of the following solutions by gastric tube: (1) 0.2 ml of a 10% ferritin solution, (2) 0.5 ml of a [illegible] blue solution, (3) 0.2 ml of a 10% hemoglobin solution, and (4) 0.3 ml of an India ink solution. The animals were killed [illegible] hours after gavage and the entire intestine from gastroduodenal junction to caecum was excised and placed in saline. A pair of fine scissors was used to open the intestine and expose the mucosa. The specimen was then rinsed in several changes of physiological salt solution and fixed *in toto* in 10% calcium-formol.

Dialysis procedure for ferritin solution

The ferritin solution used in this study was obtained from either of two commercial sources (Nutritional Biochemical Corporation, Cleveland, Ohio; Pentex Incorporated, Kankakee, Illinois). Both commercial preparations of ferritin, approximately 10 gm% in concentration, were similarly crystallized from extracts of horse spleen with cadmium sulfate. It was, therefore, necessary to dialyze the ferritin solution against a chelating agent, ethylene-di-nitrilo-tetraacetic acid (EDTA), to remove toxic cadmium contaminants (for details see Farquhar and Palade, '61). The ferritin solution was then rendered isotonic by a subsequent dialysis against Gey's balanced salt solution for 72 hours with three changes (Wissig, '64).

The final dialyzed solution of ferritin was passed through a Millipore filter (0.45 μ) and refrigerated in a sterile rubber-stoppered serum bottle. Under these conditions the ferritin solution may be stored for at least 6 months without any significant deterioration. Small quantities may be removed as needed with a sterile syringe and needle.

Karlin, '59b; Cardell et al., '67) to ant discussion here.

though the jejunal cells of suckling absorb lipid in quantity, the present itigation has not demonstrated the capacity of these cells to absorb *protein.* Ferritin was not observed in the jejunal cells, ny time interval, after it was administered to either fasting or suckling rats, *though its presence in the jejunal* en could be verified by the color of the tents at the time of autopsy.

Ileum. There were no obvious differences between jejunal and ileal lining cells newborn rats (prior to suckling) when lied by light microscopy, but an electron microscopic examination reveals a damental difference in their ultrastructure. The plasmalemma in the intermicrovillus spaces of ileal absorptive cells forms aginations extending for some distance o the apical cytoplasm (figs. 10, 11). ese invaginations often branch and anastomose with others from neighboring ermicrovillus spaces as they penetrate e cytoplasm. The resulting labyrinth of *terconnected tubules is confined to* a nd or zone across the apical part of the ll, intermingled with mitochondria and ements of the endoplasmic reticulum figs. 10, 11). Ultimately, the tubules terminate in a vacuolar structure or perhaps better term is cistern, since there are acuoles of similar size which apparently o not have tubular connections (figs. 10, 1, 14). Within the lumen of these tubules nd cisternae there is a fine, flocculent naterial, but prior to suckling the quantity f this material is not significant. After uckling, however, the dominant feature of hese cells is the abundance of this flocculent material throughout the tubular labyrinth and within the cisternae. This substance is similar to that described by Clark '59) and is a proteinaceous material, presumably derived from the milk and *succus nterius.* In addition, large vacuoles without obvious tubular connections contain his material also and appear to coalesce, orming the single, large vacuole in the supranuclear region described earlier in his paper. An amorphous material identical to that in the supranuclear vacuole was noted also in the intercellular space (fig. 3). The supranuclear vacuole persists in ileal absorptive cells, at least in those on the upper *three-fourths of the villus,* even when the animal is fasted for 18–24 hours (fig. 6). With the normal feeding cycle of the young, this means that supranuclear vacuoles *are present in the ileum from the* first meal until about three weeks after birth when the ileal cells are gradually replaced by cells typical of the mucosa of the adult. *After fasting the cells on the* basal part of the villus resemble the ileal cells from newborn animals, i.e., they contain a tubular apparatus with its associated *cisternae but lack supranuclear* vacuoles. The flocculent material within the tubular apparatus is sparse. Presumably, these are differentiating cells which have *migrated up the villus during the fasting* period.

After ferritin gavage, individual ferritin molecules were identified in the intestinal *lumen, intermicrovillus spaces,* and within the invaginations of the apical plasmalemma of ileal lining cells (figs. 11–14). Within 1.5 hours after administration of ferritin the tubular apparatus contains the ferritin tracer, as do the associated vacuoles or cisternae (figs. 13, 14). Vacuoles without associated tubules are predominant in the supranuclear area and contain ferritin in high concentration also. It is not clear whether the smooth-surfaced vacuoles bud off from the cisternae or represent cisternae which have lost their tubular connections. In any event, these vacuoles migrate to the supranuclear area where they coalesce, forming a single large vacuole (fig. 14). The progressive increase in size of this vacuole, as successive ferritin-filled vacuoles migrate from the apical area and fuse with it, ultimately results in formation of the definitive supranuclear vacuole. *The nucleus is gradually displaced* basally to accommodate the expanding vacuole.

On occasion, ferritin particles are seen in crystalline patterns within the smooth-surfaced vacuoles adjacent to the supranuclear vacuole as well as in the supranuclear vacuole itself. More often, the content of the tubular apparatus, apical vacuoles, and supranuclear vacuoles is heterogeneous, consisting of both ferritin particles and a fine, flocculent material

adult rat (Hewitt, '54; Palay and Karlin, '59b; Ladman, Padykula and Strauss, '63).

The uptake of milk lipid by the jejunal mucosa in ferritin-fed animals appeared to be the same as in normal control animals. There was no evidence of ferritin uptake by the jejunal absorptive cells, e.g., dense vacuoles; and jejunal tissue from either ferritin-fed or control animals could not be distinguished by light microscopic study.

Ileum. The appearance of ileal absorptive cells in the newborn rat prior to suckling (fig. 2) is similar to the jejunal absorptive cells in the same animal. In some young, the ileal absorptive cells contain a few small vesicles in the apical cytoplasm, deep to the area of the terminal web. Their number was variable, but in no case were there as many vesicles in the newborn as in the suckled or ferritin-fed animals.

After the young have suckled, the appearance of the ileal absorptive cells changes conspicuously, acquiring a unique character which distinguishes them from other absorptive cells in the proximal part of the intestine. A large vacuolar structure appears in the supranuclear area, displacing the nucleus toward the base of the cell (figs. 4, 9). In the suckling pig, a large cytoplasmic vacuole is formed at the base of the absorptive cell and the nucleus assumes an apical position (Payne and Marsh, '62; Mattisson and Karlsson, '66; Sibalin and Björkman, '66; Vodovar and Fléchon, '66). The contents of the vacuoles vary. They may contain either homogeneous dense material, clumps of dense material aggregated about the periphery of the vacuole; or their contents may appear as a meshwork of finely vacuolated, flocculent material. The size of the vacuole varies depending upon whether it is located in an absorptive cell on the apex or base of a villus. The largest vacuoles were usually located in cells on the apex of the villus, with a gradation in size to the smallest located in absorptive cells on the base of the villus. Cells in the crypts of Lieberkühn do not contain supranuclear vacuoles. The apparent local absorption gradient located in an apico-basal direction along the height of the villi is reminiscent of that observed during absorption of fat in the jejunum of suckling young and that reported for the adult jejunum during fat absorption (Hewitt, '54; Palay and Karlin, '59b). The vacuole persists in the supranuclear region even when the animal is fasted for periods of up to 24 hours (fig. 6).

Fine structure

The fine structure of intestinal epithelial cells of suckling rats has been described previously (Clark, '59; Kraehenbuhl et al., '66; Dunn, '67). However, some structural feautres have not been considered heretofore and, in particular, the cytological differences between the proximal half of the small intestine (duodenum and jejunum) and the distal half (ileum) have not been emphasized. These differences which are not apparent in the adult, represent a longitudinal differentiation of ultrastructure in the small intestine of the suckling rat and presumably underlie the specialized absorptive function of the proximal and distal small intestine.

Jejunum. The fine structure of jejunal absorptive cells from either newborn rats (prior to suckling) or suckling rats (which are subsequently fasted) is indistinguishable and is comparable to the fine structure of jejunal absorptive cells in fasted adult rats (Palay and Karlin, '59a; Cardell, Badenhausen and Porter, '67).

When jejunal absorptive cells of young rats were examined 30 minutes to 2 hours after suckling, osmiophilic droplets, presumably lipid, were observed distending the endoplasmic reticulum of the apical cytoplasm. At later periods, the Golgi vesicles and intercellular spaces become distended with lipid droplets, appearing finally in the lamina propria. The matrix of these lipid droplets often appears extracted and sometimes only a thin shell of dense material remains. This was a more consistent finding after glutaraldehyde fixation than with osmium fixation. In an attempt to produce better visualization of the pathway of lipid through the jejunal cell, Mazola corn oil was given by gastric tube to two suckling rats and their jejunums subsequently fixed in osmium (fig. 8). The pathway of the lipid through the jejunal cells appears to be the same whether the animal is fed milk or corn oil. In either case, it is not sufficiently different from the studies of adult jejunum (Pala

l Karlin, '59b; Cardell et al., '67) to rrant discussion here.

... the jejunal cells of suckling s absorb lipid in quantity, the present estigation has not demonstrated the ca- ... these cells to absorb protein. Fer- was not observed in the jejunal cells, any time interval, after it was admin- ... to either fasting or suckling rats, en though its presence in the jejunal nen could be verified by the color of the ... at the time of autopsy.

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After ferritin gavage, individual ferritin molecules were identified in the intestinal lumen, intermicrovillus spaces, and within the invaginations of the apical plasma- lemma of ileal lining cells (figs. 11–14). Within 1.5 hours after administration of ferritin the tubular apparatus contains the ferritin tracer, as do the associated vacu- oles or cisternae (figs. 13, 14). Vacuoles without associated tubules are predomi- nant in the supranuclear area and contain ferritin in high concentration also. It is not clear whether the smooth-surfaced vacu- oles bud off from the cisternae or represent cisternae which have lost their tubular connections. In any event, these vacuoles migrate to the supranuclear area where they coalesce, forming a single large vacu- ole (fig. 14). The progressive increase in size of this vacuole, as successive ferritin- filled vacuoles migrate from the apical area and fuse with it, ultimately results in for- mation of the definitive supranuclear vac- uole. The nucleus is gradually displaced basally to accommodate the expanding vac- uole.

On occasion, ferritin particles are seen in crystalline patterns within the smooth- surfaced vacuoles adjacent to the supra- nuclear vacuole as well as in the supra- nuclear vacuole itself. More often, the content of the tubular apparatus, apical vacuoles, and supranuclear vacuoles is heterogeneous, consisting of both ferritin particles and a fine, flocculent material

similar to that observed in suckling controls.

The successive steps in the formation of the supranuclear vacuole described above pertain to the newborn rat and the changes which occur after the first meal, whether it consists of milk or ferritin. In older suckling animals, the supranuclear vacuole is already established, so that when additional protein is ingested and taken into the apical tubules and cisternae, the smooth-surfaced vacuoles shuttle the protein directly to the existing supranuclear vacuole. In this study, the supranuclear vacuole appears to represent the terminus of the absorptive pathway for ferritin in the intestinal epithelial cell. It was not observed in association with the Golgi apparatus and there was no evidence to indicate that ferritin was transported from the lining cells to the intercellular space or lamina propria. This is in disagreement with the report of Kraehenbuhl, Gloor and Blanc ('67). A few particles were observed free in the cytoplasm, but these might represent endogenous ferritin or ferritin artifactitiously released from the tubules and vacuoles. The use of glutaraldehyde fixation proved crucial in this regard. Multiple attempts with osmium tetroxide as a primary fixative in combination with a spectrum of buffer systems, consistently resulted in the diffuse distribution of ferritin molecules throughout the cytoplasm of jejunal absorptive cells, the intercellular space, and the lamina propria. Ferritin was retained within membrane-limited systems only when glutaraldehyde was used as the primary fixative.

DISCUSSION

Morphological and functional gradients in the small intestine

The existence of an apico-basal gradient in the structure and function of an adult intestinal villus has been recognized for some time (review by Padykula, '62). The results of the present study have emphasized a longitudinal or proximo-distal gradient in the ultrastructural characteristics of the intestinal lining cells of suckling rats which is related to regional differences in their specific absorptive functions.

The proximal part of the small intest
actively absorbs lipid, but it does not ab-
sorb intact protein. This interpretation i
supported by the observation that neith
ferritin nor aggregates of amorphous pr
cipitated material originating from th
milk were identified in lining cells of th
duodenum or jejunum of the young ra
Moreover, there is no tubular apparatus i
the apical cytoplasm of jejunal lining ce
comparable to that found in the ileum, a
therefore, the jejunum appears to lack th
morphological apparatus necessary for ab
sorption of intact proteins.

In two papers reporting ferritin abso
tion by the jejunum of adult rats (Card
et al., '67; Casley-Smith, '67) the expe
ments were done using ligated intesti
segments and a much higher concentratio
of ferritin than the one employed in th
study. The quantity of ferritin taken u
under these conditions was very smal
compared with that absorbed by the ileu
of suckling rats; therefore, the ability
the jejunum to absorb significant amoun
of intact protein under normal conditio
remains to be demonstrated. The ileum
the suckling rat absorbs protein primari
but it can absorb lipid also. Cornell a
Padykula ('65) have shown that the ileu
of the suckling rat can absorb an emul
fied lipid when it is injected into the inte
tinal lumen. From the observations in t
present study and those of Cornell a
Padykula ('65), the apical tubular app
ratus and supranuclear vacuole do n
play an obvious role in the absorption
lipid, at least pariticulate lipid. In view
the current concepts of lipid absorpti
(Cardell et al., '67); Strauss and Ito, '6
it is conceivable that free fatty acids
other soluble lipids enter the tubular la
rinth where they diffuse into the api
cytoplasm and subsequently become este
fied in the endoplasmic reticulum. Und
normal physiological conditions, howev
it appears that only small quantities
lipid reach the distal part of the small
testine because of the efficiency with wh
lipid is absorbed in the proximal part.
addition to the differences in structure a
function described above, there are va
ations in the enzymatic activities betwe
the proximal and distal small intesti
which is further evidence of a line

ctional gradient; both β-glucuronidase eringova, Jirsova and Koldovsky, '65) d β-galactosidase (Koldovsky and Chytil, 5) activities are higher in the ileum than the jejunum of suckling animals. Acid osphatase activity is greater in the ileum d has been demonstrated by histochemi- means to be in association with the pranuclear vacuole and adjacent small cuoles (Cornell and Padykula, '65; Sher-y, '66).

Alkaline phosphatase activity, however, greater in the duodenum than in either e jejunum or ileum (Moog, '51, '62; Kol-vsky, Faltova, Hahn and Vacek, '61). It interesting to note that whereas alkaline osphatase activity increases after day , β-glucuronidase and β-galactosidase ac-ities decrease. All of the changes in the tivities of the enzymes mentioned above n be delayed by adrenalectomy or in-uced precociously by administration of ogenous cortisone (Moog, '62; Koldovsky d Chytil, '65; Heringova et al., '65). It ould be emphasized that in addition to gional qualitative differences in the mor-hological, enzymatic, and functional fea-ures which distinguish the jejunum and eum, there are also quantitative changes ccurring within each of these regions uring the course of the suckling period. n the rat, this period is 21 days, but in ther animals it may be considerably less. hese points have not always been appre-iated by some investigators, resulting in rroneous generalizations about intestinal istophysiology.

Mechanism of protein uptake by intestinal epithelial cells

Schechtman ('56) has emphasized that here are two distinct steps in the move-nent of proteins across a cellular layer. The first step, in the case of the intestinal pithelial cell, is the uptake of the protein t the surface of the cell and the second s the discharge of protein from the lateral and/or basal surface of the cell. The tacit assumption is that proteins cannot enter he lamina propria by passing directly hrough the intercellular spaces. There is no evidence at present for the intercellular passage of protein through the intestinal epithelium. There is, however, both physio-logical and immunological data (Brambell, '58, '67; Clark, '59) which show that proteins are incorporated nonselectively into intestinal lining cells of certain suckling animals. A variety of proteins and colloidal particles can be incorporated into the cell, e.g., albumin, gamma globulin, saccharated iron oxide (Clark, '59), ferritin (Graney, '64, '65; Kraehenbuhl et al., '67) and peroxidase (Straus, '65), but only specific proteins are discharged from the cells. The mechanism of this discharge will be discussed in the following section.

The rat is born with an antibody titer approximately one-sixteenth that of the maternal serum, but by the second or third day after birth it is equal to or exceeds that in the maternal serum (Halliday, '55b). During the postnatal period, the rat continues to absorb intact antibodies until day 20, although the quantity absorbed begins to decrease after day 15 (Culbertson, '39; Halliday, '57; Clark, '59).

The rate of transfer can be quite rapid; measurable quantities of antibody appear in the circulation of the young rat within 30 minutes after feeding immune serum. By three hours the circulating titer has attained a maximum concentration (Halliday, '55a).

Clark ('59) first suggested that protein was incorporated into the intestinal epithelial cells of suckling rats and mice by the process of pinocytosis. This concept has been reiterated for the suckling puppy (Rubin, '66), piglet (Mattisson and Karlsson, '66; Sibalin and Björkman, '66; Vodovar and Flechon, '66) and rat (Kraehenbuhl et al., '67). The morphological observations of the present study do not seem to be completely consistent with this concept. Ferritin particles enter the apices of the lining cell through a tubular labyrinth and not by incorporation into vacuoles resulting from folding of the surface membrane as in the original concept of pinocytosis (Lewis, '31). Similarly, there was no pinocytotic uptake of ferritin *in quania* (Palade, '53) by vesicles of either the smooth-surfaced or coated variety, although this phenomenon has been observed in several other systems (Wissig, '58; Fawcett, '65).

An alternative viewpoint is that the apical tubules and their cisternal terminals are a relatively stable system within the

apical cytoplasm and that the lumen of this system is confluent with the intestinal lumen, i.e., extracellular. This would provide an access for nonspecific diffusion of protein, or other substances for that matter, from the intestinal lumen through the apical tubules and into the cisternae or vacuolar terminals of the tubules. The precise means by which protein moves from the cisternae to the supranuclear vacuole remains elusive. From Clark's studies ('59) and the results obtained with ferritin there is little doubt that the nonselective incorporation of protein proceeds past the point of the cisternae and as far as the supranuclear vacuole. Whether this occurs by a pinching off of small vesicles from the cisternae or from a separate vacuolar system is unknown. There is histochemical evidence which supports to some extent the concept of two separate membrane systems. Cornell and Padykula ('65) have shown that the apical cisternae (apical droplet in their terminology) are rich in esterase, whereas the supranuclear vacuole and adjacent small vacuoles contain both acid phosphatase and esterase activities. Since the supranuclear vacuole is formed in newborn young by the successive coalescence of vacuoles in the supranuclear region, some form of a vesicular transport system must be operating.

Selective discharge of protein from intestinal lining cells

An electron microscopic examination of the ileum from at least ten rats removed at intervals varying from 30 minutes to 24 hours after feeding ferritin has not demonstrated the passage of ferritin beyond the point of the supranuclear vacuole. This disagrees with the results of Kraehenbuhl et al. ('67) who described the movement of ferritin to the basal area of the supranuclear vacuole where some of the particles were incorporated into Golgi vesicles while others appeared to migrate freely through the cytoplasm and lateral cell membrane into the intercellular space. They noted ferritin in blood and lymphatic capillaries within the lamina propria also. The free movement of ferritin through the cytoplasm and lateral cell membrane is open to question on the grounds of inadequate fixation. Their results, however, seemed to be substantiated by imm[illegible] logical data. Using a double-diffusion t[illegible] nique with an Ouchterlony plate they [illegible] tained a precipitin line between fe[illegible] antibody and sera from young rats t[illegible] had received ferritin solution 60–90 m[illegible] utes previously by intraduodenal inj[illegible] tion. In early pilot experiments of [illegible] present study, ferritin was adm[illegible] young rats by injecting the solution [illegible] rectly through the anterior abdominal w[illegible] into the stomach, which, in the [illegible] rat, was full of milk and readily visualize[illegible] However, when the abdomen was open[illegible] to remove a sample of intestinal tiss[illegible] it was obvious that a seepage of ferri[illegible] had occurred through the needle hole in[illegible] the peritoneal cavity, even though the [illegible] jection was made with a 30 [illegible] To avoid such leakage the gavage me[illegible] was subsequently employed. It [illegible] sible that a similar leakage could [illegible] from the duodenum particularly during the pumping action of peristaltic move-ments. Therefore, the intestinal uptake of ferritin reported by Kraehenbuhl et al. ('67) may actually represent absorption of ferritin from the peritoneal cavity via diaphragmatic lymphatics.

Transmission of proteins from intestin[illegible] lining cells to the circulation of young ra[illegible] is a highly selective process. Apparent[illegible] these cells can distinguish between vario[illegible] classes of serum proteins (Bangham an[illegible] Terry, '57) as well as homologous an[illegible] heterologous proteins. When immune sera are prepared from a single antigen in dif-ferent species and fed separately to suckling rats, the highest titer of circulating antibody is attained with homologous serum, while a lower titer is noted with heterologous sera from more distally re-lated species (Halliday, '57). This and other related evidence suggested that the structure or configuration of a protein is important in the process of selective trans-mission. It has been demonstrated that fragments of pepsin refined (Brambell, Hemmings and Oakley, '59) or papain di-gested antibodies (Brambell, Hemmings, Oakley and Porter, '60) are transmitted to the circulation at quite different rates. Fragments I and II, produced by Papain digestion (Porter, '58, '59) contain the antibody reactive sites but are not trans-

ed to the circulation whereas Fragt III, the antigenic part of the antibody ansmitted readily (Brambell et al., '60). nbell, Halliday and Morris ('58) have ulated that the mechanism of selection be related to receptor sites which act a receptor-substrate system recognizing pecific part of the protein. Pierce and th ('67a) have noted that interpreta of some of the studies by Brambell al. is complicated by the fact that their um assays detected only complete se n agglutinins and did not account for omplete antibodies. This is particularly nificant in light of recent findings by rris ('65) demonstrating incomplete ag tinins in the sera of young rats after ding heterologous immune sera. Similar a were obtained by Pierce and Smith 7a) using newborn pigs. In view of se facts, some reassessment of the chanism of selective transmission of oteins seems necessary; previous experiments should be repeated using more reed immunological techniques.

Molecular weights and size of proteins ve been discounted in the past as a deminant of the transmissibility of proins (Brambell, '58, '66; Hemmings and mes, '62). However, its significance may more important than previously realed. For example, a very small protein ch as bovine insulin, 36,000 M.W. in xameric form, is transmitted to the cirlation of suckling mice (Kelly, '60) and ts (Mosinger, Placer and Koldovsky, 9) in sufficient quantities to induce hypolycemia. But very large proteins, IgM mmunoglobulins, 10^6 M.W. and ferritin, 50,000 M.W. are not transmitted (Morris, 5; Graney, '64).

Besides some of the sophisticated immunological and in vitro techniques now vailable, recent advances have been made n tagging antibodies with electron opaque markers. This method should allow one to ollow the pathway of discharged proteins t the ultrastructural level. The evidence t present, though purely circumstantial, ndicates that proteins are discharged at he lateral cell membrane, pass through he intercellular space and enter the lamna propria. A similar pathway has been escribed for the yolk sac (Padykula, Deren and Wilson, '66). Amorphous precipitated material, identical to the contents of the supranuclear vacuole and blood capillaries of the lamina propria, is often observed in the intercellular spaces of the ileum during suckling, but it is not observed in the jejunum. This material is not observed in the intercellular spaces of the jejunal or ileal epithelium of the adult rat. There is, however, a similar precipitated material in the intercellular spaces of intestinal epithelium in humans who have certain protein losing enteropathies, e.g., intestinal lymphangiectasia (Dobbins, '66).

Digestion and fate of ferritin and colostral protein

There are at least two enzymatic environments which ingested proteins must survive before reaching the circulation, the first is in the lumen of the gastrointestinal tract and the second is in the ileal lining cell itslef. Proteolytic activity in the small intestine of newborn rats in high, and does not differ significantly from the activity in adult intestine (Noack, Koldovsky, Friedrich, Heringova, Jirsova and Schenk, '66). Survival of proteins in the intestinal lumen is apparently related to a low activity of digestive enzymes in the stomach. Mosinger et al. ('59) have shown that the proteolytic activity of the gastric contents is low in newborn rats, but increases significantly after day 21. These changes correspond with the morphological development of the parietal and chief cells in the gastric glands (Manville and Lloyd, '32; Kameraad, '42; Hill, '56).

Acid phosphatase and other hydrolytic enzymes have been localized histochemically in the supranuclear vacuole of ileal lining cells (Cornell and Padykula, '65; Straus, '65; Shervey, '66) which would indicate an active lysosomal activity in these cells. It should be mentioned from a comparative standpoint, an apical tubular apparatus with an esterase positive vacuolar system and a second vacuolar system with both esterase and acid phosphatase activities is found in other cell types besides the ileal lining cell, e.g., cells of the proximal convoluted tubule of the kidney (Straus, '64) and yolk sac endoderm (Padykula, '58; Sorokin and Padykula, '64; Lambson, '66). It is not known whether all proteins

must pass through the supranuclear vacuole prior to discharge, or if transmitted proteins bypass the vacuole. It is known that only about 10% of ingested proteins are actually transmitted across the epithelium (Morris, '65; Bamford, '66; Pierce and Smith, '67a,b); the remaining presumably are degradated by the lining cells.

It has been suggested (Brambell et al., '58; Leissring and Anderson, '61; Anderson, '65; Morris, '64; Brambell, '66) that the protein distributed about the periphery of supranuclear vacuoles, may be adsorbed or bound to the limiting membrane of the vacuole. It was further speculated that the binding of the protein may in some way protect it from the proteolytic activity in the vacuole. In the present study there was no evidence to indicate specific binding of ferritin to either the surface membrane or the limiting membranes of the tubular system or supranuclear vacuole. Using the tetrad structure of the micellar core of ferritin as a criterion of its integrity, there was no evidence of ferritin degradation 18 hours after feeding. This is not a good parameter, however, since all ferritin molecules are not necessarily oriented favorably to visualize the micelles. In addition, Matioli and Baker ('63) have described a ferritinic form of hemosiderin as an intermediate in the degradation of ferritin to nonferritinic hemosiderin. The half-life of intestinal lining cells in the suckling animals is about 20% longer than in the adult (Koldovsky, Sunshine and Kretchmer, '66) so that considerable intracellular digestion of protein may occur before the cells are sloughed into the intestinal lumen.

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Note added in proof: Since this manuscript was submitted for publication, two significant review articles related to this topic have appeared in *The Handbook of Physiology*, Section 6: Alimentary Canal, Volume III. Intestinal Absorption, American Physiological Society, Washington, D. C., 1968. See J. J. Deren, page 1099 and I. G. Morris, page 1491.

PLATE 1

EXPLANATION OF FIGURES

The following six figures are photomicrographs of the small intestine from three different animals: newborn rat prior to suckling, nine-day-old suckling rat and a ten-day-old rat fasted for 18 hours prior to sacrifice. Each pair of photographs represent Epon sections of jejunal and ileal tissue taken from the same animal and stained with toluidine blue. Magnification in figures 1–6 is the same, × 1744.

1 Jejunum of newborn rat prior to suckling. There are no obvious inclusions visible in the cytoplasm.

2 Ileum of newborn rat prior to suckling. Morphological pattern of the ileum not significantly different from jejunum (fig. 1).

3 Jejunum of nine-day-old suckling rat. Lipid droplets are observed in Golgi apparatus, intercellular spaces and lamina propria.

4 Ileum of nine-day-old suckling rat. Numerous clear vacuoles are noted in the apical cytoplasm. Supranuclear area occupied by large vacuole containing amorphous dense material.

5 Jejunum of ten-day-old suckling rat fasted for 18 hours prior to sacrifice. At this stage, cytoplasm is completely clear of lipid material. Typical fasting state.

6 Ileum of ten-day-old suckling rat fasted for 18 hours prior to sacrifice. Apical vacuoles and a supranuclear vacuole are still observed in the cytoplasm of the lining cells.

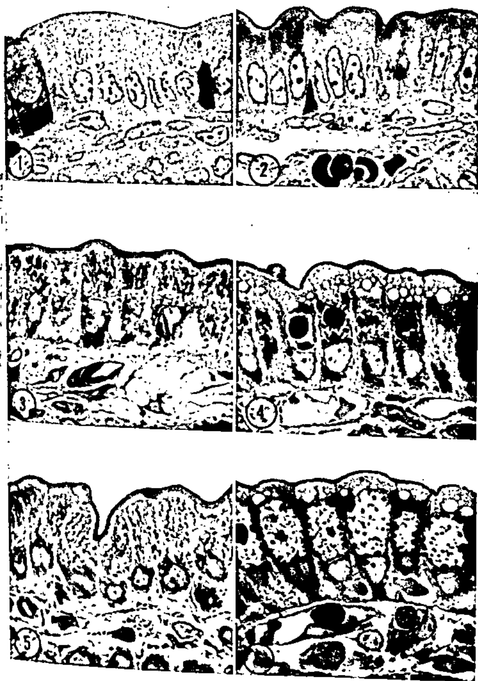

1
2
3
4
5

PLATE 2

EXPLANATION OF FIGURES

7 Jejunum of 15-day-old suckling rat fasted for 18 hours prior to sacrifice. Electron micrograph showing mitochondria, membranes of the smooth- and rough-surfaced endoplasmic reticulum. Note absence of any extensive plasmalemmal invaginations. × 29,000.

8 Jejunum of 15-day-old suckling rat, two hours after receiving 1 ml of corn oil. Osmiophilic droplets, presumably lipid, are observed in both smooth- and rough-surfaced endoplasmic reticulum. × 25,000.

PLATE 3

EXPLANATION OF FIGURE

9 Ileum of 15-day-old suckling rat. Compare amorphous precipitated material in supranuclear vacuole (SNV) with similar material in the intercellular space (ICS), lamina propria (LP) and the blood capillary marked by the red blood cell (RBC). × 13,600.

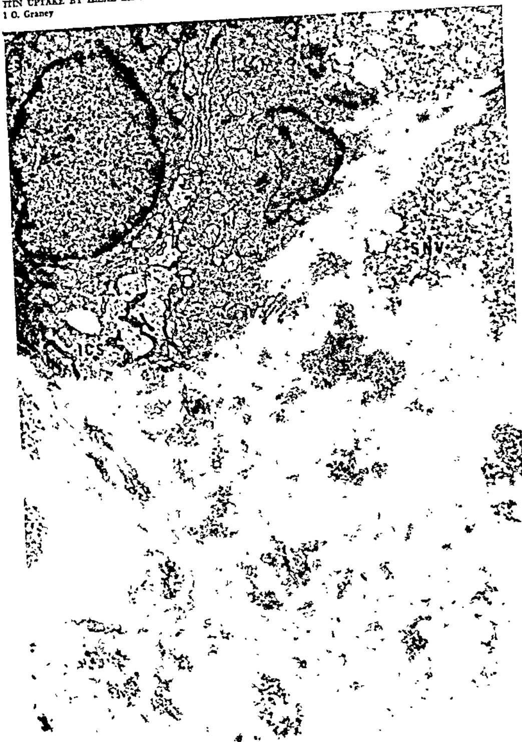

SNV
ICS

PLATE 4

EXPLANATION OF FIGURES

10 Ileum of 15-day-old suckling rat fasted 18 hours prior to sacrifice. Several cisternae (C) with tubular (arrows) connections are seen in the apical cytoplasm. The cisternae and some of the vacuoles (V), particularly those situated deeper in the apical cytoplasm, contain an amorphous precipitated material. × 14,700.

11 Ileum of ten-day-old suckling rat two hours after receiving 0.15 ml of ferritin. Low magnification electron micrograph illustrating the distribution of ferritin in the apical tubular system, apical vacuoles (arrows) and supranuclear vacuole (SNV). × 12,320.

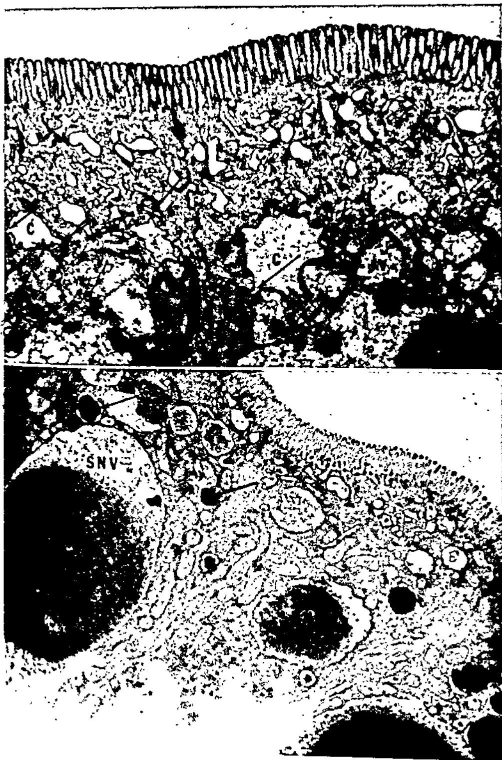

C
C
C
C
SNV

PLATE 5

EXPLANATION OF FIGURE

12 Ileum of ten-day-old rat two hours after ferritin gavage. Ferritin particles (arrows) are seen in the lumen of plasmalemmal invaginations and in tubules situated deeper in the apical cytoplasm. × 102,000. *Inset* illustrates detail of the tearad (arrow) of ferric hydroxide micelles in the center of the ferritin molecule. × 216,000.



PLATE 6

EXPLANATION OF FIGURE

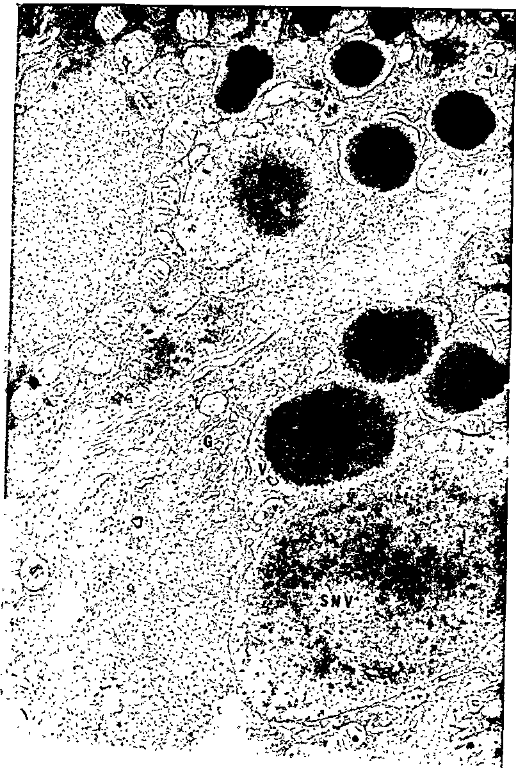
13 Ileum of 15-day-old rat 1.5 hours after ferritin gavage. Three tubules (T) are shown connecting with a large vacuole or cistern (C), containing ferritin. The intestinal lumen is toward the top of the figure. × 135,000.


T
C
13

PLATE 7

EXPLANATION OF FIGURE

14 Ileum of ten-day-old rat two hours after ferritin gavage. Medium sized vacuoles containing concentrated ferritin appear to be coalescing in the supranuclear region of the lining cell shown in the upper half of the figure. The cell in the lower half of the figure already contains a rather large supranuclear vacuole (SNV). Another vacuole (V) is seen fusing the supranuclear vacuole. The Golgi apparatus (G) does not contain ferritin. × 21,420.


G
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›rphological Features of Brown Adipose Cell ıturation *in vivo* and *in vitro* [1,2]

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ABSTRACT The maturation of brown adipose cells, derived at various time intervals from interscapular brown fat pads of neonatal rats, has been investigated electron microscopically. The major changes in *in vivo* brown adipose cells are related to lipid accumulation. In older animals, fat droplets are separated by a membrane complex, and membranous fragments are seen within large lipid inclusions. The ultrastructural characteristics of monolayer cultures of brown fat cells, isolated from neonatal brown fat pads by tryptic dissociation, and maintained for one to six days on a reconstituted collagen matrix, have also been investigated. Adipose cells *in vitro* are morphologically similar to control brown fat cells, and lipid accumulation closely parallels that seen *in vivo*. *In vitro* lipid droplet coalescence, impossible to visualize *in vivo*, has been documented by time-lapse cinematography, and membrane complexes similar to those observed *in vivo* are seen. Major changes in cultivated fat cells, compared to *in vivo* cells, involve extensive glycogen deposition and breakdown of cytoplasmic ribosomal rosettes. Older cultures contain adipose cells exhibiting characteristics of white fat cells. Effects of tissue culture environment upon the brown fat cells, and the possible conversion of the brown fat cell into a white fat cell are discussed.

Since the original description of brown ›pose tissue (Gesner, 1551), the relanship of the brown fat cell to the white · cell has been a controversial subject. me investigators are of the opinion that ›wn adipose tissue is an immature form ·white fat (Auerbach, '02; Sheldon, '24; non, '62), while others maintain that own and white fat are separate tissue enles (Rasmussen, '23; Napolitano, '63). trastructural studies have demonstrated at the morphological characteristics of mature forms of brown and white fat lls are distinctly different (Napolitano, 5). If brown fat cells do mature into hite fat cells, then brown adipose cells ould be required to undergo significant orphological and biochemical changes.

One morphological characteristic of the rown (multilocular) fat cell is the presice of numerous lipid droplets within its ftoplasm, in contrast to the single lipid roplet contained in the mature white unilocular) fat cell. If conversion of a ıultilocular fat cell into a unilocular fat ell occurs, such a process would most kely involve the coalescence of preexist-ıg lipid droplets within the multilocular at cell. There are reports in the literature that brown fat lipid droplets do coalesce, both *in vivo* (Sheldon, '24) and in organ culture systems (Sidman, '56a), but critical documentation of this phenomenon has been lacking.

It is impossible to observe the dynamic process of coalescence *in vivo* or in organ culture. However, monolayer tissue cultures provide an approach for the controlled observation of living cells, and adipose cells exhibiting the light microscopic characteristics of brown fat cells can be maintained in monolayer culture systems (Masters, '65). The cultivated adipose cells, if exposed to insulin, accumulate lipid with time *in vitro*. Time-lapse cinematographic studies of such a lipogenic system might be expected to provide documentation of lipid droplet coalescence, and investigations of this nature are reported.

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these tissues were also fixed in a cold ...tion of 1% osmium tetroxide in phos-...te buffer pH 7.2 (Palade, '52), and ...drated in alcohols. The embedding ...dium was Vestopal W (Kellenberger, ...wab and Ryter, '56) or Maraglas (Spur-..., Kattine and Freeman, '63).

In addition to examining the *in vivo* de-...pmental sequence, electron micro-...pic observations were conducted on ...tivated adipose cells maintained on re-...stituted collagen. Cultures were fixed ...*situ* in 1% osmium tetroxide in phos-...te buffer (pH 7.2) for one-two hours. ...ing alcohol dehydration, a razor blade ... used to strip the gel matrix from the ...er slip, and the matrix together with ... attached cells were processed as an or-...ary piece of tissue. These were embed-...d flat in Vestopal W. After polymeriza-...n, the light microscope was used to se-...t cells for electron microscopy. Selected ...as were cut out with a jeweler's saw and mounted in a blank Vestopal capsule ...th Tray Resin.[4]

All tissues were sectioned on a Huxley ...crotome and double stained with lead ...rate (Reynolds, '63) and uranyl acetate ...Watson, '58). All observations were made ...ith a Philips EM 200 electron microscope.

Time-lapse cinematography. Time-lapse ...nematography was used to study the ...te of lipid droplets within the cultivated ...ipose cells. Trypsinized cell suspensions ...ere prepared as previously described, and ...e cells were introduced into a Sykes-...oore perfusion chamber (Sykes and ...oore, '60). After 48 hours incubation, the ...ulture medium was changed and the per-...sion chamber was inverted and secured ... a glass slide with paraffin. In one series ... experiments the growth medium was ...placed with Krebs-Ringer phosphate buf-...r (pH 7.2) (Cohen, '49) after the initial ...8 hour incubation. The slide and at-...ched chamber was placed on the stage ... an AO Spencer phase-contrast micro-...cope which was housed in a constant tem-...erature incubator (37°C). The progress ... these cultures was monitored by time-...apse cinematography with exposures at ...he rate of one frame per minute.

RESULTS

Control electron microscopy. The following observations are concerned with the ultrastructural features and morphological alterations of rat brown adipose tissues during the first nine days after birth.

The brown fat cell of the neonatal rat contains a spherical or elliptical nucleus which is centrally or slightly eccentrically located (fig. 1, N). The nuclear chromatin is clumped, and localized at the periphery of the nucleus. The nucleoli are conspicuous within the nucleoplasm, and exhibit a distinct *pars fibrosa* and *pars granulosa* (fig. 2, NU).

Newborn brown fat cells contain relatively little lipid, and nuclear displacement occurs with an increase in the size of cytoplasmic lipid inclusions. In figure 7 (4 days post partum), the nucleus at N1 is still centrally located, but the nucleus at N2 is being impinged upon by lipid droplets, resulting in displacement toward the periphery of the cell. The majority of brown fat cell nuclei in older animals have been similarly displaced (fig. 9, N).

There are large numbers of mitochondria in brown fat cells of the newborn animal. These mitochondria are pleomorphic, and cristae orientation is quite variable. Three types of cristae orientation, characteristic of brown fat mitochondria, can be seen in figure 4, where M1 depicts cristae which are perpendicular to the long axis of the mitochondrion, and M2 contains cristae which are almost parallel to the long axis. The mitochondrion labeled M3 contains cristae which are both parallel and perpendicular to the long axis. These specializations in mitochondrial morphology provide a distinct criterion for establishing that cultivated adipose cells are representative of brown fat cells.

Ribosomes are found in great abundance in brown fat cells. The majority of these ribosomes exist as aggregates which are not membrane associated (figs. 2, 3, R), but membrane-bound ribosomes are occasionally found (fig. 2, MBR). Most brown fat cells contain little agranular reticulum, but exceptions to this have been observed. The fat cell in figure 3 contains large amounts of smooth reticulum (SR), which

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Although the ultrastructure of newborn, rat brown fat tissue has been described (Napolitano and Fawcett, '58), electron microscopic studies dealing with temporally-related morphological changes have not been reported. It should also be important to demonstrate that the *in vitro* adipose cell morphology and maturational sequences are similar to that seen *in vivo*. Therefore, ultrastructural characterization of brown adipose cells from newborn rats at various stages of development, and similar studies on the cultivated adipose cells, will be presented. Electron microscopic studies of maturing brown adipose cells might also provide information relating to lipid-droplet coalescence and cell transformation.

MATERIALS AND METHODS

Tissue culture procedures. Brown fat cell monolayer cultures were established as described by Masters ('65). Interscapular brown fat pads from newborn Wistar rats were aseptically removed, diced, and dissociated by agitation in a calcium-magnesium free isotonic solution of 0.25% trypsin (1:300) at pH 7.2 (Moscona, '52). Suspended cells were collected by centrifugation at 1000 rpm for five minutes, and were resuspended in culture medium composed of Eagle's medium (Eagle, '55) containing 2X amino acids, Hanks balanced salt solution (Hanks and Wallace, '49), and 10% bovine fetal serum. Insulin is a requirement for the accumulation of lipid in cultivated brown fat cells (Masters, '65), and was added at a concentration of 10 gamma/ml (0.2 units/ml) of medium. Penicillin was added also at a concentration of 100 units/ml.

Cell viability was determined by exposing the cell sample to trypan blue, which stains non-viable cells (Pappenheimer, '17), and cell counts were made in a hemocytometer. Leighton tubes containing flying-cover slips were inoculated with 5×10^5 cells/ml, and monolayer cultures as described by Earle, Schilling and Shelton ('50) were established. The cultures were incubated at 37°C.

Cultivated cells which were to be examined with the electron microscope were maintained on a reconstituted collagen matrix. Attachment of the fat cells to the collagen matrix provides for retention of cell to cell and cell to matrix relationships and allows circumvention of some of the technical problems associated with the handling of fat-laden cells during electron microscopic preparative techniques.

The reconstituted collagen was prepared by a modification of the technique described by Ehrmann and Gey ('56). Tendons from two rat tails were placed in a 0.1% solution of acetic acid (pH 3.5) and allowed to stand overnight in the cold. This procedure solubilizes the collagen in the tendon material. The resulting viscous solution was then filtered through large-pore gauze material to remove remnants of the tendon. Solid sodium acetate was added to the clarified collagen solution in sufficient quantity to raise the pH to 5 (ionic strength 0.125). Under these conditions the collagen precipitates out of solution in fibrillar form. The fibrils were collected by centrifugation, and all but a small amount of supernatant poured off. A drop or two of acetic acid was added to the fibril pellet, which lowered the pH and solubilized the collagen. The resulting concentrated collagen solution was then layered on cover slips which were placed in a loosely covered container and exposed to ammonium hydroxide vapors. The vapors raised the pH of the collagen solution and a transparent collagen gel was formed. Gel sterilization was effected by placing the cover slips in 70% alcohol which also partially dehydrates the gel resulting in a firmer matrix. Equilibration of the gel was carried out by placing the collagen-coated cover slips in several changes of Eagle's medium. The end product forms a non-toxic, biological matrix which is suitable for the support of cells *in vitro* and strong enough to withstand handling and preparative procedures for electron microscopy (Dyer and Ruby, '63).

Electron microscopy. In order to establish brown fat cell maturational sequences, interscapular brown fat pads were removed from rats at days one, two, four, six and nine post partum. Tissues were fixed overnight in cold 4% gluteraldehyde (Sabatini, Bensch and Barrnett, '63) buffered in McEwen's buffer (McEwen, '56), post-osmicated for one-two hours and dehydrated in a graded series of alcohols. Some

-he fat-containing cell in figure 17 is -ctly apposed to a fibroblastic cell :B), which is in contact with the sur- of the reconstituted collagen matrix :). The differences in mitochondrial : and complexity between the two cells : striking. The fibroblast nucleus (N) :attened, and the cytoplasmic morphol- : is distinct from the fat-containing cell. :all fat droplets are seen occasionally in : fibroblastic cells, but fat droplets com- -able in size to those seen in cultivated :pose cells have not been observed. The :oblasts do not contain glycogen, nor :they exhibit pinocytotic activity.

A third cell type, not seen until four :s *in vitro*, is depicted in figure 18. This , contains a large, centrally located fat lusion, and the nucleus (N), displaced ipherally, is flattened against the cell face. The cytoplasm contains many both vesicles and structures of a par- :late nature. The mitochondria are :all, and their internal structure is un- :nplicated. These cells do not contain :umulations of glycogen, in contrast to :wn fat cells in culture. This third cell :e is similar to a white adipose cell, but should be pointed out that it may rep- :ent a differentiated white fat cell origi- lly present in the brown fat tissue at : time of trypsinization.

Adipose cells after one day *in vitro* are :resentative of brown fat cells one day :t partum. Cultivated fat-cell nuclei g. 12, N), nucleoli (fig. 13, NU), and tochondria (fig. 13, M) are structurally nilar to those observed in control brown t cells. However, ribosomal rosettes, undant in *in vivo* cells, are not nearly evident in cultivated adipose cells. The ltivated fat cells do contain numerous :e ribosomes. Glycogen, rarely seen in ntrol brown fat cells, is a common fea- re of adipose cells *in vitro* (fig. 12, G). rge quantities of glycogen accumulate ith time in culture (fig. 15, G).

There are major changes in the culti- :ted adipose cells associated with lipid :cumulation. Lipid inclusions in 24 hour ultivated cells (figs. 12 and 13, LD) are milar in size and number to those ob- rved in control cells one day post rtum.

There is an increase in lipid droplet size after two days *in vitro*. The fat inclusion labeled LD in figure 14 is 14 μ in diameter, which is three times the size of the largest droplets in one day cultivated cells, and is similar in size to lipid droplets observed in *in vivo* brown fat cells four days post partum. The nuclei of two day culti- vated fat cells are peripherally located (fig. 14, N).

The multilocular fat cells after four days *in vitro* show a slight increase in the num- ber of large lipid inclusions (fig. 16). In figure 16, there is a peripheral rim of lesser density within the larger fat droplets LD1, LD2 and LD3, which is 0.3–0.4 μ in width. LD1 and LD2 contain also, profiles of lesser density which correspond in size to the smaller cytoplasmic lipid inclusions.

Cultivated fat cells six days *in vitro* con- tain numerous large lipid droplets, many of which are in contact with one another. In figure 19, LD1 is impinging upon an adjacent fat droplet at A, and is being im- pinged upon by a second droplet at B. High magnification of the interface at A reveals that the droplets are separated by a membrane complex (fig. 20), which is comparable to the membrane complexes observed between and within lipid drop- lets of control brown fat cells (figs. 10, 11, arrows).

There are many fat-containing cells after six days *in vitro* which are morpho- logically quite different from multilocular fat cells in one, two and four day cultures (fig. 21). Such cells contain large lipid inclusions (LD) which measure up to 20 μ in diameter, and occasionally, small lipid droplets are closely associated with these inclusions (arrow). The nuclei (N1 and N2) of these fat-laden cells are oval or triangular in shape, and are located near the cell periphery. Glycogen bodies (G) are usually present, and parallel arrays of endoplasmic reticulum (ER) are more ap- parent than in earlier cultivated fat cells. These cell types are similar in some re- spects to both the recognizable brown fat cell *in vitro*, and the unilocular fat cell seen in the four day culture system.

Time-lapse cinematography. The pre- ceding electron microscopic evidence strongly suggests that lipid droplet coales-

is randomly dispersed throughout the cytoplasm.

Glycogen has not been observed in brown fat cells of very young animals, but glycogen is frequently seen in the cytoplasm of capillary endothelial cells two days after birth (fig. 6, G). These particles are 250–350 Å in diameter, and exhibit the morphological characteristics and staining reactions of cellular beta glycogen as described by Revel ('64). Small quantities of similar glycogen particles were initially seen in brown fat cells six days post partum.

There is an extensive vascular system permeating brown fat tissue, and brown fat cells are routinely found adjacent to one or more capillaries (fig. 1, C). Vesicles associated with pinocytotic activity are frequently seen at the surface of the brown fat cell and the capillary endothelial cell (fig. 2, P). Pinocytosis is not restricted to the brown fat cell plasma membrane opposite capillary endothelial cells. Pinocytotic vesicles may be seen in many instances lining the surface membranes of two opposed brown fat cells (fig. 3). In figure 3, the formation of pinocytotic vesicles can be seen at P, and at P1 secondary vesicles are seen forming from the initial invagination of the plasma membrane. Similar vesicular structures, termed microvesicular rosettes by Williamson ('64), have been described at the cell surface of white adipose cells acutely depleted of lipid.

The most noticeable cytological changes in brown fat cells are related to lipid accumulation. Brown adipose cells one day after birth contain lipid droplets which vary in diameter from 0.2–4 μ. Although the larger droplets are found in many cells (fig. 1, LD), some cells contain only small amounts of lipid in the form of incipient droplets (figs. 2, 4, LD).

Brown fat cells two days post partum exhibit an increased lipid content, mainly due to an increase in the number of small lipid inclusions (fig. 5, LD). Many of these small droplets are on the order of 1 μ in diameter, which corresponds to the size of chylomicrons. Large numbers of chylomicrons are seen in capillary lumina of the two day old rat (fig. 5, CH), in contrast to the unfed, newborn animal. Fat particles the size of chylomicrons have not been seen in the extracellular spaces between capillaries and fat cells, confirming previous observations (Napolitano, '63).

There is an increase in lipid droplet size four days post partum, resulting in increased cell size, and a corresponding decrease in the dimensions of the extracellular spaces. The lipid droplet labeled A in figure 7 is 15 μ in diameter, which is three times the size of the largest droplets seen in the newborn. Smaller lipid droplets surround and in some cases are apposed to these larger fat inclusions (fig. 7). It is possible to visualize membrane complexes surrounding two closely apposed lipid droplets (fig. 8). Each membrane complex measures 70 Å, and presents the morphological characteristics of a unit membrane. It is not known whether these membrane complexes represent true unit membranes or simply a lipo-protein interface.

Brown fat cells six days post partum contain large lipid inclusions similar in size to those observed after four days post partum (fig. 9, LD), but there does appear to be a significant decrease in the number of smaller lipid droplets.

Brown fat cells nine days post partum contain many adjacent lipid droplets (fig. 10, LD1 and LD2) separated only by an intervening membrane complex. The membrane complex in figure 10 (arrow) is displaced into one of the lipid droplets. Figure 11 (arrow) demonstrates the presence of a membrane fragment, morphologically similar to that described in figure 10, within a lipid inclusion. The fragment measures 150–200 Å in width, and is composed of two outer dense lines and a central zone which is bisected by a third dense line. A lamellar array of membranes (LMS) is found also at the periphery of the lipid droplet in figure 11.

Tissue culture electron microscopy. Three cell types have been observed in the culture system. Figure 17 depicts two of these cell types. The cell containing the lipid inclusions (LD), and the large pleomorphic mitochondria (M), exhibits the general morphology of control brown fat cells. Cytoplasmic vesicles (V), and vesicles of pinocytotic origin (P) are observed frequently within these cells.

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…he presence of large accumulations of …in the cultivated cell can also be …sidered with regard to *in vitro* condi- …s. *In vivo* control mechanisms which …ntain the delicate balance of insulin …osure and glucose availability are lost … Insulin is a requirement for lipid … in the cultivated brown fat …, and it is known that this hormone can … the rate of glucose incorporation … cultivated cells (Sidman, '56b; Vann, …enberg and Lewin, '63). On this basis, …would be reasonable to expect that the …stant exposure of the fat cell to insulin …d glucose, combined with the reduced …l: extracellular fluid volume ratio *in* …ro, would result in an unusually high …cose incorporation into the cell.

Pinocytosis, which is a common feature … the cultivated fat cell, reflects changes … membrane properties consequent to in- …lin exposure. Pinocytosis is probably the …ajor factor which determines the incor- …ration rate of glucose into the cultivated …own adipose cell. A similar role for pino- …tosis has been postulated for insulin- … white adipose cells (Barrnett … Ball, '60). Since neither glycogen ac- … or excessive pinocytotic activ- …y is evident in the fibroblastic cells in …e culture system, it is possible that in- …lin exhibits a selectivity toward the cul- …vated brown fat cell, and/or that the cell … extremely sensitive to the hormone.

Turning now to a consideration of the …asmic lipid component of the brown … cell, it is apparent that there are time- …ated alterations in lipid content of *in* …*vo* and experimental cells. The cells of … systems exhibit an increase in lipid …ontent with the passage of time. The lipid …clusions in the cultivated cells become …arger than those droplets in the *in vivo* …lls, and associated with this increase in … size is a corresponding decrease in … number of fat inclusions. *In vivo* con- … mechanisms and lipid utilization may … the size of lipid droplets in the *in* … brown fat cells.

The smallest visible lipid droplets in the … fat cell are approximately 0.2 μ in diameter. These lipid inclusions are apparently synthesized within the cytoplasm of the brown fat cell, and are not incorporated as pre-formed extracellular lipid bodies. Although the capillary lumina *in vivo* contain chylomicrons, no such structures have ever been seen in the capillary cytoplasm or in the extracellular space between the capillary endothelial cell and the brown fat cell. Lipid accumulation also occurs *in vitro* in the absence of visible lipid structures in the culture medium, and is dependent upon the presence of insulin which facilitates the influx of glucose into the cell. Previous reports in the literature also indicate that lipid inclusions are formed within the fat cell and are not incorporated as preexisting structures (Napolitano, '65).

Membrane-bound lipid droplets have been demonstrated in this study, both *in vivo* and *in vitro*, in brown fat cells. The definitive membrane has been seen surrounding only the larger fat inclusions, supporting a similar claim by Simon ('65). Filamentous structures, surrounding lipid droplets in developing white adipose cells (Wood, '67), have not been observed in this investigation.

If the smaller droplets are not membrane-bound, this would indicate that the presence of a membrane is not necessary for the initial synthesis of lipid and suggests that membrane formation occurs after a critical droplet size is reached. It is possible also that the smaller fat inclusions are membrane-bound, and that the plane of section is not favorable for the visualization of such a membrane.

Recent metabolic evidence suggests that lipid in adipose cells is contained within compartments (Wertheimer, '65); a large compartment serving as a relatively inert storage site in which lipid turnover is very slow, and a second smaller compartment consisting of lipid in a very rapid state of turnover. The presence or absence of a membrane surrounding the lipid droplets might be related to such compartmentalization. The presence of a membrane around larger droplets might act as a physical barrier to lipolysis, thereby preserving a portion of the cytoplasmic lipid component. These large membrane-bound inclusions could represent the lipid com-

cence occurs in the cultivated fat cells after a period of time in culture. Time-lapse cinematography was utilized in an attempt to visualize the process of coalescence, and the results of these studies follow.

Figures 22–27 are sequential frames of a time-lapse film which demonstrates coalescence of fat inclusions in 48 hour cultivated cells placed in Krebs-Ringer phosphate buffer. Figure 22 shows two cells (a and b), which contain numerous lipid droplets. Cell a contains at least ten lipid droplets, and six are present in cell b. Cell b is triangular in shape and measures 17 μ along one side. The lipid inclusions in cell b are 5.5 μ in diameter, and the droplets in cell a measure 4 μ.

After approximately 30 minutes in phosphate buffer, coalescence is seen to occur almost simultaneously in both cells. In frame D (fig. 23, arrow), the coalescence of two lipid droplets in cell b can be seen, and further coalescence in cell b is observed in figure 24, frames G and I (arrows).

In figure 25, further coalescence is seen in cell b (frame J, arrow), and cell a in figure 25 is seen also to undergo coalescence of lipid droplets (frames J and L, arrows).

The remaining frames follow the pattern of coalescence and the final frames depict two cells, each of which contain a large, single lipid inclusion (fig. 27). The fat droplet in cell a (fig. 27, frame P) now measures 8.2 μ and the inclusion in cell b (fig. 27, frame P) is 11 μ in diameter. These droplets are double in diameter as compared with the individual droplets seen in figure 22. However, the total volume of lipid in each cell is comparable before and after coalescence.

Coalescence has also been observed with time-lapse cinematography in cells maintained on standard tissue culture medium. However, these cells have not been observed to undergo total coalescence to a unilocular stage. In addition, cells in the process of dying have been recorded with time-lapse cinematography. The lipid droplets contained in these cells were seen to fragment into smaller droplets, rather than fusing to form larger fat droplets.

DISCUSSION

The morphological data accumulat[illegible]
this investigation have provided additi[illegible]
insight into the cytodifferentiation of [illegible]
brown fat cell. The general ultrastructu[illegible]
features of brown adipose tissue in [illegible]
have been reported previously (Lever, [illegible]
Napolitano and Fawcett, '58; and Napo[illegible]
tano, '65). However, studies designed [illegible]
follow *in vivo* morphological alteratio[illegible]
as a function of time, or ultrastructu[illegible]
characterization of cultivated brown [illegible]
cells, and the temporally-related chang[illegible]
which these cells undergo, have not be[illegible]
attempted prior to this investigation. T[illegible]
present study demonstrates that cultiva[illegible]
adipose cells, derived from brown fat t[illegible]
sues, are representative of brown fat cel[illegible]
and that the *in vitro* maturational [illegible]
quences are similar to those observed [illegible]
vivo. The cultivated fat cells are not ab[illegible]
lutely comparable to control cells, becau[illegible]
they have been trypsinized and exposed
an artificial physical and chemical en[illegible]
ronment. However, these factors app[illegible]
not to have significantly disturbed in t[illegible]
fat cell maturation, as evidenced by the [illegible]
cumulation of lipid with time.

An ultrastructural comparison of the
vivo and *in vitro* brown adipose cells [illegible]
indicated two major differences. First, [illegible]
control cells contain large numbers of
bosomal rosettes, whereas cultivated
cells contain fewer ribosomal aggrega[illegible]
the ribosomes existing as individual pa[illegible]
cles. Secondly, the majority of *in vivo* c[illegible]
do not contain appreciable amounts
glycogen. In contrast, the cultivated
cells contain copious quantities of [illegible]
cogen.

The above differences can be conside[illegible]
a result of culture conditions. The app[illegible]
ent diminution of ribosomal rosettes
cultivated fat cells might reflect a decre[illegible]
in the production of enzymes which
utilized in *in vivo* biochemical reacti[illegible]
that are not employed *in vitro*. The tis[illegible]
culture environment would place ma[illegible]
emphasis on lipogenesis, with little or
need for lipolysis. This physiological s[illegible]
could result in a decreased production
lipolytic enzymes, resulting in disrupt[illegible]
of ribosomal aggregates. It could be arg[illegible]
that the ribosomal changes are a result

degeneration, but this is the least ten- interpretation if one considers that accumulation is an index of cell via- y.

he presence of large accumulations of ogen in the cultivated cell can also be sidered with regard to *in vitro* condi- s. *In vivo* control mechanisms which ntain the delicate balance of insulin osure and glucose availability are lost *vitro.* Insulin is a requirement for lipid umulation in the cultivated brown fat l, and it is known that this hormone can rease the rate of glucose incorporation o cultivated cells (Sidman, '56b; Vann, enberg and Lewin, '63). On this basis, would be reasonable to expect that the stant exposure of the fat cell to insulin l glucose, combined with the reduced l: extracellular fluid volume ratio *in vo*, would result in an unusually high cose incorporation into the cell.

Pinocytosis, which is a common feature the cultivated fat cell, reflects changes membrane properties consequent to in- lin exposure. Pinocytosis is probably the ajor factor which determines the incor- ration rate of glucose into the cultivated own adipose cell. A similar role for pino- tosis has been postulated for insulin- imulated white adipose cells (Barrnett d Ball, '60). Since neither glycogen ac- mulation or excessive pinocytotic activ- y is evident in the fibroblastic cells in e culture system, it is possible that in- lin exhibits a selectivity toward the cul- vated brown fat cell, and/or that the cell extremely sensitive to the hormone.

Turning now to a consideration of the ytoplasmic lipid component of the brown at cell, it is apparent that there are time- elated alterations in lipid content of *in vivo* and experimental cells. The cells of oth systems exhibit an increase in lipid ontent with the passage of time. The lipid nclusions in the cultivated cells become arger than those droplets in the *in vivo* cells, and associated with this increase in droplet size is a corresponding decrease in he number of fat inclusions. *In vivo* con- rol mechanisms and lipid utilization may restrict the size of lipid droplets in the *in vivo* brown fat cells.

The smallest visible lipid droplets in the brown fat cell are approximately 0.2 μ in diameter. These lipid inclusions are apparently synthesized within the cytoplasm of the brown fat cell, and are not incorporated as pre-formed extracellular lipid bodies. Although the capillary lumina *in vivo* contain chylomicrons, no such structures have ever been seen in the capillary cytoplasm or in the extracellular space between the capillary endothelial cell and the brown fat cell. Lipid accumulation also occurs *in vitro* in the absence of visible lipid structures in the culture medium, and is dependent upon the presence of insulin which facilitates the influx of glucose into the cell. Previous reports in the literature also indicate that lipid inclusions are formed within the fat cell and are not incorporated as preexisting structures (Napolitano, '65).

Membrane-bound lipid droplets have been demonstrated in this study, both *in vivo* and *in vitro*, in brown fat cells. The definitive membrane has been seen surrounding only the larger fat inclusions, supporting a similar claim by Simon ('65). Filamentous structures, surrounding lipid droplets in developing white adipose cells (Wood, '67), have not been observed in this investigation.

If the smaller droplets are not membrane-bound, this would indicate that the presence of a membrane is not necessary for the initial synthesis of lipid and suggests that membrane formation occurs after a critical droplet size is reached. It is possible also that the smaller fat inclusions are membrane-bound, and that the plane of section is not favorable for the visualization of such a membrane.

Recent metabolic evidence suggests that lipid in adipose cells is contained within compartments (Wertheimer, '65); a large compartment serving as a relatively inert storage site in which lipid turnover is very slow, and a second smaller compartment consisting of lipid in a very rapid state of turnover. The presence or absence of a membrane surrounding the lipid droplets might be related to such compartmentalization. The presence of a membrane around larger droplets might act as a physical barrier to lipolysis, thereby preserving a portion of the cytoplasmic lipid component. These large membrane-bound inclusions could represent the lipid com-

partment which turns over very slowly. The absence of a membrane around the smaller droplets would facilitate the addition of lipid to an existing droplet, and the lipid material would be also subject to rapid breakdown by cytoplasmic lipase. These non-membrane bound lipid inclusions could represent the lipid compartment which turns over very rapidly.

Coalescence of lipid droplets has been observed in the cultivated multilocular fat cell. The phenomenon of coalescence is not necessarily associated with degeneration of cultivated fat cells, for when cell death is observed *in vitro* in the time-lapse system, lipid breakdown rather than coalescence is seen to accompany the death of the cell. It is also unlikely that the time-lapse film has captured coalescence in a fibroblast, for fibroblasts in culture are spindle-shaped and contain only small amounts of lipid material, whereas the multilocular fat cells are epitheloid in shape, and lipid droplets the size of which are seen to coalesce are found only in those cultivated fat cells with the ultrastructural characteristics of the *in vivo* brown fat cell. For the above reasons, it is concluded that coalescence is a phenomenon that occurs in viable brown fat cells *in vitro*. Since it is impossible to visualize the process of coalescence *in vivo*, one cannot assume that those events which occur *in vitro* also occur *in vivo*. The increase in lipid droplet size with time *in vivo* need not be a result of coalescence, for lipid droplet size could increase by addition of newly formed lipid to existing fat inclusions.

There are certain morphological characteristics of apposed lipid droplets in cultivated and control fat cells which could be associated with the process of coalescence. The membrane complexes observed between fat droplets *in vitro* may represent the initial stages of droplet coalescence, and similar membrane complexes are seen separating fat droplets in control brown fat cells. Discontinuities of this membrane structure, observed in control cells, have not been seen between lipid droplets *in vitro*, but since coalescence of two droplets can occur in a one minute time span, as shown by the time-lapse results, it would be extremely difficult to capture membrane fragmentation at the ultrastructural level in a system that contains relatively few cells compared to the control system.

Fragmented portions of membranes als appear free within lipid droplets. If tot fragmentation of intervening membran between two lipid droplets were to occ as a result of coalescence, one would e pect to see such fragments within new formed lipid droplets. The localization membranous inclusions in lipid drople supports the postulation that coalescen does occur *in vivo*.

The problem of conversion of brown cells into white fat cells can also be dis cussed on the basis of the results of th study. If transformation of a multilocula fat cell into a unilocular fat cell were t occur, then one necessary, morphological change would be the coalescence of th numerous lipid droplets of the multilocu lar cell into a single lipid inclusion. Th time-lapse results demonstrate such a mo phological change. Conversion would r sult also in total nuclear displacement the cytoplasm, with subsequent flattenin of the nucleus as the lipid inclusion e larged, and there would be changes in t cytoplasmic organelles, especially with r gard to mitochondrial morphology a number. Some cultivated cells are morp logically different from recognizable bro fat cells and exhibit some structural f tures of the white adipose cell, and types which strongly resemble white cells also have been seen *in vitro*. H ever, the present investigation does provide sufficient evidence to the ef that these cells are brown fat cells in process of transformation *in vitro*.

It is possible that some control me anisms which are functional *in vivo* lost *in vitro*, resulting in conversion of cultivated brown fat cell into a cell w exhibits some of the morphological c acteristics of a white fat cell. That su control mechanism does exist *in vi* supported by the fact that brown adi cells are maintained as a specific t entity throughout the life-span of th bernator, and that in a non-hiber such as the rat, the tissue sites occu by brown fat cells in the newborn contain large amounts of white ad cells and very few brown adipose ce

.on this basis that some investigators .l that brown fat can be transformed .ɔ white adipose tissue. The results of .s *in vitro* study neither confirm nor .iy such a postulation, but they are con- .ent with the notion that such conver- .ns are possible.

.n conclusion, the brown fat cell exhibits .king morphological changes during its .rphogenesis. The most apparent, tem- .ally-related, morphological alterations .ich these cells undergo involve the size .l physical character of the lipid pool. .rphological data from both *in vivo* and .vitro systems strongly suggest that lipid .ithesis is initially associated with the .mation of small droplets which subse- .ently increase in size, become sur- .nded by a membrane, and finally fuse .h one another by the process of coales- .ice. The problem of conversion of the .wn fat cell into a white fat cell remains . open question, but that such a conver- .n can occur is suggested by the *in vitro* .idence.

.Since the cultivated cell is no longer ex- .sed to *in vivo* control mechanisms, the .tentialities of the cell become more ap- .rent in the tissue culture system, pro- .led that the artificial environment does .t interfere with the emergence of such .tentialities. That the fat cells *in vitro* .e representative of control brown fat cells .nnot be disputed. Some reservations .ust be maintained, however, concerning .e extent to which *in vivo* and *in vitro* .ta may be compared. This investigation .s provided an experimental system and .id a basic foundation for further correla- .ve studies which might lead to the eluci- .ition of the physiological, biochemical, .id morphological properties and capabil- .les of the brown fat cell.

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Abbreviations

C, Capillary
CH, Chylomicron
ER, Endoplasmic reticulum
FIB, Fibroblast
G, Glycogen
LD, Lipid droplet
LMS, Lamellar-membrane system
M, Mitochondrion
MBR, Membrane-bound ribosomes
N, Nucleus
NP, Nuclear pore
NU, Nucleolus
P, Pinocytotic vesicle
R, Ribosome
RBC, Red blood cell
RC, Reconstituted collagen
SR, Agranular reticulum
V, Cytoplasmic vesicle

PLATE 1

EXPLANATION OF FIGURES

1 Day one post partum. A general survey electron micrograph demonstrating the tissue organization of the newborn, rat interscapular brown fat pad. Each adipose cell is adjacent to at least one capillary (C), some of which contain a red blood cell (RBC). There are large extracellular spaces in this tissue, relative to tissues in older animals. (LD, lipid droplet; N, nucleus). × 3400.

2 Day one post partum. A portion of a brown fat cell and an adjacent capillary. The adipose cell contains small lipid droplets (LD), numerous mitochondria (M), ribosomal rosettes (R), and small amounts of membrane-bound ribosomes (MBR). Pinocytotic vesicles (P) are located along the surface of the fat cell and the capillary endothelial cell. (NU, nucleolus). × 8250.

PLATE 2

EXPLANATION OF FIGURES

3 Day one post partum. Adjacent adipose cells exhibiting numerous ribosomal rosettes (R), and elements of smooth reticulum (SR). Pinocytotic vesicle formation is observed at P, and P1 indicates a microvesicular rosette. × 9600.

4 Day one post partum. Portions of brown fat cells containing large numbers of mitochondria. M1, M2 and M3 depict the patterns of cristae orientation characteristics of brown fat cell mitochondria. (LD, droplet). × 5600.

PLATE 2

EXPLANATION OF FIGURES

3 Day one post partum. Adjacent adipose cells exhibiting numerous ribosomal rosettes (R), and elements of smooth reticulum (SR). Pinocytotic vesicle formation is observed at P, and P1 indicates a microvesicular rosette. × 9600.

4 Day one post partum. Portions of brown fat cells containing large numbers of mitochondria. M1, M2 and M3 depict the patterns of cristae orientation characteristics of brown fat cell mitochondria. (LD, droplet). × 5600.

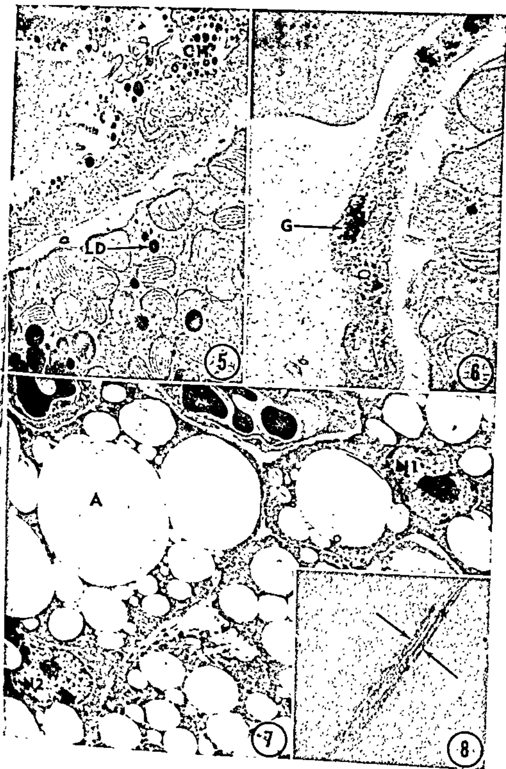

CH
LD
G
A
M1
M2
5
6
7
8

PLATE 3

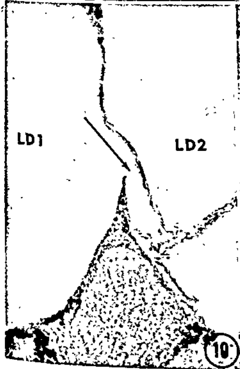
EXPLANATION OF FIGURES

5 Day two post partum. Portions of a brown fat cell and an adjacent capillary. The capillary lumen contains large numbers of chylomicrons (CH). Numerous lipid droplets (LD) are present in the brown adipose cell. × 18,720.

6 Day two post partum. Portions of a capillary and a brown fat cell. Cytoplasmic glycogen (G) is present in the capillary endothelial cell. × 25,000.

7 Day four post partum. A survey electron micrograph demonstrating large numbers of lipid inclusions in the brown fat cells four days after birth. The weak osmiophilia of the lipid is due to preparative procedures. Note the close association of surrounding lipid droplets to the droplet labeled A. There is a general increase in cell size due to lipid accumulation, and a corresponding decrease in extracellular space. (N, nucleus). × 2975.

8 Day four post partum. Two large lipid droplets separated from each other by tri-partite membrane complexes (arrows). × 146,250.


LD1
LD2
10


LMS
LD
11

PLATE 4

EXPLANATION OF FIGURES

9 Day six post partum. A survey electron micrograph demonstrating large lipid inclusions (LD) in the adipose cells. The extracellular spaces between these fat-laden cells are very small. (N, nucleus). × 5000.

10 Day nine post partum. A membrane complex between two lipid droplets (LD1 and LD2). Note the lateral displacement of the membrane complex at the arrow. × 95,800.

11 Day nine post partum. A membranous fragment (arrow) within a lipid droplet (LD). A lamellar-membrane system (LMS) is seen at the periphery of the lipid droplet. × 100,625.

N
G
12

M
NP
NU
13

PLATE 5

EXPLANATION OF FIGURES

12 Day one *in vitro*. A cultivated adipose cell exhibiting the general morphological characteristics of *in vivo* brown fat cells, except for large glycogen accumulations (G). Note the numerous small lipid inclusions (LD), and the spherical, eccentric nucleus (N). × 7660.

13 Day one *in vitro*. A cultivated brown fat cell containing a prominent nucleolus (NU), and exhibiting pores in the nuclear envelope (NP). The mitochondria (M) are characteristic of control brown fat cells. (LD, lipid droplet). × 18,000.

N
G
12
M
NP
13

PLATE 6

EXPLANATION OF FIGURES

14 Day two *in vitro*. Portions of a cultivated adipose cell exhibiting the morphological characteristics of *in vivo* brown fat cells four days post partum. (LD, lipid droplet; N, nucleus). × 5200.

15 Day two *in vitro*. Two cultivated brown fat cells which contain large accumulations of glycogen (G). × 16,380.

16 Day four *in vitro*. An adipose cell containing numerous lipid droplets, some of which exhibit a peripheral rim of lesser density (LD1, LD2 and LD3). Note the apposition of small fat inclusions to LD1 and LD2. LD1 and LD2 contain profiles of lesser density which correspond in size to the smaller lipid droplets. × 6200.

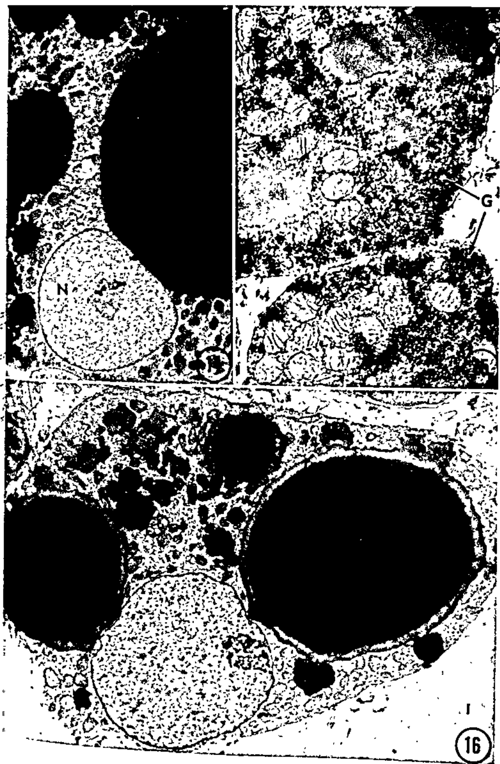

N
G
16

PLATE 7

EXPLANATION OF FIGURES

17 Day four *in vitro*. This micrograph depicts the two cell types found in the culture system. The fat cell contains lipid droplets (LD), cytoplasmic vesicles (V), numerous, large, complex mitochondria (M), and the cell surface is lined with pinocytotic vesicles (P). The fibroblastic cell (FIB) is strikingly different from the cultivated fat cells. Notice in particular the differences in mitochondrial morphology. The fibroblastic cell (FIB) is attached to the surface of the reconstituted collagen matrix (RC). (N, nucleus). × 12,250.

18 Day four *in vitro*. A cell containing a single, large lipid droplet and a peripherally displaced and flattened nucleus (N). This cell exhibits the general morphological characteristics of a white fat cell. × 7950.


M
FIB
P
N
RG
17
N
18

PLATE 8

EXPLANATION OF FIGURES

19 Day six *in vitro.* A portion of a cultivated multilocular adipose cell. Note the apposition of adjacent lipid droplets at A and B to the lipid inclusion labeled LD1. × 11,690.

20 Day six *in vitro.* A high magnification of the interlocking interface at A in figure 19, revealing the membrane complex at this region (arrow). × 100,625.

21 Day six *in vitro.* A number of adipose cells containing large lipid droplets, (LD), glycogen bodies (G), and displaced nuclei (N1, N2). Small lipid inclusions are frequently seen at the periphery of the larger droplets (arrow). (ER, endoplasmic reticulum). × 3600.

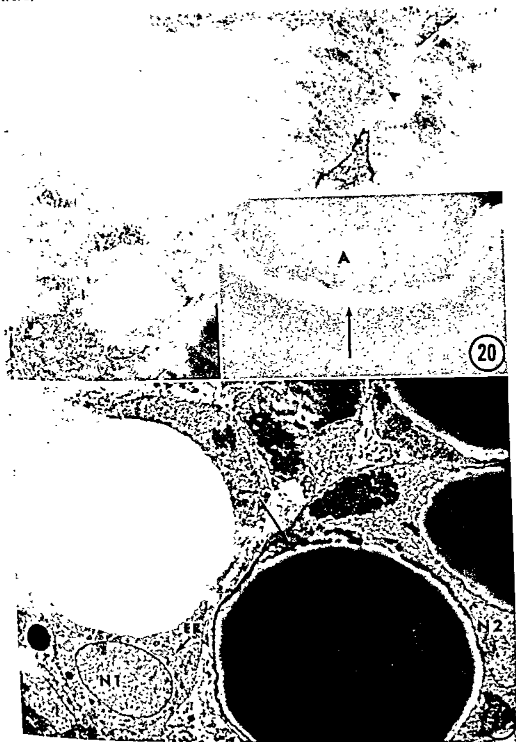

A
20
NT
ER
N2

PLATE 8

EXPLANATION OF FIGURES

19 Day six *in vitro*. A portion of a cultivated multilocular adipose cell. Note the apposition of adjacent lipid droplets at A and B to the lipid inclusion labeled LD1. × 11,690.

20 Day six *in vitro*. A high magnification of the interlocking interface at A in figure 19, revealing the membrane complex at this region (arrow). × 100,625.

21 Day six *in vitro*. A number of adipose cells containing large lipid droplets, (LD), glycogen bodies (G), and displaced nuclei (N1, N2). Small lipid inclusions are frequently seen at the periphery of the larger droplets (arrow). (ER, endoplasmic reticulum). × 3600.

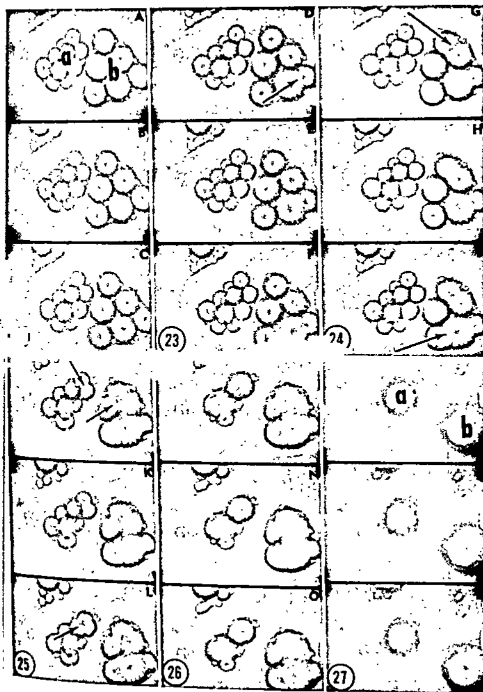

A
a
b
B
C
D
E
F
G
H
23
24
K
L
N
O
a
b
25
26
27

PLATE 9

EXPLANATION OF FIGURES

22–27 Day two *in vitro*. Sequential frames of a time-lapse film. Two fat-containing cells are seen (fig. 22, frame A, a and b). Lipid-droplet coalescence is observed at the arrows in figure 23 (frame D), figure 24 (frame G and I), and figure 25 (frame J and L). Figure 27 depicts two unilocular fat cells (frame P, a and b), formed by the coalescence of lipid droplets. × 1375.

e Ultrastructure of the Crevicular ithelium of Cat Gingiva [1]

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ABSTRACT The ultrastructure of the crevicular epithelium of clinically healthy cat gingiva was similar to that of other non-keratinizing oral epithelia. However, it differed in that the intercellular spaces were wide and extended as a continuous network from the basal lamina to the epithelial surface. Basal cells contained numerous mitochondria, Golgi apparatus, free ribosomes and a fine feltwork of evenly distributed tonofilaments. Hemidesmosomes were present in relation to the basal lamina. Cells in the intermediate layers showed a decrease in the number of organelles and an increase in concentration of tonofilaments. Superficial cells were further flattened and the remaining organelles commonly showed degenerative changes. Epithelial cells showed prominent microvilli extending into the intercellular spaces. Some of these formed desmosomes with similar processes of adjacent cells. Occasional randomly distributed tight junctions were also observed. Inactive basal melanocytes, more superficial Langerhans cells and leucocytes were seen between epithelial cells. The great majority of leucocytes were neutrophils and many of these ruptured as they approached the epithelial surface scattering their specific granules into the intercellular spaces. These findings are discussed in relation to the peculiar permeability of the gingival crevicular region of the oral mucosa to tissue fluid and leucocytes.

Electron microscopy has been used to 'estigate the structure of gingival epi- :lium in both animals (Albright, '60; :rn, '65) and in man (Kurahashi and kuma, '62; Listgarten, '64; Stern, '65; aterhouse, '65; Haim, '66; Schroeder and eilade, '66) but these studies have dealt nost exclusively with the keratinizing ithelium which forms the outer or masatory surface of gingiva. Stern ('62) d Listgarten ('66) studied the fine struc- :e of the attached epithelial cuff of gin- /a and elucidated the nature of the nto-gingival junction. Thilander ('63) idied the action of enzymes on the stra- m spinosum of the crevicular epithelium id Weinstein et al. ('67) examined it llowing the intravenous administration saccharated iron oxide. However, the ie structure of this epithelial lining of e gingival crevice has not been compre- :nsively described.

Using intravenous tracers Brill and rasse ('58) and Brill and Bjorn ('59) :monstrated that a protein-containing 3rill, '59) fluid can pass through the inically healthy crevicular epithelium 1t not through other regions of oral and asal mucosa. Recently Weinstein et al. ('67) described differences in composition between this gingival fluid and serum which suggest that permeability of the crevicular epithelium is a selective one. Sharry and Krasse ('60) and Löe ('61) demonstrated that the gingival crevice is also the site of entry into the mouth of salivary leucocytes.

The present investigation was undertaken to describe fully the fine structure of crevicular epithelium, and to provide a basis for understanding the permeability of this region of the oral mucosa.

MATERIAL AND METHOD

Strips (the marginal 2–3 mm, fig. 1) of clinically healthy gingiva were excised from the labial and buccal surfaces of canine and premolar teeth of ten mongrel cats. They were immediately sliced transversely at 0.5–1 mm intervals in a drop of fixative, and fixation was continued for one hour, either in ice-cold, phosphate-buffered (pH 7.3) 1% osmium tetroxide solution containing 5.4% glucose (Millonig, '62), or in veronal-acetate buffered

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M. J. ANAT., 123: 283–296.

grouped and oriented into prominent dles, as in keratinizing gingival epithe- n (fig. 5).

ine circular vesicles, often arranged in s in close relation to the cell mem- ne, were common in the cytoplasm of basal cells. Indentations and invagina- s of the basal cell plasma membranes suggested pinocytotic activity (figs.). Occasional dense rounded bodies h a finely granular, or sometimes multi- icular, matrix were present within the oplasm of crevicular epithelial cells g. 3) and larger lipid droplets were also nmon (fig. 10).

The desmosomes of crevicular epithe- m consisted of opposing zones of cell mbrane specialization, subjacent cyto- smic electron density and tonofilament ncentration, as seen in the hemidesmo- mes, separated by an intercellular in- val of some 200 A (fig. 9). The inter- llular lamellae which form part of the smosomal complex in keratinizing gingi- l epithelium were often not as apparent similarly treated crevicular epithelium. ight junctions, characterized by a fusion the outer leaflets of adjacent cell mem- anes and the absence of adjacent cyto- lasmic modification, were occasionally en amongst groups of desmosomes. They ppeared to be randomly distributed and ere not characteristically found in the perficial strata (fig. 9).

Passing from the basal to intermediate ell layers there was a progressive flatten- g of the epithelial cells, nuclear outlines nded to become more irregular, and there as a diminution in the number of mito- hondria. Conversely, the number of tono- ilaments tended to increase (figs. 4, 8) ut unlike keratinizing gingival epithelium here was no tendency for them to form rominent bundles (fig. 5). In the super- icial cells cytoplasmic lipid droplets were more common (fig. 10) but cytoplasmic organelles were fewer and tended to be poorly defined and degenerate. Intracellu- lar lysosome-like bodies at this level often had bizarre forms and membranous whorls resembling myelin figures were also ob- served.

At all levels the intercellular spaces were wide (figs. 3, 6, 8, 10, 11, 13) with local- ised dilatations up to several microns in width. A finely granular material (figs. 6, 11), generally similar in appearance to the ground substance of connective tis- sue, filled the intercellular spaces which formed an interconnecting network ex- tending from the basal lamina densa to the external environment. In the interme- diate (fig. 8) and superficial (fig. 13) layers, the intercellular spaces frequently contained cellular debris and membrane- bound dense bodies which exhibited a vari- ety of shapes (fig. 13). Some resembled the intracellular lysosome-like bodies already described. Microvilli of various lengths pro- jected from epithelial cells into the inter- cellular spaces and often contacted similar processes from adjacent cells (fig. 8).

Basal melanocytes were recognised by their characteristically indented nucleus and a cytoplasm rich in mitochondria but devoid of tonofilaments and cytoplasmic granules (fig. 7). These cells were found close to the epithelial basal lamina but were usually separated from it by fine processes of the adjacent Malpighian cells. They did not form desmosomes with the epithelial cells (fig. 7). Melanocyte proc- esses were seen in the intercellular spaces at all levels. Langerhans cells with simi- larly indented nuclei and cytoplasm rich in mitochondria were present in the inter- mediate and superficial layers.

Leucocytes were also present within the intercellular spaces of crevicular epithe- lium. Although lymphocytes and larger cells of the monocyte series were observed, neutrophils were the most common type. The latter were readily recognised by their nuclear polymorphism and rich cytoplas- mic content of densely staining "specific granules" (fig. 11). The cell membranes of neutrophils in the more superficial parts of the epithelium frequently showed dis- continuities not evident in the plasma membranes of adjacent epithelial cells (fig. 11). Rupture of these migrating neu- trophils (figs. 11, 12) as they approached the salivary environment seemed the most likely explanation for the presence of lyso- some-like bodies within the intercellular spaces of the crevicular epithelium.

DISCUSSION

Although the fine structure of the cre- vicular epithelium was found to be gen-

(pH 7.3) 2% osmium tetroxide containing 4.5% sucrose (Caulfield, '57). After dehydration in graded ethanol and infiltration with propylene oxide, the triangular gingival slices were embedded flat in discs of Araldite (Luft, '61) or in an Araldite and Epon mixture (Yamamoto, '66).

The polymerized discs were then viewed with a dissecting microscope to distinguish the strongly osmiophilic keratinizing epithelium, the lightly stained crevicular epithelium and the virtually unstained connective tissue, each of which formed one margin of the slice of gingiva. As an additional check on the localisation of the block face, 1 μ sections were cut and stained with 0.5% alkaline methylene blue for light microscopy. Sections (700–1200 Å in thickness) were stained with uranyl acetate (Watson, '58) and lead citrate (Reynolds, '63) and examined with a Hitachi HU-11A electron microscope.

OBSERVATIONS

Under the light microscope the crevicular epithelium varied from 3–4 cells in thickness over connective tissue papillae to several times this thickness between them (figs. 1, 2). Wide-bore capillaries were numerous just deep to the basement membrane separating the collagenous connective tissue from the basal epithelial cells (fig. 2). The germinative cells stained deeply with methylene blue and were frequently observed in mitosis (fig. 2). In the intermediate layers, cells showed a transition from the cuboidal or low columnar basal cells to the more flattened superficial cells, a change accompanied by a loss of basophilia and nucleolar prominence (fig. 2). Although sometimes smooth, the surface of the crevicular epithelium was often ragged (fig. 2).

With the electron microscope the junction of the epithelium and connective tissue was seen to be irregular due to projections and invaginations of the basal epithelial cells (fig. 4). The junctional zone or "basement membrane" consisted of two lamellae. A relatively clear layer, the lamina lucida, some 250–400 Å thick separated the plasma membranes of the basal cells from an electron dense layer, the lamina densa, which was of similar thickness (figs. 4, 5, 6). Fine fibrils about 100 Å in diameter with faint but irregular transverse striations radiated from the lamina densa between the larger (250–600 Å) light staining and characteristically striated collagen fibres most of which were parallel to the basal lamina. These fine fibrils were more numerous beneath keratinizing gingival epithelium (fig. 5) than deep to the crevicular epithelium (fig. 4). The lamina densa was continuous across the spaces between basal cells but there was little distinction between the lamina lucida and the intercellular material (fig. 6).

Hemidesmosomes were present along the junction of the crevicular epithelium and connective tissue (fig. 4) but were not as numerous as those beneath keratinizing gingival epithelium. At the half desmosome, the epithelial cell membrane showed an increased separation of its component leaflets and an intensification in staining of the inner one. The subjacent cytoplasm exhibited a diffuse electron density which shaded off into the concentration of tonofilaments usually present. On the connective tissue side of the modified plasma membrane an additional dense lamina 60 Å thick, and separated from the cell by 100 Å, was present in the lamina lucida. There was a tendency for the lamina densa in these regions to show an increased density.

The basal epithelial cells had centrally placed ovoid nuclei bounded by a nuclear membrane containing nuclear pores. Mitochondria were numerous in these cells particularly in the basal parts of the cytoplasm and in the vicinity of the Golgi apparatus (fig. 3). One or two electron-dense granules were frequently present within these mitochondria in relation to the inner mitochondrial membrane. The series of parallel lamellae and vesicles which comprised the Golgi apparatus tended to be arranged parallel to the nuclear membrane and quite close to it (fig. 3). Rough surfaced endoplasmic reticulum was rarely present in any significant amounts but free ribosomes, often grouped to form polyribosomes, were abundant and distributed throughout the cytoplasm amongst the even feltwork of short fine tonofilaments. The tonofilaments of crevicular epithelium (fig. 3) were not as numerous, nor were

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erally similar to that of non-keratinizing oral epithelia (Zelickson and Hartmann, '62, '63) it differed in two significant respects, namely, the continuity of its intercellular spaces with the oral cavity and the presence within it of large numbers of leucocytes.

Prominent intercellular spaces, like those of crevicular epithelium, are also seen in amphibian epidermis but in the latter, belts of membrane fusion (zonulae occludentes) bind the superficial cells into a continuous sheet (Farquhar and Palade, '64, '65). Diamond and Tormey ('66) have reviewed the role of long extracellular channels in the transport of fluid across epithelia. However, in the epithelia they discuss (intestine, gall bladder, kidney) the intercellular spaces are closed externally by tight junctions as in amphibian epidermis. Such tight junctions as were observed between the cells of the crevicular epithelium appeared to be insufficient in number to form such a barrier, and the few present were not limited to the superficial layer of cells. The possibility that the presence of such wide intercellular spaces was a fixation artefact was eliminated by comparing the crevicular epithelium with the keratinizing epithelium in the same slice of gingiva. In the latter epithelium, as in non-keratinizing oral epithelium (Zelickson and Hartmann, '62, '63), the superficial intercellular spaces were virtually obliterated.

Although a study using saccharated iron oxide as an electron-dense tracer (Weinstein et al., '67) did not produce conclusive evidence of intercellular passage, it seems probable that the width and continuity of the intercellular spaces is the morphological basis for the outward movement of tissue fluid (Brill and Krasse, '58; Brill and Bjorn, '59) through the crevicular epithelium, but not through other regions of healthy oral mucosa. The recovery of intravenous sodium fluorescein has been used to demonstrate the flow of this gingival fluid in the dog (Brill and Krasse, '58), man (Brill and Bjorn, '59), and rabbit (Brown-Grant and Browne, '66) and we have found that it also enters the gingival crevice in the cat (unpublished results). The differences in ratios of components between this fluid and serum (Weinstein et al., '67) indicate that it is not a simple transudate. Two morphological features of the crevicular epithelium could be related to this selectivity. They are the epithelial basal lamina which appeared to form the only continuous "barrier" between the connective tissue and the oral cavity, and the prominent microvilli which suggest cellular modification of the content of the intercellular spaces.

Similarly, the wide intercellular spaces could be the route for inward passage of materials through this epithelium. Egelberg ('63) has demonstrated that histamine placed in the gingival crevice produces changes in the underlying vasculature while Rizzo and Mitchell ('66) have produced both a local and a systemic antibody response to the repeated application of egg albumin to the crevicular region.

In view of this inward permeability, the presence of leucocytes in the gingival tissues could be interpreted as an inflammatory response to the bacterial flora of the gingival crevice (Gavin and Collins, '61). However, Orban ('57) believes that the presence of such cells does not, in itself constitute a pathological condition. This latter view is supported by the presence of leucocytes in the gingiva of germ-free mice (Rovin, Costich and Gordon, '66) and the other mucous membranes of the genital (Beaver, '60), respiratory and digestive tracts (Kent, '66) which are not commonly inflammed. Until it is determined whether gingival leucocytes represent an actual, or only a potential, defense mechanism, the histological control of the selection of healthy gingiva will remain unreliable. Many of the neutrophils observed in the crevicular epithelium appeared to be disintegrating and discharging their granules into the intercellular spaces. Indeed, the prominence of these spaces in the material examined might be related to this leucocytic degranulation, for both leucocytic enzymes and hyaluronidase increase their width (Thilander, '63). The latter enzyme may be a product of the crevicular bacterial flora (Thilander, '64).

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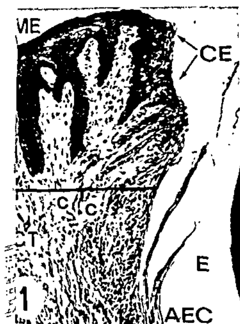
PLATE 1

EXPLANATION OF FIGURES

1 Light micrograph of cat gingiva showing keratinized masticatory epithelium (ME), the crevicular epithelium (CE) related to the gingival crevice, and the attached epithelial cuff (AEC) related to the space (E) occupied by the enamel prior to decalcification. These epithelia cover a collagenous tissue (CT) which is well supplied with capillaries (c). The horizontal line indicates the level of the scalpel cut used to obtain specimens of crevicular epithelium. Paraffin section, Masson's trichrome stain. × 40.

2 Light micrograph of crevicular epithelium showing the basophilic basal epithelial cells one of which is in mitosis (mf), the more flattened intermediate cells (i) which have prominent nucleoli, and the lightly staining, somewhat ragged superficial cells (s). Several capillaries (c) are present just deep to the epithelium. Araldite section, methylene blue stain. × 400.

3 Electron micrograph of parts of four basal cells of crevicular epithelium. Such cells have an ovoid nucleus (N), numerous mitochondria containing small dense granules (M), Golgi vesicles (G), pinocytotic vesicles (P), lysosome-like granules (L), and a fine feltwork of tonofilaments (T). These cells are separated from one another except at the desmosomes (D) by the intercellular spaces (S) into which microvilli (MV) project. × 12,500.


ME
CE
C
C
T
E
1
AEC


S
mf
C
C
2
S

PLATE 2

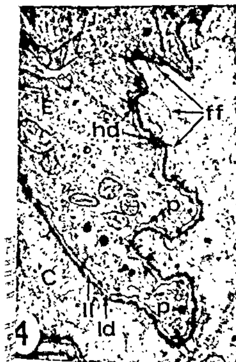
EXPLANATION OF FIGURES

4 Electron micrograph of the convoluted junction between crevicular epithelium (E) and the underlying connective tissue (C). The junction consists of a lamina lucida (ll) and a lamina densa (ld) from which fine fibrils (ff) radiate between the lightly staining collagen fibres beneath. The basal epithelial cell membrane shows invaginations reminiscent of pinocytosis (p) and an increase in electron density at hemidesmosomes (hd). × 25,000.

5 Electron micrograph of the junction between keratinizing gingival epithelium and the underlying collagenous tissue. The aggregation of tonofilaments (TF) into bundles is shown within the epithelial cells which form hemidesmosomes (hd). Fine fibrils (small arrows) radiate from the lamina densa (ld) between underlying collagen fibres (large arrows). × 45,000.

6 Electron micrograph of the junction of crevicular epithelium (E) and the underlying connective tissue (C). The dilated intercellular space (S) is separated from the connective tissue ground substance by the lamina densa (arrows). × 20,000.

7 Electron micrograph of a basal melanocyte in crevicular epithelium. This cell has a prominently indented nucleus (N) and a cytoplasm rich in mitochondria (m) but devoid of tonofilaments. Processes of adjacent epithelial cells (E) separate the melanocyte from the basal lamina and underlying connective tissue (C). × 13,500.


E
hd
ff
p
C
lf
ld
p
4


hd
TF
ld
5


C
S
E
E
6


N
m
E
E
C
7

PLATE 3

EXPLANATION OF FIGURES

8 Electron micrograph of portions of several intermediate cells of crevicular epithelium. These show desmosomes (D) perinuclear mitochondria (m) and microvilli (mv) which project into the wide intercellular space (S) containing a free dense body (arrow). × 10,000.

9 Electron micrograph of the contact relationship of two intermediate level cells of crevicular epithelium (E). Portions of three desmosomes (D) and two regions of tight junction (arrows) interrupt the continuity of the intercellular space (S). × 50,000.

10 Electron micrograph of portions of three flattened superficial cells of crevicular epithelium. They contain a lipid droplet (Li) and a lysosome-like body (L). The intercellular spaces (S) also contain free dense bodies (arrows). × 15,000.





S
LI
I
S
L
S
S
10

PLATE 4

EXPLANATION OF FIGURES

11 Electron micrograph of several intermediate level cells (E) of crevicular epithelium. The dilated intercellular space (S) contains a neutrophil leukocyte identified by its polymorphic nucleus (N) and by its cytoplasmic granules. Arrows indicate breaks in the cell membrane of the neutrophil but adjacent epithelial cell membranes are intact. × 6,000.

12 Electron micrograph of part of a superficial cell of crevicular epithelium (E) and related intercellular space (S). This space contains a process from a neutrophilic leukocyte which has ruptured and is discharging its cytoplasmic granules into the intercellular space. × 25,000.

13 Elecrton micrograph of an intercellular space between superficial cells (E) of the crevicular epithelium. It contains cellular debris (CD) and numerous dense bodies (arrows) which are ovoid (1), indented (2), elongated (3), round (4) or irregular (5) in form (cf. fig. 12). × 22,000.


E
S
E
N
11

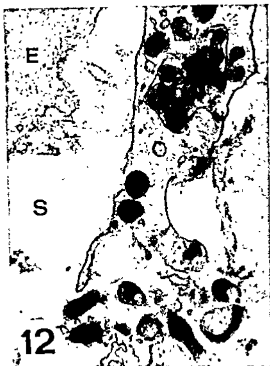

E
S
12

PLATE 4

EXPLANATION OF FIGURES

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'e Fine Structure of the Adrenal Cortex of the ·Lined Ground Squirrel [1]

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ABSTRACT The adrenal cortex of the 13-lined ground squirrel was examined by means of phase contrast and electron microscopy. At the most, two cell types were observed. One of these two types was observed in three morphologically different stages of activity. They had some common characteristics, although they were very different in general appearance. The findings substantiate Symington's ('60, '61) statements that the zona reticularis contains the active cells and the zona fasciculata contains the inactive cells of the deep adrenal cortex. The cell types do not stay within the bounds of the three classical zones, and it is suggested these be used for general topography only. A number of modifications and derivations of mitochondria are described, one of which lends strength to Lever's ('55) suggestion that mitochondria might be converted into lipoidal masses.

Sheridan and Belt ('64) state, "Fine ɪrphology of an organ is usually similar : most instances from one species to .other. This generality, however, does not ɪply in the case of the mammalian adreɪl cortex since there has been considerable ɪriability among the species that have ɜn observed."

'Due to the recorded differences and ɜcause the periductular steroid cells reɪmble and functionally replace the adreɪl cortex of the adrenalectomized 13-ɪed ground squirrel (Seliger, Blair and ɪossman, '66), an investigation of the ɪltrastructural morphology of the adrenal ɔrtex of this animal was considered a vital ɪrst step in our study of the origin, difɜrentiation and controlling factors of ɪese cells.

MATERIALS AND METHODS

Fourteen male and fourteen female 13-ɪned ground squirrels (*Citellus tridecemɪneatus*, Mitchell) were used in this invesɪigation. All had been trapped as juvenile ɪnimals and had been in captivity approxiɪnately a year.

A great deal of care was exercised to ɪttempt to keep the environmental condiɪions and the mechanics of sacrifice and ɪissue-handling constant. Half of the aniɪnals in each group were decapitated while asleep. The other half were carefully lifted from their cages with a gloved hand, carɪied rapidly to the laboratory and killed immediately by a sudden cervical blow. The adrenal glands were removed and immediately sliced serially while immersed under the fixative.

Fixation was accomplished with 6.25% glutaraldehyde, buffered to pH 7.7 with Sorensen's phosphate buffer, chilled to 4°C. After being rinsed in buffer, the tissue was stained with cold 1% osmium tetroxide in the same buffer. A graded series of ethyl alcohol was used for dehydration. Propylene oxide was the clearing agent. Epon 812 (Luft, '61) was used as the embedding material. The tissue was stained by uranyl acetate followed by lead acetate.

OBSERVATIONS

Phase contrast microscopic observations

Despite apparent control of environment and processing methods, phase contrast study of 2 μ sections reveals a wide variation in the adrenal cortices of the animals studied.

The outermost cells contain large, round or oval, smooth-walled vacuoles. These cells occupy the zona glomerulosa and, to a variable degree, the outer portion of the zona fasciculata. Their location varies in different specimens and, to some extent,

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e mentioned in the electron microic observations.

Electron microscopic observations

he round-vacuole cells are polygoned with vesicular, generally round lei. Cells located near the capsule are ewhat elongated and have flattened lei. The nuclei possess a narrow, rather ilar nuclear cistern and obvious nuclear s, evenly dispersed granular material the center, and peripherally concened chromatin which projects toward center at several regions, one of which often have a nucleolus on its summit. he extranuclear compartments contain ge numbers of round, membrane-bound uoles, the majority of which are usually ated on one side of the cell (usually that ing a blood vessel), causing the nucleus be eccentric. A flattened, elongated, anular sac or series of sacs are usually sely pressed against the outer surface these vacuoles.

The matrix of the cytoplasmic compartnt, including the intervals between cuoles, is moderately dense and contains sely scattered small vesicles of agranr endoplasmic reticulum interspersed th groups of ribosomes and mitochonla. The mitochondria are usually round oval. They have a dense, faintly granular atrix and contain primarily tubulo-saclar cristae. Most mitochondria contain me vesicular and lamellar-form cristae. perinuclear Golgi complex is often obrved, but occasionally one is located near e plasma-membrane.

Osmiophilic, membrane-enclosed rods e found in the perinuclear zone of the und-vacuole cells of some of the animals fig. 4). Many of the rods show clear areas ithin their confines (fig. 6). Often more an one rod is found in a membranous c. Large, irregular membrane-enclosed rms of similar texture and density also ave been observed. Some specimens also how similar rods which are not membranenclosed, but merely surrounded by the ytoplasmic matrix. Some of these cells ontain osmiophilic, membrane-enclosed orms which vary from round to club or up shaped. These structures have not been nalysed but have the ultrastructural apearance of lysosomes.

In the deeper regions of the round-vacuole cell layer, there is a gradual change. First the mitochondria become modified by a loss of the dense matrix and a loss or reduction of the cristae. Often the cristae are seen in longitudinally organized bundles of tubular membranes. These modified mitochondria commonly show internally located membrane-enclosed areas, either clear, full of osmiophilic material, or as multilayered, partially collapsed sacs. These latter occasionally appear to be in the process of being extruded from the mitochondria (fig. 8), and other examples located near the mitochondrial wall appear to have their outer membrane fused with the inner membrane of the mitochondrial wall. Similar multilayered, collapsed figures are occasionally seen in the cytoplasmic compartments of these cells.

The first irregular vacuoles observed deep to the round vacuoles are round with a scalloped outline (fig. 5). These cells also contain small vacuoles which appear to be enlarged examples of the vesicular endoplasmic reticulum of these cells, and it appears as if the scalloped outline of the irregular vacuoles is caused by the remnants of the small vacuoles which are coalescing into the large vacuoles. The deeper lying irregular vacuoles have the outlines of partialy collapsed sacs (fig. 7). Often one of the latter will have a pseudopod-like process which extends into an adjacent vacuole or mitochondrion, and occasionally a mitochondrion will have a similar process which extends inside of an irregular vacuole.

Lysosome-like structures appear in higher numbers in these cells than in the previous form. The cytoplasm is filled with vesicular, agranular endoplasmic reticulum which is interspersed with groups of ribosomes.

Many of these cells show dilated nuclear cisterns which have a varicose appearance and accentuate the nuclear pores. The dilated nuclear cisterns are commonly associated with the presence of the small vacuoles described above.

The electron photomicrographs of the transitional halo cells show a sparsity or, most often, an absence of vacuoles. They show the presence of vesicular endoplasmic reticulum and, occasionally, tubules of

in various areas of the same specimen (fig. 1). Occasionally cells of this type extend deep into the zona fasciculata, and some sections show a transition zone followed by another layer of these round-vacuole cells. Occasionally an isolated cell of this type is found anywhere in the other parts of the adrenal cortex. These cells are arranged in compact, block-like groups, two or three cells wide, with blood vessels and connective tissue stroma separating adjoining groups. The large, round vacuoles are of various sizes but essentially of one size within any one cell. Most of the vacuoles appear clear and refractile. A small number of cells, often near the capsule, show vacuoles which are filled with osmiophilic material or vacuoles which have peripheral zones of osmiophilic material and clear centers. The various types are usually uniform for an individual cell. To determine if different degrees of osmium penetration caused this difference, usual fixation and osmification procedures were employed on some tissues. These blocks were thoroughly rinsed and re-osmificated with extensive agitation. Most of the vacuoles appear clear. These findings, plus some of our observations on cells showing clear vacuoles and also showing intensely osmificated structures within the confines of the same cells, lead to the conclusion that the various forms of these vacuoles indicate the degree of saturation of their enclosed lipoidal material.

The second cell form, the irregular-vacuole cell, often appears to blend gradually with the round-vacuole cell. In some deeper areas, the round-vacuole cells contain both irregular, non-refractile vacuoles and smooth, round, refractile vacuoles (fig. 1). In areas like this it is impossible to draw a line of demarcation between the two cell forms. In other areas the two cell forms appear as contiguous cells and a distinct line of demarcation can be drawn. Between these two cell forms, cells are found occasionally which show no cytoplasmic detail with phase contrast microscopy. This cell is designated as a transitional halo cell and is associated morphologically with the halo cell (fourth cell form).

The cytoplasmic compartments of the irregular-vacuole cells are filled with non-refractile, irregular, "clear" spaces. This gives a foamy appearance to the cells (fig. 2). Examination with the phase contrast microscope at high magnification discloses that many of these "clear" spaces actually do contain light gray material. The electron photomicrographs show that these spaces containing the light gray material are actually modified mitochondria.

The irregular-vacuole cells are found deep to the round-vacuole cells, and their extent varies. They vary inversely with the fourth form, the halo cell, which occupies the remainder of the deep adrenal cortex to the cortico-medullary junction, except for one or two layers of transitional halo cells (third form) sandwiched between the irregular-vacuole cells and the halo cells. Three of the decapitated animals' adrenal cortices show these irregular-vacuole cells present, almost exclusively, deep to the round-vacuole cell layer and extending to the medulla. The remaining decapitated animals and a number of hand-held control animals have irregular-vacuole cells deep to the round-vacuole cells, occupying from half to three-quarters of the remaining deep adrenal cortex. The balance of hand-held control animals show layers of irregular-vacuole cells that vary from a narrow, irregular layer to approximately a third of the cortex deep to the round-vacuole cells.

The halo cells are designated "halo" cells because the nuclei are closely encircled to a varying degree by gray, halo-like masses. The gray masses vary: Some appear to be homogeneous, some to be composed of small gray particles, and others have irregular or flattened clear spaces scattered throughout them (fig. 3). The electron photomicrographs show the gray masses to be concentrated accumulations of mitochondria. Except for size, these halo cells are the same, whether they are found in the zona reticularis or the zona fasciculata. The irregular-vacuole cells and the halo cells occupy the majority of the cortex deep to the round-vacuole cells and are inversely proportional to each other in the volume they occupy.

Small black structures become common in the deeper cells of the round-vacuole cell area and occur with a greater frequency in the deeper cells of the adrenal cortex. They are the lysosome-like bodies

ɩample of both in the round-vacuole ;egion of the gland. We agree with ('55) that these are not degenerative but rather are probably in a different of activity. The "dark" cells appear : a greater concentration of mitodria, dense cytoplasmic matrix and compact and irregular nuclei.

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Zonation

Phase contrast study of 2 µ sections of ·ie adrenal cortex shows great variability f the cells' locations and boundaries plus ·he apparently grandual blending of the ı'ell forms. Since special care had been aken to attempt to control the conditions f the care and sacrifice of these animals, his was very unexpected. In an attempt o understand this, sections were cut from blocks of Epon-embedded tissues from a pilot study concerned with stimulation and inhibition of the adrenal cortex. Two ani- mals had been killed — one hour and 24 hours — after administration of 4 units/kg of ACTH gel (stimulated animals). Two animals (for each drug) had been killed — one and 24 hours — after receiving 10 mg/kg cortisone acetate; 10 mg/kg hydroxycortisone acetate; and 5 mg/kg desoxycortisone acetate (inhibited animals).

All one-hour animals showed a rather normal pattern of cells, typical of the handheld control animals. This consisted of an outer layer of round-vacuole cells followed by a variable layer of irregular-vacuole cells, not exceeding half of the remainder of the adrenal cortex. The remainder of the area down to the junction with the medullary cells was filled with halo cells. The tissues from the 24-hour animals which were inhibited showed predominantly irregular-vacuole cells deep to the round-vacuole cell layer, and the stimulated glands showed halo cells, almost exclusively, deep to the round-vacuole cell layer.

This information is the result of only a pilot study, but it indicates that the wide variation that occurred in the adrenal cortices of the control glands of these animals was due to a variation in the degree of stress they felt under seemingly identical environments. These findings substantiate Symington's ('60, '61) statements that the zona reticularis contains the active cells and the zona fasciculata the inactive cells of the deep adrenal cortex.

DISCUSSION

Our findings in the 13-lined ground squirrel's adrenal cortex indicate that the three classical zones should be used only for general orientation and localization. Since there is considerable variation in the location of the cells, the glands should be subdivided according to cell forms and the volumes they occupy. This great variability of the cells' locations and boundaries, plus the gradual blending of cell forms, suggests that no more than two cell types, in various stages of production and activity, are involved. The variation in the ratio and position of the cell forms appears to be a result of hidden stresses.

It is naive to believe that animals kept under outwardly identical environmental conditions would be under the same degree of stress. The general health of these animals could vary a great deal without being obvious. Variation in the degree of adjustment to captivity, the cage size, different eating and drinking habits, the odors of

endoplasmic reticulum organized into bundles which present a honeycomb-like appearance on cross section (fig. 11). There are no ribosomes or obvious matrix present between these tubules. Most of the mitochondria are elongated and thin except at the ends, and are arranged into cupped masses which DeRobertis and Sabatini ('58) refer to as "chondriospheres" (fig. 10). A smooth-walled membranous sac is pressed intimately between the adjacent mitochondria of these masses (fig. 10). This arrangement causes the contiguous mitochondrial surfaces to appear thickened or more intensely stained. The internal membranes of these mitochondria are vesicular and occasionally tubular, but the intense contrast between matrix and internal membranes which is seen in other mitochondria is absent.

Although the general appearance of these cells is very different from the halo cells, the mitochondria-membranous sac relationships are the same. Intermediate forms containing chondriospheres as well as the aligned mitochondria which are characteristic of halo cells have been observed (fig. 11).

The halo cells occupy a variable amount of the deeper portion of the zona fasciculata and the entire zona reticularis. The cells of the zona reticularis are smaller and lose the column-like arrangement, but they are morphologically the same in both zones. The nuclei are generally round, with peripherally concentrated chromatin and often contain a central nucleolus which may be in a ring form or in the form of a folded nucleolonema. The nuclear cisterns vary from the narrow, regular type to the varicose type which is associated with the dilation of certain intermitochondrial sacs to be described.

The mitochondria occur in packed masses which closely encircle the nuclei to a variable extent. They vary in shape from round or elongated to branching, in the same cells. Their internal membranes are tubulo-saccular, vesicular and lamellar-form in one mitochondrion. Occasionally, a number of very elongated mitochondria occur in parallel groups orientated radially from the nucleus with parallel lamellar-form cristae running the entire length of the mitochondria (fig. 16). Elongated sacs of granular endoplasmic reticulum occur between adjacent mitochondria (figs. 12, 13). They are often flattened to the extent that the lumina are very narrow. Halo cells which have varicose nuclear cisterns and a preponderance of modified mitochondria usually show dilated intermitochondrial sacs (figs. 14, 15). No direct connection has been observed between the nuclear cistern and these intermitochondrial sacs. Occasionally, cells from the junction between the outermost halo cells and the transitional halo cells show mitochondria arranged in this same manner but possessing agranular intermitochondrial sacs (fig. 11).

The cytoplasmic compartments of the halo cells are packed with vesicles of smooth endoplasmic reticulum which are interspersed with small clumps of ribosomes.

Generalities

Centrioles are commonly found in cells of all forms and regions (fig. 13). A cilium was observed in one round-vacuole cell. Wheatly ('67) recorded similar observations in the rat adrenal cortex.

The lysosome-like structures are present in cells of all forms. They are least common in the round-vacuole cells. In the deeper regions, similar dark structures are seen which contain empty round vacuoles, several of which are often larger than the osmiophilic portions of the structures (figs. 15).

A multivacuole structure has been observed in cells of all forms. This large structure is full of empty-appearing vacuoles which usually maintain a slightly stained, contant-width interspace which gives the appearance of a network of tubules (fig. 17). Select sections show a double-membrane outer wall, and sufficient examples of transition between these structures and mitochondria have been observed to state that they are derived from these organelles (fig. 18). Some of these structures have thick walls, which are made up of concentric layers of osmiophilic lamellae giving them a myelin-like appearance.

"Light" and "dark" cells have been observed among all cell forms and in all zones of the adrenal cortex. Figure 4 contains

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other animals, noises, and lighting could affect the animals differently. Jones ('63) felt the 13-lined ground squirrels he used were subjected to different degrees of stress due to the positions of their cages in the battery.

Although some viruses — especially insect viruses — are rod-shaped, and incomplete ones could have clear areas within them, the described rods and the similar large irregular masses are probably similar to the rods of crystalloid material shown in the interstitial cells of the testes (Fawcett, '66). To date, demonstration of a crystalloid matrix in these rods and masses has not been possible.

The mitochondria seem to have no definite shape in the various parts of the adrenal cortex, but most of them appear round or oval on section. They seem to be more commonly elongated in the deeper-lying halo cells.

The mitochondria containing the membrane-bound osmiophilic structures indicate that some product is formed and is being stored in the mitochondria. Another internal configuration, myelin figures, appear to be formed within the mitochondria by apposition of ever increasing numbers of membranes. The myelin-like figures have been observed apparently penetrating the walls of the mitochondria. It is not certain they are being extruded, but since it appears they are built up within the mitochondria, this assumption seems logical. These forms may collapse after voiding their contents to the exterior of the mitochondria. The single, clear, round, membrane-bound vacuoles seen within the mitochondria may be the beginning of these myelin-like figures.

The large, multivacuole structures described appear to be the result of modification and enlargement of mitochondria. What appear to be more advanced stages show thick osmiophilic laminated walls. Some of the above observations appear to reinforce Lever's ('55) statement that mitochondria may form the lipoidal material of these cells. When one of these structures is present in a cell, it is common to see others within the confines of that cell.

There is an almost constant relations[illegible] between the degree of varicosity of [illegible] nuclear cistern and the degree of dilatio[illegible] of some of the endoplasmic reticul[illegible] membranes. The rough membranes th[illegible] occur between the organized mitochondri[illegible] in the internal cells are often very dilated. Since the nuclear cistern is considered t[illegible] be continuous, to some degree, with th[illegible] endoplasmic reticulum, this is not an u[illegible] expected finding. This same relationsh[illegible] also seems to exist in the irregular-[illegible] cells, with some of the smaller vacu[illegible] or membranous sacs. It is our belief [illegible] these smaller vacuoles are actually dila[illegible] examples of the vesicles of the endoplasm[illegible] reticulum which fill these cells.

ACKNOWLEDGMENT

We would like to acknowledge Docto[illegible] P. Epling, of our department, for his b[illegible] advice, and for allowing us to use var[illegible] pieces of equipment from his elect[illegible] microscope laboratory.

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PLATE 1

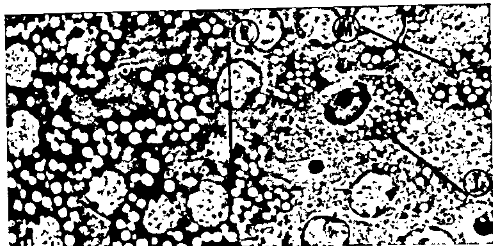
EXPLANATION OF FIGURES

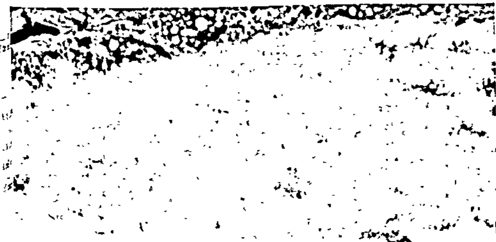
In all three photomicrographs the small, dark structures in the cytoplasmic compartments are the lysosome-like structures of the electron photomicrographs.

1 Area of the junction of round-vacuole cells (R) with irregular-vacuole cells (I). Mixed form (M) containing mostly round vacuoles. Hand-held control female animal, phase contrast photomicrograph. × 1,600.

2 Area full of irregular-vacuole cells from the zona fasciculata. Hand-held control female animal, phase contrast photomicrograph. × 1,600.

3 Cells with "halos" of mitochondria (H) around the nuclei. A cell with dilated intermitochondrial sacs (D). The nuclei of the latter are usually darker and have more irregular outlines. Hand-held control female animal, phase contrast photomicrograph. × 1,600.





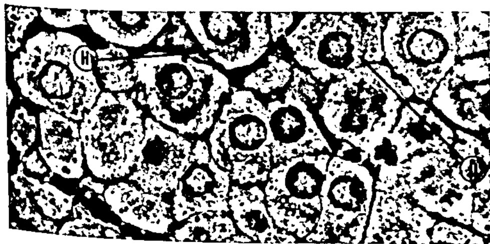


PLATE 2

EXPLANATION OF FIGURES

4 Round-vacuole cells from the zona glomerulosa. Portions are seen of two "light" cells, left, and two "dark" cells, right. Membrane-bound rods (R) are seen in the "light" cells. Modified mitochondria (MM) and a Golgi complex (G) are seen in the "dark" cells. Hand-held control female. × 6,400.

5 Portions of cells from deep in the round-vacuole cell layer. Cell to the right shows scalloped irregular vacuole (S) with surrounding small vacuoles. The mitochondrion below the scalloped vacuole has a process (P). The cell to the left shows a typical round vacuole (R) for comparison. Hand-held control female. × 16,000.

6 A group of membrane-enclosed rods and two of the larger irregular structures of similar texture. The nucleus of this round-vacuole cell is just evident in the upper right corner (from the zona glomerulosa). Hand-held control female. × 28,000.

R
MM
G
S
4

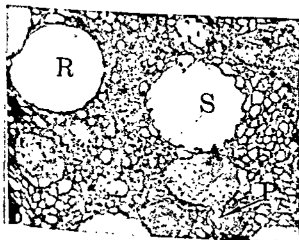




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9

PLATE 3

EXPLANATION OF FIGURES

7 Portions of four irregular-vacuole cells. Modified mitochondria (MM) and irregular vacuoles, many of which have pseudopod-like processes (P), are very plentiful. The small vacuoles (sV) which appear to be dilated, vesicular endoplasmic reticulum also are very plentiful. Hand-held control female. × 9,000.

8 Mitochondria from an irregular-vacuole cell, one of which shows tangentially sectioned, organized tubular cristae. The other shows myelin figure-like structures, one of which appears to be in the process of being extruded (arrow) (from the zona fasciculata). Hand-held control male animal. × 25,000.

9 A portion of an irregular-vacuole cell showing the extensive contact often present between the modified mitochondria (arrows) and the clear-appearing irregular vacuoles (from the zona fasciculata). Hand-held control female. × 22,000.




9

PLATE 4

EXPLANATION OF FIGURES

10 Portion of a transitional halo cell. Mitochondria are elongated and cupped into chondriospheres. Arrows indicate the ends of the collapsed agranular sacs located between contiguous mitochondria (from the outer zona fasciculata). Hand-held control female. × 23,000.

11 Transitional halo cell from a slightly deeper portion of the gland than the cell shown in figure 10. It shows an intermediate form containing both chondriospheres (right) and aligned mitochondria similar to those seen in halo cells (left). The arrows indicate the ends of the intermitochondrial sacs. Surrounding these mitochondria is organized tubular agranular endoplasmic reticulum viewed on cross section (from the outer zona fasciculata). Hand-held control female. × 13,000.

12 Portion of a halo cell showing the organization of mitochondria and granular intermitochondrial sacs (from the zona reticularis). Hand-held control female. × 10,000.

13 Portions of two halo cells showing a centriole (C) and cell junction complexes (J). Hand-held control female. × 9,000.









PLATE 5

EXPLANATION OF FIGURES

14 Portions of two halo cells. Cell A shows marked dilation of the nuclear cistern, the intermitochondrial sacs and a large number of the small vacuoles believed to be enlarged vesicles of smooth endoplasmic reticulum, and modified mitochondria. Cell B shows a narrow, regular nuclear cistern, undilated intermitochondrial sacs, unmodified mitochondria and a more regular cytoplasmic matrix (from the zona reticularis). Hand-held control female. × 5,600.

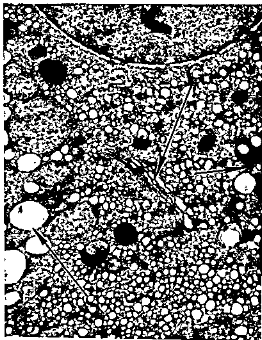
15 A cell similar to that shown in figure 14 showing dilated, granular intermitochondrial sacs cut on cross section (S), a Golgi complex (G), and several of the osmiophilic bodies containing large clear vacuoles (arrow). Hand-held control female. × 15,000.

16 Portion of a halo cell showing various forms of mitochondria seen in these cells. Hand-held control female. × 15,000.

17 Multivacuole structure (arrow). Hand-held control female. × 6,000.

18 These two mitochondria are the best examples observed of the transition between the multivacuole structures and mitochondria. The dark granular material is precipitate. Hand-held control male. × 18,000.











e Innervation of the Bone Marrow .Laboratory Animals [1,2]

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ABSTRACT The nerve pattern of the bone marrow of the monkey, rabbit, rat and mouse was studied in serial sections specially stained to demonstrate myelinated and nonmyelinated nerve fibers. Features that are common to the different species studied were found in the bone marrow of the femur of the rat in a simple arrangement.

A scheme is proposed based on thirteen photomicrographs demonstrating the nerve pattern of the bone marrow and the relation of the nerve fibers with the arterial tree, the sinusoids and the blood forming cells. This scheme shows that the necessary anatomical conditions exist permitting a direct influence of the nervous system on the function of the bone marrow. These conditions are the presence of myelinated nerve fibers traversing the parenchyma of the marrow, and the existence of nonmyelinated fibers ending in sphincterlike structures at the origin of the arterioles.

Single myelinated nerve fibers and small bundles of both myelinated and nonmyelinated fibers lie in close contact with the endothelial wall of the sinuses during long trajects. Only the very thin endothelium of the sinuses separates the newly formed blood and the nerve fibers. An interaction between both elements is possible along these areas of contact.

The function of an organ cannot be fully iderstood without considering also the rvous system, yet the study of the innertion of the bone marrow has long been glected. Its study is of interest in normal orphology and physiology, and may prove be of importance in pathology.

The purpose of this paper is to present servations concerning the presence, the iture and the distribution of the nerve ers in bone marrow of normal adult aniəls commonly used in the laboratory, as ell as the relations between the nerve fiers and the cells of the parenchyma, the rterial vessels and the sinusoids. It is oped that the description of the normal rchitecture of the bone marrow, with the erve pattern included, may throw some ght on the understanding of the mechnisms that regulate the proliferation and elease of the blood cells, and also on the laintenance of homeostasis and on the daptation of the function of the bone narrow to different stimulations. The hotomicrographs of normal nerve fibers lemonstrated here with well standardised istological methods may serve later for omparison with pathological material.

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MATERIAL AND METHODS

The bone marrow of the long bones of the normal adult mouse, rat, rabbit and monkey was studied. For the demonstration of the nonmyelinated fibers, the marrow was impregnated en bloc by the silver nitrate method of Cajal after fixation and decalcification of the bone with the fluid of Castro ('25). The femur, tibia and humerus of the small animals (mice and rats), with their bone marrow inside, was embedded in paraffin and cut entirely in serial sections, 10 μ thick. This method permits the study of the bone marrow in relation to the bone. The marrow of the large animals (rabbit and monkey) was extracted from the bone as cylinders of approximately 15 mm in length and impregnated with the silver nitrate. Cylinders of marrow from the area of the nutrient foramen, the diaphysis and the distal epiphysis were studied separately. From these regions, serial sections were cut and at intervals a number of slides were counter-

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stained with the Masson's trichrome or with Hematoxylin-Eosin. The sections counterstained with these methods provided evidence of the selectivity of the silver impregnation for nerve fibers and gave information concerning the relations between the nerve fibers and the other structures of the bone marrow. Blocks of tissue from the same region were fixed in formalin and stained with the Luxol fast blue-Periodic-acid-Schiff-Hematoxylin technique as described by Margolis and Pickett ('56) for the demonstration of the myelinated fibers and the blood vessels. The reticulum fibers were stained in sections of this formalin fixed material with the method of Wilder and compared with the nerve fibers for additional control of selectivity of the silver impregnation of the nerve fibers by the Cajal — Castro method. In some cases, the nerve and the nutrient artery with a fragment of tissue at the level of the nutrient foramen were fixed and stained with osmic acid for demonstrating myelinated fibers and fat. A total of 330 long bones were prepared of which 27.000 sections were cut and studied.

RESULTS

General findings

The long bones of the animals have a small foramen in a constant position, the so-called nutrient foramen. The nutrient artery and a nerve pass through foramen, providing the blood supply and the nerve supply for the marrow. This is well defined from its origin in the spinal nerve that corresponds to the territory in which the foramen is located, until its termination in the periosteum and the bone marrow. Following the nomenclature given by Gros (1846) we call diaphysary nerves those nerve trunks extending between the corresponding spinal nerve and the foramen. The diaphysary nerve gives branches that spread over the periosteum and also branches that penetrate into the bone cavity. The latter are the medullary branches of the diaphysary nerve and constitute the subject of our study. The medullary branches immediately upon entering the marrow cavity, divide repeatedly and give off numerous nerve bundles that follow a distribution roughly parallel to the branches of the nutrient artery [illegible] throughout the marrow, both distally [illegible] proximally to the nutrient foramen. [illegible] nerve bundle contains a number of [illegible] fibers surrounded by a thin [illegible] connective tissue, the perineurium. [illegible] bundles have both myelinated and [illegible]myelinated fibers. The nonmyelinated [illegible]bers are thinner than those covered [illegible] myelin. With the reduced silver nitrate [illegible] Cajal — Castro, these fibers are [illegible] impregnated by the silver salt and [illegible] as black wirelike structures sharply [illegible]marcated on a yellowish background. [illegible] structure which becomes impregnated [illegible]responds to the neurofibrillary [illegible] of the nerve fiber. The neurilemma [illegible]mains almost unstained but it can be [illegible]tinguished as a pale sheath around [illegible] neurofibrils. The oval nuclei, sparsely [illegible]tributed along the nerve fibers, are [illegible] of the Schwann cells that form the [illegible]emmal sheath. The different cells [illegible] connective tissue fibers of the [illegible] are faintly stained but the various [illegible]tural types of blood vessels are [illegible]ognizable. The distinction between [illegible]les and capillaries is very easy owing to [illegible] clear demarcation of the muscular [illegible] of the vessels in which every smooth [illegible]cle cell is stained brown with the [illegible]lar method used and appears clearly [illegible]rated from the others.

The myelinated fibers are thicker [illegible] less numerous than the nonmyeli[illegible]. They can be found isolated among [illegible] cells of the bone marrow or in the [illegible] nerve bundles intermixed with [illegible]linated fibers. The osmic acid stains [illegible] myelin black leaving the nonmyelinated [illegible]bers unstained. The relation between [illegible]linated fibers, cells and blood vessels [illegible] be beautifully demonstrated by the Lu[illegible] fast blue-Periodic acid-Schiff-Hem[illegible] method. The myelinated fibers are stai[illegible] in sky blue color, the blood vessels [illegible] bright red and the the nuclei of the [illegible] in blue-violet.

In most of the long bones studied [illegible] nerves enter the bone cavity at one [illegible] only, but in the humerus of the rat [illegible] are two separated entries. The main [illegible] is through the nutrient foramen located [illegible] the proximal part of the bone. A small[illegible] entry is located near the trochlea. [illegible]

l innervation of very small areas of marrow was observed in the long s, by the entry of single nerve fibers ıgh some of the largest Haversian ls.

summary then, except for the minor e contributions mentioned above, most ıe bone marrow is innervated by nerve ıches entering with the nutrient artery ugh the nutrient foramen. The nerve ıches divide repeatedly and are related a) the arterial component of marrow ulation (b) the sinusoidal part of the row circulation and (c) the parenma.

a) *Innervation of the arterial tree.* : outstanding features of the innervaı of the bone marrow is the close associ-ın of the nerve pattern with the arterial tem. All arteries of the marrow are ompanied by nerve bundles (fig. 1) t form a wide-meshed network with nches often winding around the arter-: The main branches of the nutrient ery are surrounded by a number of ve bundles each of them containing merous nerve fibers (fig. 2). The thinr branches of the artery are accompanied only one or two bundles, each having y two or three nerve fibers (fig. 3). The erioles and the capillaries often are acnpanied by only one naked fiber atched to the vessel wall. The thicker rve bundles are individually ensheathed a thin membrane of connective tissue d may lie either in close contact with e arterial wall, or run parallel to the vesl, leaving a few cells of the parenchyma tween nerve and artery. Thin nonmyeated fibers leave the nerve bundles, ach the wall of the arteries and spread nong the smooth muscle fibers dividing peatedly into branches that end in conct with the muscle fibers. Particularly ch is the innervation of the circular layer the smooth muscle fibers located around e opening of the initial part of some of e branches of the nutrient artery (fig.). The presence of well innervated annuır muscle fibers is a characteristic of the hincters working under immediate nervus control.

The capillaries are in contact with very in nonmyelinated fibers only in certain rts of their course. The fibers may follow the capillary in intimate contact with its endothelial wall for some distance, then leave the vessel and join another capillary that results from the division of a neighbouring vessel (fig. 5). It appears as if a number of these very thin nerve fibers never reach another vessel but divide further and eventually end among the cells of the parenchyma.

(b) *Innervation of the sinusoidal tree.* The sinusoids appear to be poorly innervated in comparison with the arterial vessels. In some sections of the diaphysary marrow it is possible to see isolated myelin fibers alongside the wall of the central sinus (fig. 6). They can be followed for a long way, straight and undivided, changing their course only to curve a little around the walls of the mouths of the sinus recti, which in this area enter the central sinus perpendicularly. In the epiphysary marrow, the central sinus ramifies towards the trabecular bone of the epiphysis following the long axis of the bone. Some of the myelinated fibers described along the central sinus in the diaphysary marrow continue along the wall of the sinusoids as they ramify towards the epiphysis. The fibers become very thin and disappear among the cells of the parenchyma or end in contact with the endothelium of the sinusoids. Other fibers, either isolated or in loose bundles, run parallel to the central sinus approximately at middle distance from the sinus to the endosteum, establishing contact with numerous sinusoids (fig. 7). The small bundles of nerve fibers become loose because they are not ensheathed by the perineurium as in the main nerve trunks. When the bundles meet the wall of a sinusoid, their fibers spread and follow the rounding of the sinusoidal wall, rejoining again on the other side as the bundle proceeds towards the epiphysis. In this form all the fibers of each bundle make contact with the endothelium of numerous sinusoids.

Myelinated and nonmyelinated fibers that lie in contact with the wall of the sinusoids are separated from the blood only by the thin sinusoidal wall. The fibers end by dividing into extremely thin naked fibers that taper away in contact with the endothelium of the sinuses.

(c) *Innervation of the parenchyma.* From repeated division of the nerve entering the bone cavity, sharply demarcated nerve bundles detach and proceed proximally and distally across the parenchyma of the bone marrow following the length of the bones independently of the arterial branches. An exchange of fibers occurs between some of the nerve bundles in the form a chiasmas (fig. 8). The nerve bundles divide into thinner branches in their course towards either epiphyses. They are composed of a number of nerve fibers (two to seven) both myelinated and nonmyelinated. Other branches consist of a few nonmyelinated elements.

The nerve bundles give off single myelinated fibers (fig. 9) that travel a long way undivided across the parenchyma before they end by ramifying freely among the blood forming cells (fig. 10).

No one cell type or group of cells of the parenchyma appear to have special attraction for the nerve fibers. The same single nerve fiber touches successively but indiscriminately every cell that lies in its path. Nonmyelinated fibers follow a more tortuous course. After a long path among the cells of the parenchyma they end in the wall of an arteriole, on a capillary (fig. 11), or wander for some distance among the blood forming cells as very thin naked fibers (i.e., nerve fibers without Schwann cells).

The usual linear course of the fibers is occasionally modified by parenchymal cells lying in their path (fig. 12).

Fat globules appearing in the way of the nerve fibers also cause them to take a curved path (fig. 13).

A few nonmyelinated fibers turn from the axial direction towards the surface of the marrow cylinder ending among the elements of the endosteum or penetrating occasionally into a Haversian canal. The Haversian canals can therefore be innervated by either fibers entering the bone cavity from the periosteum or by fibers originating within the parenchyma as branches of the medullary nerve bundles.

The nerve fibers are asymmetrically distributed in the bone marrow. They are abundant in the vicinity of the nutrient foramen (upper third of the femur), but sparsely distributed in the distal epiphyses. The nerve fibers are mostly to be found midway between the endosteum and the central sinus of the marrow cylinder.

DISCUSSION

The above separate description of the innervation of the sinusoids and innervation of the parenchyma is artificial, and was intended for descriptive simplicity since sinusoids and blood forming cells form a labyrinth of vessels and islands of cells intimately related to each other.

The long bones of the laboratory animals have a bone marrow with similar architectural characteristics. In the marrow of the femur of the rat exist the basic anatomical details in almost diagrammatic simplicity. Therefore this marrow was chosen for the detailed description.

The findings presented leave no doubt that the bone marrow is innervated by myelinated and nonmyelinated nerve fibers. These fibers form a characteristic pattern that in part parallels the vascular architecture and in part appears related to the cellular parenchyma. The *vascular architecture* has been studied extensively by Fliedner et al. ('56) who proposed a schema that describes the marrow as an organ with a well defined relation between vessels and parenchyma. The present study complements their description by adding the nerve pattern to the study of the bone marrow as a well innervated organ. It now appears possible to propose the following scheme (fig. 14) for the relation between nerves, vessels, and marrow cells. The wide central sinus collects the blood carried by the sinusoids. The arteries divide giving branches on either side of the central sinus and end in capillaries that pervade the marrow throughout. The arteries and arterioles are shown as tubes with lines across that represent smooth muscle fibers, while in the capillaries these fibers are absent. In the upper part of the figure a fragment of the diaphysary bone appears perforated by innervated Haversian canals.

The nerve pattern comprehends the main findings and is composed and based on the 13 photomicrographs described in the previous pages. The number of the photomicrographs corresponds with the same number on the scheme. The relation between the nerve fibers and the nutrie

ssels are shown in numbers one to five. mulation of these nerve fibers will in-ence the blood flow by changing the meter of the lumen of the arteries that pends on the degree of contraction of innervated smooth muscle fibers of the terial wall. The nerve bundles that ap-ar to have a course independent of the itrient blood vessels are described as re-ted to the parenchymal cells and the usoids. The numbers 6 to 13 illustrate e peculiarities of these relations. In mber 7 a group of myelinated fibers versing the parenchyma spread and nd around the wall of the sinusoids that pear across their path. Changes in diam-er of the sinusoids are likely to stimulate e nerve fibers. Myelinated nerve fibers are so bent by the cells, as demonstrated in mber 12. An increase in cellularity can rve the nerve fibers in many areas caus-g an elongation of the fibers that is also kely to originate nerve impulses. Only a ry thin, fenestrated endothelium sepa-ttes the newly formed blood from the erve fibers that lie in contact with the all of the sinuses. This circumstance ermits an interaction between nerve fi-ers and blood. The composition of the ewly formed blood may affect the conduc-ivity of the nerve fibers, while the stim-lation of the nerve fibers may induce the elease of substances that influence the ermeability of the vessels' wall.

The presence of myelinated fibers in the one marrow was mentioned by Variot und emy (1880) and by Castro ('29) and ere interpreted as afferent fibers respon-ible for the transmission of nerve impulses o the brain where they could be evaluated s pain. The existence of these afferent ibers is well proved by the repeated clin-cal observation of *patients that feel pain* during the performance of a bone mar-row aspiration.

The nonmyelinated fibers were first found in the marrow by Variot and Remy (1880). Ottolenghi ('02) described the in-timate relations that these fibers establish with the arteries, and Castro ('29) dem-onstrated anatomically the autonomic na-ture of the nerves of the bone marrow of the occipital bones by following the nerve fibers in serial sections to their origin in the superior cervical ganglion.

An indirect influence of the nervous system on the blood cell count by way of nerve fibers carried by the splanchnic and vagi nerves ending in the abdominal vis-cera, was schematically presented by Ko-miya ('56). Our scheme demonstrates that the myelinated and nonmyelinated nerve fibers carried by the spinal nerves estab-lish intimate anatomical relations with the bone marrow structures permitting a *di-rect* influence of the nervous system on the bone marrow function. This direct influ-ence does not exclude but rather comple-ments the other regulatory mechanisms that are known to influence the physiology of the bone marrow.

ACKNOWLEDGMENTS

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PLATE 1

EXPLANATION OF FIGURES

Bone marrow, femur, rat. Reduced silver nitrate of Cajal-Castro

1 One of the main branches of the nutrient artery divides. The smooth muscle fibers are clearly demonstrated. Two of the nerve bundles (arrow) accompanying the blood vessels are visible in this section. Each nerve bundle is composed of numerous nerve fibers. × 500.

2 Cross section of one of the main branches of the nutrient artery in the vicinity of the nutrient foramen. The blood vessel appears surrounded by five nerve bundles, each containing a different number of nerve fibers. × 500.

3 An arteriole accompanied by three nerve fibers. The nerve fibers contact the layer of smooth muscle fibers of the arterial wall. × 800.


2


3

PLATE 2

EXPLANATION OF FIGURES

Bone marrow, femur, rat. Reduced silver nitrate of Cajal-Castro

4 A thin branch of the nutrient artery gives off an arteriole on the right hand side of the figure. One of the nerve fibers accompanying the artery (arrow) ends in a bulbous sphincterlike structure at the beginning of the arteriole. × 500.

5 In the center of the figure a capillary divides (arrow). One isolated nerve fiber bends at the level of the division and follows one of the branches of the vessel in intimate contact with its endothelial wall. The fiber appears interrupted in the photograph but it can be followed in the serial sections as it crosses from the left (top) to the right of the vessel wall (bottom of the figure). × 800.

6 A thick nerve fiber (arrow) is visible in the wall of the central sinus. Only the endothelial wall of the sinus separates the fiber from the newly formed blood carried by the vessel. The nerve fiber disappears in the center of the figure bent by the sinus ending perpendicularly to the central sinus, but in the serial sections the fiber can be followed a long distance attached to the wall of the sinus. × 800.


4


5


6

PLATE 3

EXPLANATION OF FIGURES

Bone marrow. Femur. Rat.

7 In the lower half of the figure, an arteriole is followed by a nerve bundle. Another bundle of nerve fibers appears out of focus in the upper left of the figure and becomes clearly visible (arrow) at the right, where the nerve fibers spread and curve around one of the sinusoids appearing in the path of these fibers. Reduced silver nitrate. × 800.

8 The lower half of the figure shows an exchange of fibers between two nerve bundles, producing a chiasma. In the upper half, another nerve bundle appears at the right. Reduced silver nitrate. × 800.

9 A myelinated nerve fiber traverses the parenchyma in intimate contact with the various cell types of the marrow. The arrow shows a node of Ranvier. Osmic acid. × 2,500.


8


9

PLATE 4

EXPLANATION OF FIGURES

10 A nerve fiber (arrow) ramifies in the center of the figure, ending among the cells of the parenchyma. Bone marrow. Femur. Rat. Reduced silver nitrate. × 2,000.

11 Nonmyelinated fiber (arrow) reaches the wall of a capillary, divides and disappears among the cells in the vicinity. Bone marrow. Femur. Rat. × 800.

12 Myelinated fiber arching around a cell at the right hand side of the figure. The myelin appears as an unstained sheath around the fiber. The nucleus of a Schwann cell is shown at the left (arrow). Bone marrow. Femur. Rat. × 2,000.

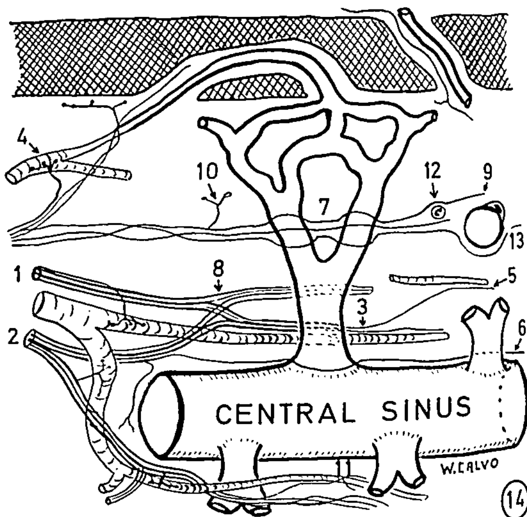
13 Two nerve fibers following the curvature of a fat globule in the center of the figure. Bone marrow. Femur. Rabbit. × 800.


10




12





EXPLANATION OF FIGURE

14 Schematic representation of nerve pattern of bone marrow. The relation of the nerve fibers with the arterial tree, the sinusoids and the parenchyma of the organ are based on the 13 photomicrographs described in the text. The numbers of the photographs correspond with the same numbers in the scheme. The innervation of the nutrient vessels are shown in numbers one to five. The innervation of the sinuses and the parenchymal cells in numbers 6 to 13. Harversian canals innervated by fibers originated from nerve bundles within the bone marrow and in the periosteum are represented in the upper part of the figure.

.trastructural Changes in the Sperm Head during ·rtilization in the Rabbit

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ABSTRACT Spermatozoa have been observed with the electron microscope at various stages of their approach to and penetration of the rabbit ovum. No significant change in fine structure is observed in uterine sperm, or in many of the sperm at the periphery of the granulosa investment around the ovum. By contrast, a majority of sperm lying between the granulosa cells or on the surface of the zona pellucida display various stages of the "acrosome reaction"; this involves fusion and vesicle formation between the plasma membrane and the outer membrane of the acrosome. Loss of these vesicular elements, and content of the acrosome cap, takes place before sperm begin to penetrate the substance of the zona. The constricted posterior "equatorial" segment of the acrosome cap does not take part in the acrosome reaction and remains with its content intact during penetration of the zona; neither does the content of the apical sub-acrosomal region (perforatorium) or post-acrosomal region appear to change in traversing the zona. The hypothetical zona lysin is thus presumed to be closely associated with the persistent inner membrane of the acrosome, which now becomes the limiting membrane around the anterior part of the sperm nucleus.

No "penetration filament" has been observed, but sperm within the zona pellucida of ageing eggs are often preceded by a straight or curved fissure in the substance of the zona. The possible nature of capacitation and its relation to the acrosome reaction and to the process of penetration, are discussed briefly.

The ultra-structural features of mam-alian spermatozoa have by now been irly well documented (Fawcett, '58, '65; wcett and Ito, '65; Blom and Birch-An-rsen, '65; Nicander and Bane, '62, '66; acke and Almquist, '64; Hadek, '63b; edford, '64, '67b; Kojima, '66). Although ecies differences are seen in the form nd proportions of certain organelles, the erm head is always constructed accord-ng to the following basic arrangement. he dense nucleus is covered rostrally by e acrosome cap, a membrane limited sac lled with a fairly homogeneous material hich is presumed to possess hyaluroni-ase activity, though this has not as yet een demonstrated unequivocally. Between he inner membrane of the acrosome and he frontal border of the nucleus, except n man (Bedford, '67b), is found the perforatorium" or "apical body," which is, n fact, a subacrosomal space of variable ize occupied in most species by electron ense material. This material may possibly xtend posteriorly as a thin layer over the hole subacrosomal surface of the nucleus, to become confluent with material enveloping the post-acrosomal region (Bedford, '67b). The sperm head is covered by a plasma membrane which is always closely applied to the post-acrosomal region, but appears rather more loosely apposed to the surface of the acrosome.

Although we have a fair understanding of the structural arrangement of the mammalian sperm head, our present conception of the events which occur in sperm during their approach to and penetration of the egg is scanty at best; little is known, moreover, of the precise function or fate of the sperm organelles during fertilization. The substance of the acrosome cap has been shown to be absent in rodent and rabbit sperm penetrating the zona pellucida (Austin and Bishop, '58b; Moricard, '61; Austin, '63; Hadek, '63a; Piko and Tyler, '64), and this has sometimes left an impression that structural changes in the acrosome cap occur during capacita-

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tion. Rabbit uterine sperm, however, do not show obvious structural modifications which can be equated with the physiological change of capacitation when examined with the light, or the electron microscope (Adams and Chang, '62; Bedford, '63a, '64; Austin, '63), though evidence from *in vitro* fertilization experiments shows these sperm to be functionally capacitated. Furthermore, the fact that some component of seminal plasma can reversibly inhibit the capacitated state (Chang, '57; Bedford and Chang, '62), seems to militate against the idea that profound structural changes occur in sperm as a feature of capacitation per se.

It has not proved possible to visualize clearly the acrosome cap of living rabbit spermatozoa, but, in phase contrast microscope observation of guinea pig and rodent sperm moving within the cumulus oophorus, the acrosome cap often appears elevated or separated from the sperm head before it reaches the surface of the zona pellucida (Austin and Bishop, '58b; Ohnuki, '59; Yanagimachi, '66). In the present investigation, the question of the significance and fate of the rabbit acrosome during the approach to and penetration of the ovum has been studied with the electron microscope. The results show that with the exception of the narrow posterior segment, the substance and outer membrane of the acrosome are lost before commencement of penetration into the zona pellucida. Breakdown of the acrosome in the vicinity of the egg apparently occurs as a result of multiple fusion between the plasma membrane and outer membrane of the acrosome cap; this latter process has been discussed recently by Barros, Bedford, Franklin and Austin ('67).

MATERIALS AND METHODS

Relatively few spermatozoa are present in the ampulla of the oviduct following natural mating. For this reason, in the present study 2.0–2.5 × 10^5 rabbit sperm obtained from the cauda epididymidis were inseminated in 0.025 ml of Ringer solution into each oviduct of oestrous rabbits, two hours before administration of an ovulation injection of 50 i.u. human chorionic gonadotrophin. The timing of these events was arranged to allow ample opportunity for the sperm to become fully capacitated in the Fallopian tube, before arrival of the ova (Adams and C… '62). Ova were flushed from the F… tubes with 4% glutaraldehyde in … phate or collidine buffer. In the … of observations, the ova were recov… about 12.5 hours after the ovulation injection — at this time the bulk of the … lus oophorus had dispersed, but most ova were completely covered by cells of the corona radiata together with variable numbers of cumulus cells. The eggs … the greatest mass of surrounding granulosa cells were later chosen for sectioning. In the second series, eggs were recovered 17–18 hours after an ovulation inj… the majority of these were completely … nuded of corona cells, and showed a … central area in the ooplasm, typical of … pronuclear stage.

Ova were allowed to fix in phosphate … collidine buffered 4% glutaraldehyde … two hours, and after immersion for … eral hours in either phosphate or coll… buffer alone, were transferred to … 1% osmic acid for one hour. Soon … exposure to osmic acid, the ova becom… brown and are more easily seen … The eggs were dehydrated in ascendin… concentrations of alcohol up to 100% … cohol, during one hour, followed by … mersion in propylene oxide for two … Great care was taken not to disturb … eggs during dehydration, as the surroun… ing granulosa cells tend to become d… tached rather easily at this stage. After … hydration, the eggs were placed in … capsules containing Epon, and were … bated for several days at 60°C; it was … essary to push the eggs to the bottom … the capsule with a fine wire, as they … ally would not sink by way of gravity … the viscous resin. Sections were cut w… an automatic microtome (Porter-Bl… Sorvall) using a diamond knife. T… were stained with uranyl acetate and … citrate, and were examined in a Phil… 200 electron microscope.

OBSERVATIONS

Many spermatozoa were observed … ious stages of their approach to, … penetration of the zona pellucida. Rath…

-xpectedly, sperm were seen occasion-
y also within corona cells around sev-
l ova. The examples shown in figures 4
d 5 both have lost the content and outer
mbrane of the acrosome, but other in-
nces were observed where intact sperm
ads had come to lie within the corona
l.

The pattern of change in morphology of
e sperm head, in relation to its spatial
sition, was never clear-cut, but the fol-
wing general impression and sequence
events nevertheless made itself evident.
ost spermatozoa situated at the periphery
the granulosa cell coat showed no
ange in the continuity of the plasma
embrane, or in the form and consistency
the acrosome cap; thus their structure
d not differ essentially from that of the
ajority of mature epididymal or ejacu-
ted rabbit sperm (Hadek, '63b; Bedford,
). However, a minority of free sperm
the periphery did exhibit various de-
ees of acrosomal change. In some of
ese, the plasma and outer membranes
d content of the acrosome had disap-
ared (see fig. 3); in others, the plasma
embrane had apparently fused at various
oints with the outer acrosome membrane,
form a chain of vesicles arrayed around
e rostral half of the nucleus (see figs.
2).

In contrast to sperm present at the pe-
phery of the cells about the egg, a ma-
rity of the sperm seen within the granu-
sa mass, wedged in the interstices of the
ells, showed a variety of stages of the
rocess of acrosomal breakdown (acro-
ome reaction). Some of these sperm had
eached the point of vesiculation (figs.
, 2) with concomitant release of the con-
ents of the acrosome. In others the proc-
ss was complete, leaving only the sharply
utlined inner membrane of the acrosome
s a shroud for the anterior half of the
ucleus (fig. 3). Unchanged sperm heads
ere present on occasion between the co-
ona cells, and on one egg covered only
parsely by such cells, two intact sperm
were adherent to the surface of the zona
pellucida (figs. 7, 8). However, most
sperm among the corona cell processes
near to the surface of the zona in young
fertilizable eggs (fig. 1), showed some de-
gree of acrosomal change.

As reported by Barros et al. ('67), sperm displaying vesiculation of the plasma and outer acrosomal membranes are often to be found adhering to the zona surface (fig. 2). By contrast, sperm entering (fig. 6) or within the zona (figs. 9, 10, 11) do not retain any acrosomal material, with the exception of that present in the stable posterior (equatorial) segment (fig. 12), which remains intact during sperm passage as far as the peri-vitelline space (fig. 16). Observation of more than 100 sperm at different stages of penetration of the zona, in several ova, indicates that all remnants of the outer acrosome/plasma membrane vesicular complex are lost before penetration begins (fig. 6). Thus it is the rostral part of the inner membrane of the acrosome which must first interact with the substance of the zona pellucida during the initial phase of penetration, and it is this relatively stable membrane which, after the "acrosome reaction," becomes the limiting cell membrane over the anterior surface of the sperm head. At this stage the plasma membrane still remains intact over the posterior part of the head and over the tail (fig. 14), and thus breakdown of the original plasma membrane is confined to that region which covers the acrosome. Following the acrosome reaction, continuity of the cell surface is ensured by fusion of the equatorial segment with the remaining plasma membrane; this arrangement is shown in a sperm traversing the zona pellucida in figure 12, and can be seen in a diagrammatic representation of the acrosome reaction in text-figure 1.

The absence of any change in density or quality of the zona beneath the point of contact with the plasma membrane of the intact sperm head (figs. 7, 8), contrasts with the obvious digestion which precedes the bared inner acrosome membrane of sperm beginning to penetrate the zona. The examples shown in figures 6 and 13, might suggest that the sperm traverses the zona only through a narrow fissure; in many freshly ovulated eggs, however, the sperm begins to enter within a wide cone of superficial digestion which narrows to become a slit in the deeper regions of the zona.

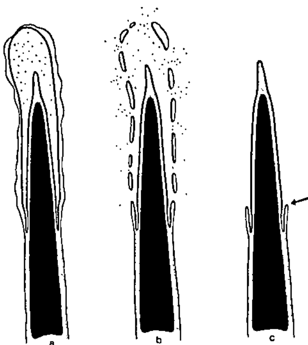


Fig. 1 Diagrammatic representation of the acrosome reaction in rabbit spermatozoa. (a) Intact sperm head as seen in samples of ejaculate, epididymal and capacitated uterine spermatozoa. (b) The outer membrane of the acrosome cap fuses at several points, progressively, with the overlying plasma membrane, to form a series of vesicles around the rostral part of the sperm head — the acrosomal content escapes through the ports which appear between the vesiculated membranes. (c) The vesicles are shed, leaving the sharply defined inner acrosome membrane as the limiting border of the anterior sperm head; continuity of the sperm surface is ensured by fusion at the anterior border of the persistent "equatorial" segment between the acrosome and plasma membranes which remain (arrowed).

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Once the sperm has undergone the acrosome reaction and has lost the outer vesiculated elements, no further structural changes occur in traversing the zona pellucida. Perivitelline sperm appear no different, structurally, from those about to penetrate (cf. figs. 3, 16), and no indication was obtained that the material which occupies the sub-acrosomal space (perforatorium) or the post-acrosomal ... in any way dissipated during ... through the zona.

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DISCUSSION

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It is difficult at present to ascribe any function to the substance of the protruding apical sub-acrosomal space or "perforatorium," during penetration of the zona (see Dan, '67; Colwin and Colwin, '67); more information is needed in mammals about the mode of fusion of the fertilizing sperm with the vitelline membrane, before a firm idea of the significance of this apical region can be gained. There are marked species differences in the form and size of the apical space, which varies from the prominent hook in rodent sperm (Clermont, Einberg, Leblond and Wagner, '55; Austin and Bishop, '58a), to the negligible protuberance in sperm from the cat and horse (Nicander and Bane, '66); the perforatorium as a special structure or rostral concentration of sub-acrosomal material is entirely absent in ejaculated human spermatozoa (Bedford, '67b). Evidence has been obtained that the cytoplasmic material of the "perforatorium" may extend posteriorly as a thin layer over the sub-acrosomal surface of the nucleus to join that present in the post-acrosomal space (Hadek, '63b; Bedford, '67b). These facts raise a question as to whether the localized apical concentration of this sub-acrosomal material, as such, has special functional significance. The material which overlies the post-acrosomal and other regions of the nuclear surface may be equally important in possible roles involving activation of the vitelline surface or of ooplasmic organelles.

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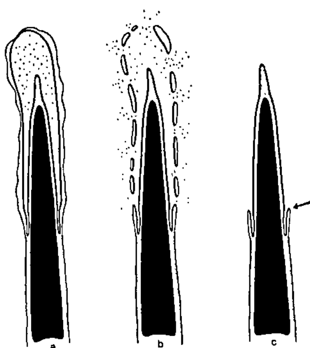


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DISCUSSION

The form and rostral position of the acrosome cap of mammalian sperm suggest an important role for this organelle in fertilization, but its function has remained uncertain. It is known that the acrosome cap is no longer present in sperm which have penetrated the mammalian egg (Austin and Bishop, '58b; Hadek, '63; Austin, '63; Piko and Tyler, '64), but the extent of its involvement at the time of passage through the zona pellucida has been unclear. Present observations reveal that the majority of the content of the rabbit acrosome is lost *before* penetration

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among the granulosa cells in the vicinity of the fresh egg in which the outer elements of the acrosome have disappeared (fig. 3) or are undergoing vesiculation (figs. 1, 2). This raises the possibility that substances in the vicinity of the mammal egg stimulate or initiate the acrosome reaction (text-fig. 1), and that viable rabbit sperm are able to react in this manner only after the prior conditioning which constitutes capacitation. Such sperm-stimulating substance(s), obviously comparable to the "fertilisin" of sea urchin eggs (Lillie, '19), could emanate from the surrounding granulosa cells and/or the ovum. The cells of the corona radiata around ovulated eggs often display a well-preserved and extensive ribosome-studded endoplasmic reticulum as well as a widespread system of Golgi lamellae. Clearly, these cells may still discharge some secretory function after ovulation. It should be noted, however, that some fresh rabbit ova remain fertilizable, though less so, after loss of the cells of the corona radiata (Chang and Bedford, '62; Bedford and Chang, '62); thus their presence is not essential for fertilization.

The exact significance of capacitation at fertilization is not wholly clear, but it is evident that sperm penetration through the zona pellucida is able to occur only after capacitation has been accomplished. Apparently, this functional change in the sperm also ensures establishment of initial contact between the gametes early in the fertile life of the egg, probably by facilitating sperm passage across the granulosa cell "barrier" (Austin, '60; Bedford, '67a). The mode of the acrosome reaction, discussed by Barros et al. ('67), and in the present paper (text-fig. 1), make it seem likely that capacitation involves changes of a destabilizing nature in the plasma membrane of the sperm. This membrane, though morphologically unchanged following capacitation, would then be able to react appropriately to the hypothetical stimulatory substances arising from the granulosa cell/egg mass, by local breakdown and fusion with the underlying outer membrane of the acrosome. This vesiculation reaction would in turn allow coordinated release in the vicinity of the egg, of the acrosomal content, which, by virtue of its presumed hyaluronidase activity must act to dissipate the matrix between the granulosa cells (Barros et al., '67). In the second place, loss of the outer acrosomal elements involved in the vesiculation reaction serves to reveal the inner membrane of the acrosome; this latter phenomenon appears an essential pre-requisite for penetration of the zona substance (cf. figs. 6, 9, 10, 11). In the above scheme of events the decapacitation factor in seminal plasma (Chang, '57; Bedford and Chang, '62a) might be presumed to inhibit the fertilizing ability of capacitated sperm by acting to block membrane receptors or to "stabilize" the plasma membrane in some other way, so as to prevent the onset of the vesiculation reaction. These last speculative suggestions are now open to experimental investigation.

In the present study, a few intact sperm were situated between the granulosa cells but it must be remembered that abnormally large numbers of sperm were deposited at the site of fertilization. The intact sperm shown in figures 4 and 5 were adherent over relatively denuded parts of the zona. Since the sperm were taken from the cauda epididymidis it is possible that these may still have been relatively immature, but this finding, nevertheless raises the question as to whether all viable spermatozoa in the ejaculate are capable of undergoing capacitation.

The plasma membrane of mammalian sperm, though physically continuous, appears to be regionally differentiated in its character, as judged by differences in the antigenic nature of the surface over the head and tail, respectively (Henle, Henle and Chambers, '38; Smith, '49) by the auto-agglutination behavior in various media, (Kato, '36; Bedford, '65), and by the typical tail-anode orientation of mature mammalian sperm in an electric field (Nevo, Michaeli and Schindler, '61; Bangham, '61; Bedford, '63). It is interesting, therefore, that in capacitated sperm approaching the ovum, the plasma membrane breaks down only locally, where it overlies the acrosome. The cell surface about the posterior part of the head and the tail (fig. 14) is apparently untouched by the factors which promote the coordinated

face breakdown seen during the acro- reaction.

n a collation of available evidence con- ning the nature of the acrosome reac- , Colwin and Colwin ('67) bring to the basic morphological similarity of event in widely separated groups in- the Enteropneusta, Echinoder- Mollusca, and speculatively, in mmalia. Present evidence shows their culation with regard to mammals to be thus the inner acrosome mem- of the mammalian sperm can be dered as the homologue of the mem- ne which delineates the acrosomal fila- or tubule, extruded during the acro- ne reaction in Arbacia or Saccoglossus. ere is, however, no extension of this mbrane as a concomitant of the acro- me reaction in rabbit sperm. Phase con- st microscope studies of fertilized ova m the rabbit, pig and sheep, have shown at appears to be a filament extending the rostral border of sperm within e zona pellucida (Dickmann, '64; Dick- ann and Dziuk, '64; Dziuk and Dick- ann, '65). In the phase contrast micro- ope this element may precede the sperm a light zone (fig. 15 — inset) or as a rk filamentous projection, depending on the exact level of focus (personal servation). No filament has been ob- rved in the electron microscope in rabbit erm at various stages of penetration rough the zona, (figs. 6, 9, 10, 11, 15) nd since the apical border always re- ained intact in these sperm, it is diffi- ult to visualize how such a filament ight arise. In contrast to the concept of filament, it seems more probable that he image of an anterior projection in the hase microscope is produced by the nar- ow fissure (fig. 15), which appears com- monly before sperm within the zona of rabbit eggs recovered at the end of their ertile life. Since this type of channel was rarely seen to precede sperm in the zona of young fertile eggs, it is possible that its appearance is associated with some change in the physical properties of the zona, as a concomitant of the ageing process.

The mode of formation of the fissure extending in front of the sperm head, is an open question. It is difficult to imagine how a process of predigestion could produce such a directional effect in the seemingly homogeneous material of the zona. Although it is conceivable that such a gap might be left after retrograde movement of the sperm within the zona, this suggestion is weakened by the fact that in some cases the fissure is curved to a much greater degree than is the rigid straight sperm nucleus. The remaining possibility that comes to mind is that this fissure develops as a physical split, or rupture of the zona substance brought about by forward movement of the sperm at a time when the ageing zona is becoming less resilient. In this case one would assume that sperm in this situation have reached the limit of their penetration and are probably destined to remain within the zona.

The finding of sperm within several corona cells was unexpected, and to the author's knowledge this has not been observed heretofore. It should be remembered, however, that relatively a large number of sperm were placed at the site of fertilization, and one cannot be certain that movement of some sperm into corona cells occurs after natural mating. Although both examples in plate 3 show sperm which have already undergone the acrosome reaction, several structurally intact sperm have been observed within corona cells. This implies that the entry process was probably phagocytic rather than the result of active penetration by the sperm. The reason for the withdrawal and incarceration of some sperm by the cells which surround the ovum, is not readily apparent, though this might conceivably serve to shield the female gamete from "defective" spermatozoa.

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1
2
av
Z

PLATE 1

EXPLANATION OF FIGURES

1 A rabbit sperm is approaching the zona pellucida (Z) between corona cells (C) and corona cell processes (P). The acrosome cap has been transformed into a series of vesicles which still cover the rostral region of the sperm head. × 13,000. Inset: higher magnification to show the arrangement of the vesiculated acrosome. × 27,000.

2 Transverse section of a rabbit sperm head on the surface of the zona pellucida of a fertilized ovum, recovered approximately two and one-half hours after ovulation. The outer investment of the acrosome has undergone vesiculation (av), with loss of the acrosomal content. No change is evident in the zona substance (Z) in contact with the vesiculating acrosomal elements. × 23,600.


iam
e
3

PLATE 2

EXPLANATION OF FIGURES

3 Rabbit sperm between corona cells (C) approaching the surface of the zona pellucida, which lies out of view beyond the upper border of the plate. In this case the vesiculated acrosomal investment has been discharged before contact with the zona pellucida, exposing the inner membrane (i.a.m.) of the acrosome cap. Note the intact posterior remnant of the cap (c) which persists during penetration of the zona. × 40,000.


iam
3
e

PLATE 3

EXPLANATION OF FIGURES

4–5 Both figures show rabbit sperm nuclei lying within corona cells. Examples have been observed of intact sperm similarly placed within corona cells. Note the distribution of Golgi lamellae and ribosome-studded endoplasmic reticulum within the cells. × 15,800.

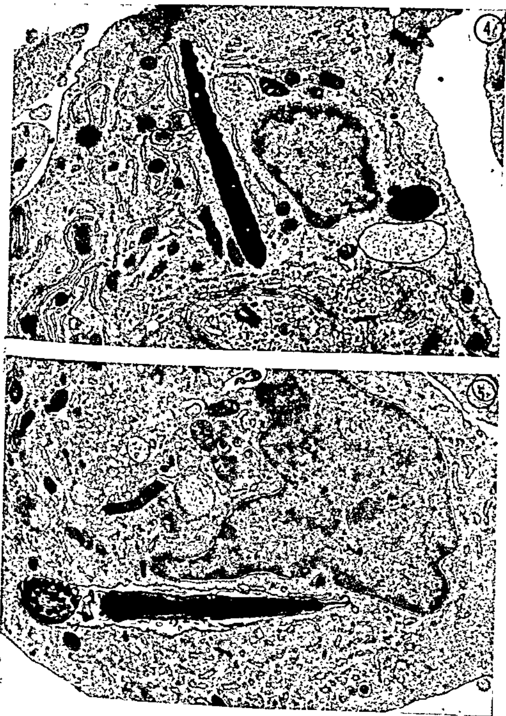

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5

PLATE 4

EXPLANATION OF FIGURE

6 Rabbit sperm head beginning to penetrate the zona pellucida. No acrosomal vesicles remain about the rostral surface of the head; thus, the zona interacts solely with the inner membrane of the acrosome cap during the initial stages of penetration. This ovum has already been activated by another sperm as evidenced by the absence of cortical granules in the vitellus. × 17,000.


6
Z

PLATE 5

EXPLANATION OF FIGURES

7 An intact sperm head is adhering to a relatively denuded part of the zona surface (Z), though a corona cell (C) is seen immediately above the sperm head. The zona pellucida appears in no way affected by contact with the sperm plasma membrane (arrowed). × 38,000.

8 Sperm head with intact acrosome cap, and most of the covering plasma membrane (arrowed), adhering to a denuded region of the zona pellucida (Z) of a fertilized ovum recovered about two and one-half hours after ovulation. Although the sperm head has indented the surface, no change is apparent in the substance of the zona. × 38,000.

7

P

Z

8

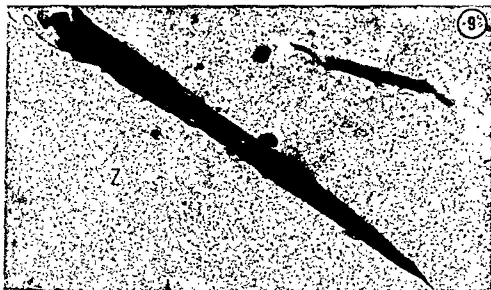
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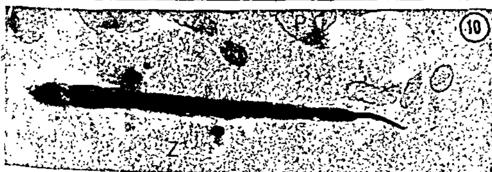
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PLATE 6

EXPLANATION OF FIGURES

9–11 Examples of sperm lodged wholly within the zona pellucida of freshly ovulated ova. No vesiculated acrosomal elements remain about the sperm head. All figures × 22,000.


9
Z

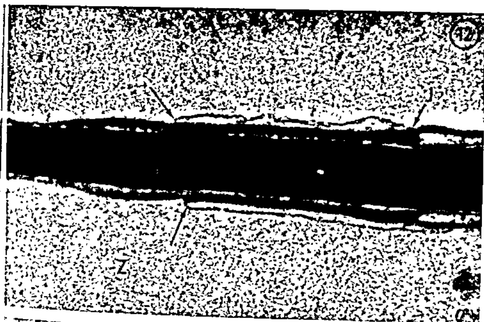

10
Z


11
Z

PLATE 6

EXPLANATION OF FIGURES

9–11 Examples of sperm lodged wholly within the zona pellucida of freshly ovulated ova. No vesiculated acrosomal elements remain about the sperm head. All figures × 22,000.


12
Z

13
PC
P
Z
V

PLATE 7

EXPLANATION OF FIGURES

12 Central portion of a sperm head lying within the zona of a freshly ovulated ovum. Arrows denote the point of fusion between the outer membrane of the acrosome cap and the overlying plasma membrane which extends caudally to cover the posterior head and tail surface; J. indicates the posterior limit of the acrosome cap. × 70,000.

13 Section of a fertilized ovum recovered approximately six hours after ovulation. This shows the typical form of the penetration curve (p.c.) formed by sperm in traversing the zona pellucida (Z). A sperm head is seen in the peri-vitelline space at the right of the picture, and a remnant of a corona cell process (p) is still present within the substance of the zona. × 11,800.

14
ZS
15
16
Z
V



PLATE 8

EXPLANATION OF FIGURES

14 Section of sperm tail below the zona surface (ZS). At this stage the plasma membrane remains intact over the posterior head and the tail region of the sperm. × 56,000.

15 Sperm head lodged within the zona pellucida of a fertilized ovum recovered approximately seven hours after ovulation. The fissure which precedes the head, is seen commonly in eggs taken at this stage, and is thought to be responsible for the appearance of a projection (see inset) which sometimes extends from the rostral border of such sperm viewed with the phase microscope. × 25,500.

16 Sperm head lying in the peri-vitelline space of a fertilized egg. Note that the space of the perforatorium is occupied by electron-dense material. The posterior equatorial segment of the acrosome also remains intact in peri-vitelline sperm. × 25,500.


18



PLATE 9

EXPLANATION OF FIGURES

17 Section of the fertilizing sperm head in an ovum recovered about two hours after ovulation. The swelling envelope around the dense nucleus may be nuclear membrane. × 23,500.

18–19 Higher magnification of sections of a fertilizing sperm head. Note the appearance of areas of rarefaction within the dense nucleus (arrow), and fibrils (NF), emanating from the surface of the dense nuclear chromatin. Figure 18, × 64,000; figure 19, × 69,000.

17
Z
pvs





PLATE 10

EXPLANATION OF FIGURES

20 Tail of the fertilizing sperm (T) entering the vitellus. Note the absence of a limiting membrane around the tail filaments within the ooplasm (cf. fig. 14). It is uncertain whether the membranes within the peri-vitelline space derive from the vitelline surface, from the limiting membrane of the sperm, or from both. × 51,000.

21 Appearance of the male pronucleus in an egg recovered about six hours after ovulation. Note the homogeneous nucleoplasm, and the prominent nucleoli, which are not membrane bound. At this stage the pronuclei become surrounded by many newly formed Golgi vesicles and lamellae. × 8,000.

22 Higher power of part of a male pronucleus (PN); the sperm tail (T) often lies in close apposition to the pronucleus at this stage. × 25,000.


20
T


21

·utoradiographic Investigation of ·alvarial Growth in the Rat [1]

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ABSTRACT Results of autoradiographic investigations utilizing tritiated proline reveal an intricate mechanism of rat calvarial growth and reshaping from age two days through 75 days. Generalized intramembranous bone growth dominates the growth process from two through eight days; however, this simple bony enlargement is terminated early. At eight days a differential apposition-resorption process begins dorsally in the calvarium and progresses rostrally within a ten day period. This process includes differential apposition on the ectocranial and endocranial periosteal surfaces accompanied by a differential apposition-resorption pattern on the endosteal surfaces. The wave-like process begins in the occipital bone and progresses ventrally to the frontal bone resulting in a flattening of the bony components. At 35 days bone accretion is again generalized on most calvarial surfacse. However, at 40 days another change in the growth process evolves on the occipital and frontal bones. These bones are now seemingly displaced in a superior direction as bone apposition continues on their superior surfaces, namely, the ectocranial periosteal and endocranial endosteal surfaces and resorption progresses on the two inferior surfaces. The magnitude and duration of the process described in this paper is sufficient to account for calvarial flattening in the absence of bony spatial reorientation due to bending at sutures. The findings described here when compared to proposed in vitro studies can help put the importance of the so-called "functional matrix" related to bone growth into its proper perspective.

The specific sites of bone apposition and ·esorption in the growing rat calvarium ·ave not been clearly defined. The role ·layed by the areas of apposition and re·orption in the general growth and reshap·ng of the calvarium is subject to con·troversy (Baer, '54; Brash, '34; Brodie, ·41; Craven, '56; Giblin and Alley, '42, ·44; Massler and Schour, '51; Moss, '54; Young, '62).

Moreover, assessment of the influence of environmental factors such as soft tis·sue function upon bone growth remains a difficult task. Genetic control over the primordia of bones is generally accepted. However, the point at which environment assumes primary importance, if there be such a point, remains unknown. Function, according to its latest proponent, Moss (Moss and Baer, '56), is of prime importance in bone growth. Consequently, enlargement of the brain, periosteal sliding and other soft tissue systems are considered by him to be determinants of growth of the neurocranium. This hypothesis may well be true; however, definite concepts cannot be postulated without knowledge of the precise mechanism of bony changes and without experimental evidence that a specific pattern of bone growth would be altered in the absence of soft tissue or functional requirements.

The autoradiography technique (Leblond et al., '50) with tritiated proline was utilized in the experiments described in this paper in order to assess small increments of growth on decalcified sections, a task beyond the resolution afforded by previous Alizarin Red S studies (Cameron, '30; Jarabak, '51; Moore, '49; Schour, '36, '41; Massler and Schour, '51; Baer, '54). The experiments were designed to observe the growth process from birth to seventy-five days in the male albino rat calvarium. In order to prepare for the major study, two preliminary investigations were performed.

The first preliminary experiment was designed to determine the precise pattern and the magnitude of differential apposi-

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[2] A post-doctoral Fellow at Forsyth Dental Center and Harvard University School of Dental Medicine and supported by the National Institute of Dental Research at the time this work was done.

TABLE 3

Growth of male albino rat calvarium. Experimental design. Major investigation

Number of animals at each age	Age in days at first injection.[1] These animals were injected a second time after two days and sacrificed *six hours later*	Age in days at first injection.[1] These animals were injected a second time after four days and sacrificed *six hours later*	Age in days at first injection.[1] These animals were injected a second time after six days and sacrificed *six hours later*
2	2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, *32, 34, 36, 38, 40, 42, 44*	46, 50, 54	58, 64, 70

[1] One microcurie of tritiated proline per gram of body weight.

t four days of age and sacrificed six hours later. Two four-day-olds were injected with comparable amounts of proline and again injected at six days of age; then sacrificed six hours later. This injection scheme was continued through animals of 46 days of age. Two 46-day-old animals were injected and again injected at 50 days, then sacrificed six hours later. This four-day injection interval was now used for animals up to 58 days and a six-day period up to 75 days (table 3).

After sacrifice with ether, the tibia and calvarium were removed and fixed in 5% formol-saline, decalcified with formic-acid-sodium-citrate, embedded in paraffin and cut sagitally at 6 µ. The sections were dipped in Kodak NTB 2 emulsion and exposed for 11 to 14 day intervals. They were then processed and post-stained with hematoxylin and eosin.

RESULTS

Sutures. From two through 20 days of age the sutural connective tissue at the parietal-frontal, anterior lambdoidal and posterior lambdoidal sutures diminished in width as generalized intramembranous growth of adjacent bones continued through 18 days (Gans and Sarnat, '51; Pritchard, Scott and Girgis, '56).

At 20 days the sutural growth became more appositional in nature as sheets of osteoblasts lined the bone edge on either side of the suture. The three sutures mentioned remained active through the 75-day experimental period in contrast to the earlier closure if these sutures proposed by Massler and Schour, '51 (figs. 2, 3).

Ectocranial and endocranial surfaces (fig. 1). Apposition on all surfaces of the

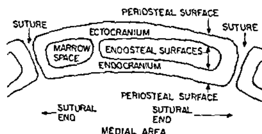


Fig. 1 Diagrammatic representation of calvarial bone with surfaces to be discussed labeled.

frontal, parietal, interparietal and occipital bones through eight days was observed, with no differential pattern exhibited during this period (figs. 4, 5).

At eight days, however, a differential pattern of apposition became evident. The ectocranial periosteal surface of the interparietal bone exhibited marked and rapid growth at the suture ends with little or no change at the medial area of this surface (figs. 6, 7). The endosteal side of this surface, however, was an active site of apposition. The endocranial periosteal surface exhibited more growth at the medial and less at the suture borders. The endosteal side of this surface was an active resorption site. At no time was resorption observed on the endocranial periosteal surface. While the differential pattern continued on the interparietal bone, the generalized apposition on the parietal and frontal bone continued (fig. 8).

At 12 days the parietal bone began the differential growth pattern described for the interparietal bone while the frontal bone still exhibited a generalized apposition and remodelling.

TABLE 1

Preliminary experiment 1
Experimental design

Number of animals injected [1] in each group	Age of animal when given first injection	Age of animal when given second injection and sacrificed six hours later	Age of animal when given second injection and sacrificed seven days later
2	21	28	
2	28	35	
2	35	42	
2	42	49	
2	21		28
2	28		35

[1] One microcurie of tritiated proline per gram of bodyweight.

tion and resorption in the neurocranium as well as the duration of growth at the neurocranial sutures.

The second preliminary experiment was designed to determine an injection regimen leading to the major study of calvarial growth from birth through 75 days. Its purpose was to assess the optimal interval between injections of ^{3}H-proline in representative age groups during the experimental period so that two injections would be far enough apart for good definition and at the same time avoid the loss of the first injection due to remodelling.

METHODS AND MATERIALS

Preliminary experiments

Preliminary experiment 1. Twelve Holtzman male albino rats ranging in age from 21 through 49 days were studied. Two 21-day-old animals were injected with one microcurie of ^{3}H-proline per gram of body weight. These two animals were again injected at 28 days and sacrificed six hours later. This same scheme was followed over the next two seven-day periods. Animals with two injections seven days apart were sacrificed after 14 days and also studied (table 1).

After sacrifice with ether, the tibia and calvarium were removed, fixed in 5% formol-saline, decalcified with formic acid sodium citrate, embedded in paraffin and cut sagitally at 6 μ. These sections were dipped in Kodak NTB 2 emulsion and exposed for five through 14-day intervals. They were then processed and post-stained with hematoxylin and eosin.

Preliminary experiment 2. Sixteen male albino rats of the Holtzman strain were studied. Four animals were injected with 1 μCi of ^{3}H-proline per gram of body weight at 2, 21, 35 and 55 days of age. Two of each group were injected a second time two days later and sacrificed after six hours. The other two in each group were injected once more after four days and sacrificed six hours later (table 2).

The tibia and calvarium were removed from each and treated as in preliminary experiment 1.

RESULTS

The results of the preliminary experiments revealed that a two-day interval between injections was most suitable in animals up to 46 days, four-day intervals from 46 through 58 days, and six-day intervals from 58 through 75 days. The magnitude of the major remodeling process, based on findings of preliminary experiment 1, will be discussed later. The major experiment was then designed, utilizing knowledge from these two pilot studies, to observe the growth process from birth through 75 days.

METHODS AND MATERIALS

Major investigation

Fifty-six Holtzman male albino rats ranging in age from two to 75 days were studied. Following the findings of preliminary experiment 2, two 2-day-old animals were injected intraperitoneally with 1 μCi of tritiated-proline per gram of body weight. These animals were injected again

TABLE 2

Preliminary experiment 2
Experimental design

Number of animals injected [1] in each group	Age in days when first injected. These animals were injected a second time two days later then sacrificed after six hours	Age in days when first injected These animals were injected a second time four days later then sacrificed after six hours
2	2, 21, 35, 55	2, 21, 35, 55

[1] One microcurie of tritiated proline per gram of bodyweight.

arily a reaction to environmental stimuli, ch as soft tissue function. Only by organ ulture means can the validity of such a oncept be tested. While it is true that ork by Fell and Robinson ('29) has own a strong genetic effect in growth long bones, the changes we have de- ribed above in the cranial bones are so uch more intricate and time dependent, at these bones lend themselves ideally to rther investigations of this kind.

ACKNOWLEDGMENTS

The author is indebted to Drs. James T. ying, Coenraad F. A. Moorrees and Miles Crenshaw for their guidance in this udy and to Mr. John D. Heeley for his chnical assistance.

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At 14 days the same pattern of change occurred in the frontal bone. As differential apposition continued on the periosteal surfaces the marrow spaces exhibited the same interesting pattern of change. The ectocranial endosteal surface was lined with osteoblasts laying down bone while the endocranial endosteal surface was actively resorbing bone as evidence by many Howship's lacunae filled with osteoclasts, and no apposition occurred at this surface. This intricate pattern continued on the occipital, interparietal, parietal and frontal bones until 35 days.

At 35 days, the differential pattern of apposition and resorption was lost and a more generalized system of growth was exhibited on all periosteal surfaces (fig. 9).

At approximately 40 days, the frontal and occipital bones showed a different pattern than that previously descrived. Apposition now was evident on the entire ectocranial periosteal surface and the entire endocranial endosteal surface while resorption occurred on the ectocranial endosteal surface and endocranial periosteal surfaces (fig. 10).

DISCUSSION

Growth of the neurocranium up to eight days was shown to be a result of generalized intramembranous bone growth which occurred within a fibrous connective tissue capsule and progresses from each bone center radially until the presumptive sutures are compressed into the adult form (Pritchard, Scott and Girgis, '56).

However, after eight days a differential pattern of apposition and resorption began within this capsule. This pattern started dorsally in the calvarium and progressed rostrally within a few days. The changes described led to a flattening of the occipital, interparietal, parietal and frontal bones. These bones flattened individually until 35 days when they apparently reached their final configuration and thereafter grew in a more generalized, uniform manner.

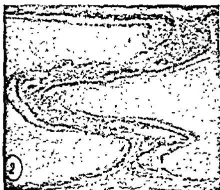
At approximately 40 days another change in pattern was noticed at the occipital and frontal bones. These bones were now displaced in a superior direction similar to the process in long bones called "drift" by Enlow ('63) (fig. 11).

In view of this finding, Enlow's concept of alteration of environment in long bones may apply to membranous bones as well. Enlow states that during lengthening of long bone by epiphyseal growth, changes along the shaft are induced by the altered environment as muscle attachments change and periosteal sliding occurs.

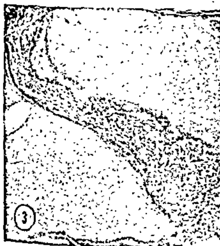
In light of my investigations, the flat bone counterpart of the epiphysis may be the suture and the outer and inner tables of the calvarial bones can be compared to the shaft of long bone. Thereby, the sutures, acting as pacemakers like the epiphyses, may induce concurrently adjustive growth changes along the length of membranous bones between them as function and environment are altered.

The magnitude and importance of the differential pattern of apposition and resorption described was made clear by preliminary experiment 1. In this experiment it was established that a ^{3}H-proline injection given 14 days prior to sacrifice was now located at the resorptive endocranial endosteal surface of the parietal bones. This surface therefore replaced itself in a 14-day period from 21 days through 35 days (fig. 12). An apposition-resorption pattern of this magnitude can easily lead to a change of shape of this bone, and the concurrent process in the other components of the calvarium could certainly result in the flattening of the calvarium with growth. This flattening process was also described by Baer ('54) but it was explained by the spatial reorientation of each bone by a bending at the sutures. The conclusions obtained from my own investigations indicate that very little reorientation is needed.

The results of the foregoing experiments indicate the specific sites of alteration of growth in the calvarial bones at different stages of development. The next question is to consider the various influences which control these alterations in growth, environmental or genetic. While there is no doubt that both these factors play a role, it is our intention to undertake organ culture experiments to ascertain if similar changes are brought about in the absence, as far as possible, of usual environmental conditions. Moss's theory, for example, is that bone growth and remodeling is pri-


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3


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8


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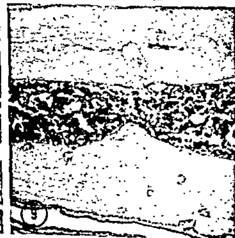

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PLATE 1

EXPLANATION OF FIGURES

2 Parietal-frontal suture. Animal injected at 70 days and 75 days, then sacrificed six hours later. Note continued growth of both bones at the suture. (× 110, H & E).

3 Anterior lambdoidal suture. Animal injected at same intervals as in figure 2. Growth takes place at this suture also. (× 110, H & E).

4 Parietal Bone — generalized intramembranous growth at age six days in control animals. Osteoblasts evident on all surfaces. Ectocranial surface (a). Endocranial surface (b). All other photomicrographs will have same orientation. (× 110, H & E).

5 Same age as figure 4. Autoradiograph shows generalized labeling of bone surfaces (× 110, H & E).

6 Interparietal bone — medial area. Animal injected at eight days and ten days, then sacrificed six hours later. Note apposition on ectocranial endosteal surface and endocranial periosteal surface. The ectocranial periosteal surface appears quiescent while the endocranial endosteal surface is rapidly resorbing as shown in figure 12. (× 110, H & E).

7 Lateral portion of same bone as in figure 6. The periosteal ectocranial surface at this site (arrow) is actively depositing bone in contrast to the medial area distant from the suture. (× 200, H & E).

8 Parietal bone — medial area, same age period as in figures 6 and 7. Bone apposition on both periosteal surfaces with no apposition at endosteal side. No differential pattern is evident here. (× 110, H & E).

9 Frontal bone — medial area. Animal injected at age 34 days and 36 days, then sacrificed six hours later. General periosteal apposition has returned. (× 110, H & E).

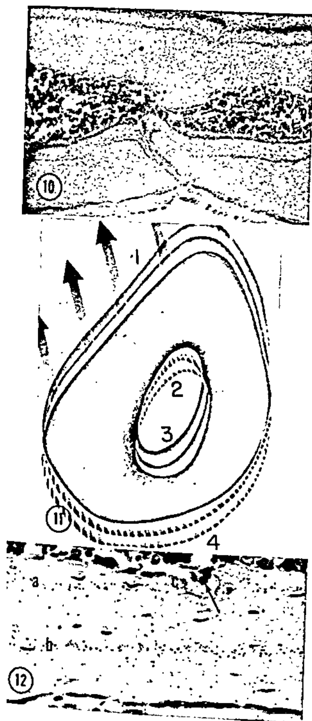

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a
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b
12

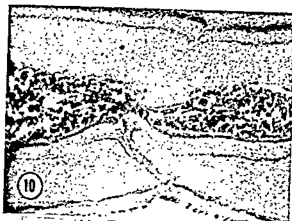
PLATE 2

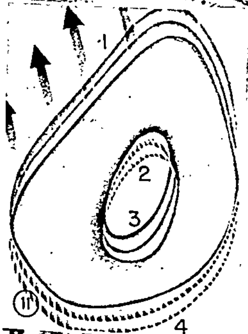
EXPLANATION OF FIGURES

10 Frontal bone — medial area. Animal injected at ages 42 days and 44 days, then sacrificed six hours later.

11 Cross section of long bone from Enlow. Drift phenomenon similar to that process described in calvarial bone of figure 10.

12 Parietal bone — medial area. Animal injected at 21 days and 28 days, then sacrificed at 35 days. 21 day injection (a), 28 day injection (b), endocranial endosteal resorbing surface (c). Osteoclast at label given 14 days prior to sacrifice (arrow). This bone surface has replaced itself in a 14 day period. (× 440, H & E).


10


1
2
3
4
11


a
b
12



ıe Fine Structure of Uterine Smooth Muscle of the ıt Uterus at Various Time Intervals Following :Single Injection of Estrogen[1]

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ABSTRACT The fine structure of smooth muscle cells of the rat uterus was studied at 6, 12, 24, 48, 72 and 96 hours after a single subcutaneous injection of estradiol dipropionate (10 µg). The control rats were ovariectomized, but did not receive hormone treatment. Changes in the ultrastructure of the muscle cells occurred soon after estrogen stimulation. An increase in the number of ribosomes on the outer nuclear membrane and evagination of the membrane into the surrounding cytoplasm was noted at six hours. The amount and dilatation of the granular endoplasmic reticulum (GER) increased gradually from 6 to 96 hours. At 24 and 48 hours the mitochondria appeared to increase in number and at later time intervals they increased in size. The Golgi complex was more prominent at 72 and 96 hours than at the earlier time intervals. An increase in free ribosomes was observed at 6 and 12 hours, but a marked accumulation of them occurred at 24 hours and continued to 96 hours. The glycogen granules were very pronounced at 24 and 48 hours with a reduction at 72 and 96 hours. Many pinocytotic vesicles and dense bodies were present in the controls and at 6 and 12 hours. A decrease in number occurred in both at the later time intervals. Organelles accumulated at the nuclear poles beginning at 24 hours and extending to 96 hours. The observations made provide additional information on the fine structure of uterine smooth muscle cells following estrogen stimulation and support previous biochemical studies on glycogen concentration and RNA synthesis in the rat uterus.

Estrogen increases the glycogen concen-ıtion of the rat uterus (Boettiger, '46). ıe polysaccharide is localized primarily smooth muscle cells (Bo and Atkinson, 2) and appears shortly after initiation of ırmone treatment (Walaas, '52; Bitman, ıcil, Mench and Wrenn, '65; Bo, Maraspin ıd Smith, '67). Similarly, glycogen syn-ıetase activity increases in the myo-ıetrium of the rat uterus soon after ıstrogen stimulation (Bo and Smith, '64; o, Maraspin and Smith, '67). Shortly after ıstrogen treatment there is also an increase ı the amount of phospholipids, RNA syn-ıesis and protein synthesis in the rat ıterus (Aizawa and Mueller, '61; Mueller, ıerranen and Jervell, '58; Noteboom and ıorski, '63).

The literature contains a few reports ıhich attempt to correlate the fine struc-ıure of uterine smooth muscle cells with ıhe chemical changes that occur following ıstrogen stimulation. Laguens ('64) re-ıorted an increase in the number and size ıf the nucleoli at 6, 12, 24 hours after a ıingle injection of estrogen (10 µg). At 48 and 72 hours the large nucleoli lay peripherally and in the cytoplasm some ribosomes, rough endoplasmic erticulum (RER) and slightly enlarged mitochondria were present. Ross and Klebanoff ('67) observed that muscle cells in the diestrous rat uterus were elongated, spindle-shaped, contained ellipsoid nuclei and cytoplasmic organelles present were located adjacent to the nucleus. In the cytoplasm the RER had only a few scattered ribosomes on its membranes, Golgi complexes were well developed, scattered free ribosomes appeared and numerous vesicles occurred at the surfaces of the cells. At estrus, Ross and Klebanoff ('67) noticed a marked increase in the amount of RER and Golgi complexes; the surface of the smooth muscle cells had numerous outpocketings and infoldings. Following estrogen stimulation for three consecutive days the fine structure of uterine smooth muscle was similar to the estrous condition (Ross and Klebanoff, '67).

[1] This investigation was supported by USPHS grants AM-08029-04 and HD-00606. The authors are grateful to Dr. Norman Sulkin for making the electron microscope available.

In order to better understand the progressive ultrastructural changes that occur in the smooth muscle cells of the rat uterus following estrogen stimulation, this investigation was designed to study the cells at different time intervals following a single subcutaneous injection of the hormone. The study will attempt to correlate, where possible, the changes with some of the chemical alterations known to occur.

MATERIAL AND METHODS

Twenty-four adult virgin female rats (weighing 200 ± 10 gm) of the Wistar strain (Charles River were used. All the animals were bilaterally ovariectomized and maintained four to a cage with free access to food. Ten days following ovariectomy four control animals were killed; the remaining rats received a single subcutaneous injection of 10 µg estradiol dipropionate [2] in 0.1 ml cottenseed oil. The animals (3 or 4 at each time interval) were killed with ether vapor at 6, 12, 24, 48, 72 and 96 hours after administering the hormone. The uterus was quickly removed, placed in a cold solution of 2% osmium tetroxide buffered with veronal acetate (pH 7.4) and cut in small pieces. The tissue was fixed for one hour, dehydrated through a graded series of alcohols and embedded in an epoxy resin, Epon 812. The sections were cut with a Portor-Blum ultramicrotome and stained for 30 minutes with lead citrate (Reynolds, '63).

RESULTS

Ovariectomized animals. In many areas a relatively large amount of intracellular connective tissue was present and the smooth muscle cells were small and spindle-shaped. Only a few elongated and centrally located nuclei appeared in the smooth muscle cells. In a few areas the outer nuclear membrane formed small evaginations into the cytoplasm and only a few ribosomes were attached to the membranes. The myofilaments were parallel to the long axis of the cell, but absent in areas containing organelles. The plasma membrane was fairly free of evaginations throughout its extent.

The few mitochondria present lay primarily close to the periphery of the cell although some of them were present at the nuclear poles. The shape of mitochondria varied, being either elongated, round, or short rod-like structures with well developed lamellar cristae. Only a very small amount of granular endoplasmic reticulum (GER) was noted along the sides of the nucleus and at the nuclear poles. Well defined Golgi complexes appeared lateral to the nucleus. Small vesicles made up the complex with only a few cisternal elements. Numerous pinocytotic vesicles were attached to or near the plasma membrane. Several multivesicular bodies lay beside the nucleus. A few ribosomes were scattered around the nucleus and along the plasma membrane. The rare glycogen granules appeared close to the plasma membrane or at the nuclear poles. Dense bodies present had a variable morphology. Some of them contained linear, closely packed structures, while others had small rounded dense structures of various sizes surrounded by a light background matrix (figs. 1, 2).

Six hours following estrogen treatment. Several changes in the structure of uterine smooth muscle cells manifested themselves at this time interval. The nucleoli appeared to be closer to the nuclear envelope than in the control rats. More ribosomes were located on the outer nuclear membrane than in the controls; numerous evaginations of the membrane projected into the cytoplasm. These evaginations became especially prominent at the nuclear poles. In addition to being present along the plasma membrane mitochondria and GER occurred at the nuclear poles. Mitochondria varied in shape as in the control group of animals. The amount of GER increased and some of it was dilated, forming branching cisternae. The Golgi complex, located at the side of the nucleus and at the nuclear poles, contained more vesicular than cisternal elements. More free ribosomes and glycogen granules located at the sides of the nucleus and at the nuclear poles existed than in the ovariectomized rats. The dense bodies varied in morphology; the most common form had an enclosing membrane with a small space between the membrane and the contents. The light matrix contained granules of variable size. A few of the

[2] The estradiol dipropionate was supplied through the courtesy of Ciba Pharmaceutical Products, Summit, New Jersey.

nse bodies had clear vacuoles resembling ose in lipofuscin pigment.

Twelve hours after estrogen treatment. ith a few exceptions the fine structure of e smooth muscle cells was similar to that six hours. The cells appeared larger, e plasma membrane protruded in some aces into the connective tissue and fewer aginations of the outer nuclear membrane were present. Glycogen granules, osomes and GER increased slightly in nount from that observed at six hours.

Twenty-four hours after estrogen treatment. The muscle cells were more elonated than observed previously due to the tension of the cytoplasm from the nuear poles. Adjacent muscle cells intergitated frequently at this time interval. wo nucleoli occurred in a few cells. Evaginations of the outer nuclear membrane ere not as pronounced as at 12 and 6 ours. At this time interval the greatest ange consisted of the accumulation of rganelles at the nuclear poles. The GER creased, its cisternae enlarged, the mitohondria appeared to have increased in umber, but the Golgi complex remained pproximately the same size as previously. marked increase in the number of free bosomes and glycogen granules located oth at the nuclear poles and along the ytoplasmic extensions was evident. There ere fewer pinocytotic vesicles than noted arlier (fig. 3).

Forty-eight hours after estrogen stimulation. The cells had increased in size and more interdigitation of adjacent muscle ells occurred than at the preceding time nterval. There were indentations of the uclear envelope, but these may have resulted from contraction of the muscle cells. The outer nuclear membrane had only a few ribosomes with only a few areas showing evaginations of the membrane into the surrounding cytoplasm. Some of the nucleoli lay closer to the nuclear envelope than in the previous groups. The amount of GER greatly increased and appeared primarily at the nuclear poles and in the cytoplasmic extensions of the cells. In some areas the GER consisted of large tubular-like structures. The Golgi complex lay beside and at the poles of the nucleus; its size did not appear to vary from the preceding time intervals. Although mitochondria were located primarily at the nuclear poles some occurred next to the nucleus and in the cytoplasmic extensions. Their number appeared to have increased at this time interval.

A marked increase in glycogen concentration occurred at this time interval. Its component particles were located close to the plasma membrane, at the nuclear poles and in the cytoplasmic extensions. Also free ribosomes, located primarily at the nuclear poles and in the cytoplasmic extensions, increased greatly. Very few pinocytotic vesicles were seen along the plasma membrane (figs. 4–6).

Seventy-two hours after estrogen stimulation. At this time interval GER, mitochondria and Golgi complexes became concentrated at the nuclear poles and in the cytoplasmic extensions of the muscle cells. There were fewer glycogen granules than at 48 hours, and they appeared primarily along the plasma membrane (fig. 7).

Ninety-six hours after estrogen stimulation. The structure of the muscle cells was similar to that seen at 72 hours. However, he GER appeared to have increased in amount and often in many areas it was dilated. In some of the cytoplasmic extensions mitochondria, GER, ribosomes and scattered glycogen granules occupied the entire part of the cell (fig. 8).

DISCUSSION

The results demonstrate that the fine structure of the smooth muscle cells of the rat uterus undergo changes soon after estrogen stimulation. At six hours, ribosomes were abundant on the outer nuclear membrane which was evaginated into the surrounding cytoplasm. Although some ribosomes appeared on the outer nuclear membrane and some evagination occurred at later time intervals they were not as pronounced as at six hours. A gradual increase in amount and dilatation of the GER occurred from 6–96 hours. Mitochondria appeared to increase in amount at 24 and 48 hours and at the later time intervals they enlarged. The Golgi complex was seen throughout the periods studied, but seemed to be more prominent at 72 and 96 hours at the nuclear poles. An increase in free ribosomes appeared at 6 and 12 hours, but a marked accumulation of them occurred

at 24 hours and continued to 96 hours. The glycogen granules were very pronounce at 24 and 48 hours followed by a reduction at 72 and 96 hours. Many pinocytotic vesicles and dense bodies were present in the controls and at 6 and 12 hours. A decrease in number of vesicles and bodies occurred at the later time intervals.

The findings of the present study at time intervals (48–96 hours) are similar to the observations made by Ross and Klebanoff ('67) in the uteri of rats during estrus and in ovariectomized rats following estrogen treatment for three days. The glycogen concentration was not discussed by the above investigators. The present data differ from those of Laguens ('64) who did not detect any cytoplasmic changes until 48 hours after a single injection of estrogen (10 μg); free ribosomes were increased in number, some rough ER appeared and the mitochondria were enlarged. No comment was made on the glycogen concentration at the different time intervals. The observations obtained in the present study on the glycogen concentration at different time intervals following estrogen stimulation are similar to the results of biochemical studies reported previously (Bo, Maraspin and Smith, '67).

Noteboom and Gorski ('63) in a biochemical study did not detect any significant increase in protein synthesis in the nuclear, mitochondrial and ribosomal fractions of uterine cells at two hours after estrogen treatment; four hours after hormone stimulation a marked increase was noted in all the above fractions. It has been suggested by Ui and Mueller ('63) that estrogen stimulates the synthesis of certain proteins in the uterus which are necessary for the synthesis of new RNA and that the new RNA supplies the protein synthesizing machinery the essential information necessary for effecting the early anabolic effects of the hormone. The cellular RNA is primarily ribosomal. Moore and Hamilton ('64) reported that ribosomes were more abundant in the cytoplasm of the cells of the entire uterus following estrogen stimulation than in those deprived of the hormone; 4 to 24 hours after estrogen treatment newly formed ribosomes appeared in the cytoplasm. The increase in the amount of free ribosomes and GER following estrogen stimulation which was noted in this investigation supports the biochemical results obtained by Moore and Hamilton ('64).

Hamilton ('63) suggested that estrogen stimulates intranuclear reactions resulting in the production or regulation of messenger RNA which precedes the acceleration of protein synthesis. If the synthesis of new RNA is blocked by actinomycin D, the early acceleration of phospholipid and protein synthesis due to estrogen do not occur (Ui and Mueller, '63). Whether the accumulation of glycogen in uterine smooth muscle following estrogen stimulation is dependent on newly synthesized RNA has not been established, but the present ultrastructural study provides suggestive evidence of a linkage of the two phenomena.

The dense bodies were more evident in the smooth muscle cells of the controls and at 6 and 12 hours than at the later time intervals following estrogen stimulation. Whether the dense bodies were lipofuscin pigments or lysosomes was not established. The significance of dense bodies in smooth muscle cells has not been determined. Pigment granules have been observed to accumulate in smooth muscle cells of vitamin E deficient intact rats (Mason and Emmel, '45). Kaunitz, Slanetz and Atkinson ('49) reported that in ovariectomized rats pigmentation did not develop to the degree characteristic of vitamin E deficiency, but following prolonged estrogen treatment during the course of avitaminosis pigmentation developed which was similar to that seen in intact vitamin E deficient rats. They may accumulate in the cell when the normal metabolic activity of the cell is altered. This has been shown to occur in skeletal muscle (Odor, Patel and Pearce, '67).

An increase in the number of mitochondria was noted at 24 hours and 48 hours after estrogen treatment and at 72 and 96 hours the organelle appeared to increase in size. The changes seemed to occur at the time of greatest metabolic activity of the smooth muscle cells. Rouiller ('60) reported that in skeletal and heart muscles there is a relationship between tissue activity and the number of mitochondria

...ce in active striated muscle (red) the ...tochondria are numerous and in white ...iated muscles there are fewer mitochon...a. Whether or not the enlargement of ...tochondria observed at 72–96 hours was ...e to change in metabolic activity was ...t determined. Swelling of mitochondria ...curs in various physiological and patho...gical states (Rouiller, '60).

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PLATE 1

EXPLANATION OF FIGURE

1 Electron micrograph of several uterine smooth muscle cells from an ovariectomized rat. The smooth muscle cells are small and the intercellular material (IC) is fairly abundant. Along the plasma membrane (PM) are pinocytotic vesicles (PV). In the cytoplasm can be seen several dense bodies (DB) of variable structure. Also present in the cells are a few mitochondria (M), a small amount of granular endoplasmic reticulum (GER) a few scattered free ribosomes (R) and the Golgi complex (GC). × 20,720.


DB
R
IC
GER
M
DB
PV
PM
GC
1

PLATE 2

EXPLANATION OF FIGURE

2 Longitudinally cut smooth muscle cells from an ovariectomized rat showing ultrastructural similarities to those seen in figure 1. More mitochondria (M), pinocytotic vesicles (PV) and fewer dense bodies (DB) are visible. × 20,720.

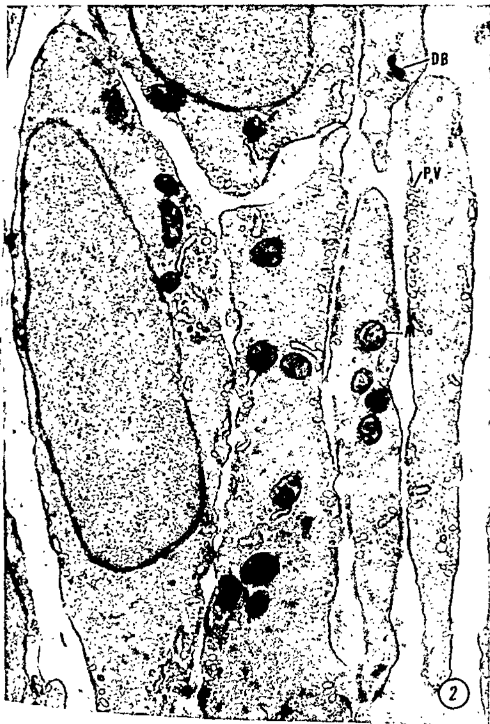

DB
P.V
2

PLATE 3

EXPLANATION OF FIGURES

3 An electron micrograph demonstrating the beginning of accumulation of granular endoplasmic reticulum (GER), mitochondria (M), Golgi complex (GC) and free ribosomes (R) at the nuclear pole. A dense body (DB) lies in close relation to the nucleus (N). The rat was autopsied 24 hours after estrogen stimulation. × 30,900.

4 This electron micrograph demonstrates a portion of a smooth muscle cell of the uterus from an ovariectomized animal autopsied 48 hours after a single injection of estrogen. There is some interdigitation (ID) of adjacent cells. There is a great accumulation of glycogen (G), free ribosomes (R) and granular endoplasmic reticulum (GER) at the nuclear pole. A few mitochondria (M) and multivesicular bodies (MVB) are located in this area. Ribosomes (R) are present in the outer nuclear membrane (ONM). × 20,720.

PLATE 4

FXPLANATION OF FIGURES

5 Cytoplasmic extensions of two smooth muscle cells are seen in this electron micrograph. Interdigitation (ID) of adjacent cells is evident. Note the marked accumulation of glycogen (G) granules close to the plasma membrane (PM). There are some free ribosomes (R) and scattered mitochondria (M) in the cells. The animal was autopsied 48 hours after estrogen treatment. × 20,720.

6 An electron micrograph demonstrating an accumulation of ribosomes (R) and granular endoplasmic reticulum (GER) at the nuclear pole of a smooth muscle cell. Also in this area are a few glycogen (G) granules, a dense body (DB), a few mitochondria (M) and a multivesicular body (MVB). In an adjacent cell (top right) glycogen (G) granules are present. The animal was autopsied 48 hours after estrogen stimulation. × 20,720.

PLATE 5

EXPLANATION OF FIGURES

7 This electron micrograph shows an accumulation of structures at the nuclear pole of the smooth muscle cell. Note the presence of the granular endoplasmic reticulum (GER), mitochondria (M), Golgi complex (GC) and free ribosomes (R). A portion of the nucleus (N) can be seen. The rat was autopsied 72 hours after estrogen treatment. × 30,900.

8 A cytoplasmic extension of a smooth muscle cell from an animal autopsied 96 hours after estrogen stimulation. Mitochondria (M), granular endoplasmic reticulum (GER), free ribosomes (R), scattered glycogen (G) particles and Golgi complex (GC) occupy almost the entire area. × 30,900.


G
GC
GER
M
8

Morphogenesis of the Placenta and Fetal Membranes of the Tree Shrews (Family Tupaiidae)[1]

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ABSTRACT The paucity of information on the early developmental stages and the necessity for re-evaluation of the phylogenetic implications of the tupaiid fetal membranes have stimulated the present study.

Neonatal tupaiids possess prominent bilateral endometrial pads that run the length of each uterine horn. These gland-free pads are the sites of implantation and placentation in adults. Implantation is bilateral and superficial. Multinucleated giant cells differentiate from the cytotrophoblast of the attachment sites; they facilitate the marginal expansion of the placental disc by detaching the paraplacental epithelium from its basement membrane.

Prominent bilateral chorio-vitelline placentae develop and remain functional through early limb bud stages. Decidual cells fuse to form "decidual knots" during this period.

The bilateral chorio-allantoic placentae are labyrinthine endotheliochorial. The trophoblast of the labyrinth is entirely syntrophoblastic, similar to the condition in the armadillo placenta. Epithelioid mesenchymal cells in the labyrinth are strikingly similar to those of the shrew placenta. The large allantoic vesicle and free vascular yolk sac persist. Amniogenesis occurs by folding.

Uterine glands remain secretory throughout pregnancy; iron is probably supplied to the fetus in this manner.

The tree shrews of Southeast Asia possess many characteristics that suggest affinities with the mammalian orders Insectivora and Primates, but the absence of unquestioned fossil tupaiids obscures their precise phylogenetic relationships. Many authors classify the family Tupaiidae within the suborder Prosimii of the Primates (Fiedler, '56; Clark, '62; Buettner-Janusch, '66). Recent re-examinations of the skeleton and dentition (van Valen, '65; McKenna, '66) and the nervous system (Campbell, '66), as well as immunological comparisons of serum proteins (Goodman, '67), suggest that the Tupaiidae are not as closely related to the prosimian Primates as previously believed.

The lack of unquestioned fossil remains necessitates the search for comparative anatomical and biochemical characteristics that may be useful in determining the phylogenetic affinities of tree shrews. Mossman's ('37, '53) extensive comparative analysis suggests that the developmental characteristics of the placenta and fetal membranes have remained conservative features during the evolution of mammals, presumably because of their relative isolation from the selective effects of the external environment. This conservative nature of the fetal membranes suggests that they have considerable significance in characterizing the higher taxa, particularly at the ordinal and subordinal levels.

Hubrecht (1894b, 1895, 1899, '08) examined over 450 *Tupaia javanica* in all stages of pregnancy. The single fetus in each uterine horn is connected to two bilateral placental discs. Hubrecht observed further that implantation, formation of the yolk sac placentae, and their subsequent replacement by the chorio-allantoic placentae all occur on the lateral walls of each horn. These specialized sites are evident in the virgin uterine horn as gland-free endometrial "cushions." Hubrecht reported that the invasive chorio-allantoic villi surround the maternal capillaries, and that the capillary endothelium then degenerates, resulting in the circulation of maternal blood within trophoblastic lacunae.

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On the basis of Hubrecht's description, de Lange ('33) included the placentation of *Tupaia* within the labyrinthine hemochorial type of Grosser ('09).

Mossman ('39) stated that the placenta of the tree shrew is endotheliochorial, but he presented no evidence supporting this view. Actually, his conclusions were based upon his unpublished observations of several specimens of *Tupaia javanica* that J. P. Hill had obtained from the Hubrecht Laboratory (Mossman, '67a).

Van der Horst ('49) disagreed with Hubrecht concerning the fate of the maternal capillary endothelium in the placental labyrinth. He traced the endothelial continuity between the maternal arterioles entering the placenta and the capillaries in the labyrinth. He stated that maternal capillary endothelium persists in late stages of pregnancy and classified the *Tupaia* placenta as endotheliochorial.

Meister and Davis ('56, '58) examined placental material from *Tupaia minor* and *Tupaia tana* (= *Lyonogale tana* Conisbee '53), consisting of one early and two late stages of chorio-allantoic placentation. They described the earlier stage as labyrinthine endotheliochorial, but they believed that the maternal endothelium is lost in the labyrinth of the near term specimens, and thus classified them as labyrinthine hemochorial. Meister and Davis concluded that the fetal membranes of *Tupaia* are significantly different from those of the Insectivora, but they found a similarity between the fetal membranes of *Tupaia* and those of the more generalized members of the families Cebidae and Cerocopithecidae of the Primates. This evidence of the placenta and fetal membranes has been used to support the contention that the tupaiids are closely related to the Primates (Clark, '62; Buettner-Janusch, '66).

Recent studies of the placentation of *Tupaia and Anathana* support van der Horst's ('49) conclusion that the endotheliochorial relationship persists in late stages of pregnancy (Hill, '65; Verma, '65). Furthermore, Hill contended that there is no justification for classifying the tupaiids with the Primates on the basis of their placentation.

The lack of agreement about certain details of placentation, the paucity of information available on the earlier developmental stages of the fetal membranes, and the necessity for re-evaluation of the phylogenetic implications of the morphogenesis of their fetal membranes have stimulated the present study.

MATERIALS AND METHODS

The 52 specimens examined during this study belong to 4 of the 6 genera of tupaiids recognized by Lyon ('13). Reproductive tracts were obtained from 10 *Tupaia* javanica, 6 *T. longipes*, 6 *T. minor*, 5 *T. glis*, 4 *T. palawanensis*, 1 *T. picta*, 1 *T. chinensis*, 1 *T. montana*, 1 *T. gracilis*, 1 *Lyonogale tana*, 6 *Urogale everetti*, and 1 *Dendrogale murinus*. The material was collected in Malaysia, Thailand and the Philippine Islands through the cooperation of a number of individuals. The majority of the specimens were obtained from Professor Clinton H. Conaway, Department of Zoology, University of Missouri and Doctor Kenneth L. Duke, Department of Anatomy Duke University Medical Center. Professor B. I. Balinsky, Department of Zoology, University of the Witwatersrand, South Africa kindly loaned the collection of slides prepared by the late C. J. van der Horst (see van der Horst, '49). Single preserved reproductive tracts were obtained from each of the following: Barbara Harrisson, the Sarawak Museum, Malaysia (see Harrisson, '63; Doctor Joseph C. Moore, Chicago Natural History Muscum; Professor Clinton N. Woolsey, Department of Neurophysiology, University of Wisconsin Medical School; and Doctor P. D. Nieuwkoop, Hubrecht Laboratory, Utrecht, the Netherlands.

The reproductive tracts were fixed in 10% formalin, Bouin's fixative or AFA (alcohol, formalin and acetic acid), and subsequently stored in 70% alcohol. Paraffin sections were cut at 6–10 μ; serial sections were mounted and stained from the early stages of pergnancy, whereas only every tenth section was examined from some of the later stages. The ovaries were also sectioned in order to provide additional information about the reproductive history of these animals. Sections were stained with Groat's tetrachrome (unpublished). Harris' hematoxylin and eosin, Masson's trichrome, Movat's pentachrome,

an silver, PAS (Periodic Acid-Schiff), mori's basic fuchsin, Turnbull's blue, or tchison's iron stain.

Measurements of the greatest diameters length of the early developmental stages re recorded from serial section slides th the aid of an ocular micrometer. ross measurements were made of the own-rump length of preserved embryos henever possible.

OBSERVATIONS

To understand the uterine changes that cur during pregnancy, it will be useful consider briefly the uterine histology of e immature and non-pregnant adult tree rew. The elongate uterine horns unite a short common corpus; the general ternal appearance of the uterus has been scribed previously (Wood Jones, '17; leister and Davis, '56; Clark, '62).

Non-pregnant uterus

At 12 days postnatum the endometrium f each horn exhibits two characteristic gions: (1) the mesometrial and anti-mesometrial surfaces bear short, straight ubular glands in the superficial third of he endometrium, and (2) gland-free endo-metrial pads or cushions occupy the lateral r orthomesometrial walls and bulge rominently into the uterine lumen. These ads are actually bilateral ridges which ex-tend the entire length of the horn. In cross ection the pads are fungiform. (fig. 5).

The subepithelial zone of the endometrial pads consists of densely packed cellular stroma, whereas the stroma of the base of the pads and of the mesometrial and anto-mesometrial regions is more loosely ar-ranged. The columnar epithelial lining of the pads measures about 13 μ in height. Prominent endometrial pads are also seen in the uterine horns of the neonate, but there is no sign of uterine gland formation at this stage.

Uterine glands extend to the base of the endometrium by 50 days of age. The endo-metrial pads remain gland-free, although coiled gland tubules occur in the stroma immediately beneath them. Serial sections show that these basal glands curve around the pads to open into the uterine lumen. The pads are similar in appearance to those of earlier stages.

In more mature nulliparous specimens the endometrial pads are less prominent because of the relatively greater thickness of the glandular endometrium (fig. 6). The basal glands are coiled slightly, and the antimesometrial glandular endometrium is about three times as thick as that of the mesometrial region.

During the preovulatory phase of the estrous cycle of mature tupaiids the basal halves of the uterine glands are moderately coiled. At this stage the gland-free endome-trial pads no longer bulge into the uterine lumen; hyperplasia of the adjacent glan-dular endometrium has resulted in a rela-tively uniform thickness of the lateral walls. The arterioles which supply the endo-metrial pads of parous animals are coiled in the region between the myometrium and the base of the pads, but there is little or no evidence of coiling of the smaller arterioles which directly supply the pads (fig. 7).

Preimplantation stages of pregnancy

In one specimen of *Tupaia glis* the ampulla of each oviduct contains an eight cell stage surrounded by the zona pellucida. These cleavage stages measure 76 × 84 μ and 82 × 89 μ, exclusive of the zona; in comparison, the average dimensions of ova from six Graafian follicles in *T. glis* are 114 × 123 μ.

The earliest blastocyst found occurs in the isthmus of one oviduct of a specimen of *Urogale everetti;* three late morulae also occur in the same region. The tubal blasto-cyst measures 107 × 108 μ. A unilaminar blastocyst becomes distributed to the mid-dle region of each uterine horn, even in those cases where double ovulation occurs in one ovary and none in the other. At this stage the inner cell mass is 3–4 cells thick; there is no thinning of the trophoblast (Rauber's layer) covering the inner cell mass.

Two specimens contain expanded free bilaminar blastocysts in the middle of each uterine horn (fig. 8). The cuboidal tropho-blastic epithelium is about 10 μ thick and is considerably thinner than in earlier stages. Rauber's layer is complete in the younger of these two blastocysts, but it is reduced in thickness and incomplete in places over the embryonic knot of the older

Abbreviations

A, Amnion
AD, Allantoic duct
AM, Allantoic mesenchyme
AP, Attachment plaque
AV, Allantoic vesicle
B, Basement membrane
C, Cytotrophoblast
CM, Basal chromatic masses
D, Basal decidua
E, Endothelium
EC, Endometrial capillaries
ED, Endoderm
EX, Exocoelom
G, Cytoplasmic granules
K, Decidual knots
L, Labyrinth
M, Macrophage
P, Endometrial pad
PV, Primary villi
S, Syntrophoblast
SC, Smooth chorion
T, Trophoblast
TG, Trophoblast giant cell
U, Uterine epithelium
UG, Uterine gland
VS, Vascular splanchnopleure
VV, Vitelline vessels
Y, Yolk sac cavity

specimen. A squamous endodermal layer forms a continuous internal lining for the blastocyst cavity.

There are no remains of the zona pellucida around any of the expanded free blastocysts. An attenuated zona surrounds the unilaminar uterine blastocysts; but it is uncertain whether loss of the zona results from continued stretching or from its dissolution as a result of acid fixation. The zona still surrounds the expanded unimplanted blastocysts of the badger, ferret, mink and rabbit (Wright, '66; Orsini, '67).

Coiling of the uterine glands increases during passage of the blastocysts from the oviduct to the uterus; they are considerably coiled in uteri containing expanded bilaminar blastocysts. The stroma of the endometrial pads remains densely packed and cellular throughout most of the preimplantation period, although some stromal edema is evident in the expanded blastocyst stage.

A single *Tupaia longipes* was sacrificed six days following postpartum copulation in captivity. The ovaries contain two generations of corpora lutea: smaller corpora of lactation and larger corpora of pregnancy. A bilaminar blastocyst occurs on the uterine side of the tubo-uterine junction (fig. 9). Rauber's layer is complete and there is no evidence of a zona pellucida; the general appearance of the blastocyst is very similar to that of the expanded bilaminar preimplantation blastocysts of other specimens. This specimen suggests the possibility of lactational delay of implantation in *Tupaia*, similar to the situation described in the mouse and rat (Enzmann, Saphir and Pincus, '32; Krehbiel, '41). Secretory material is abundant in the lumina of the

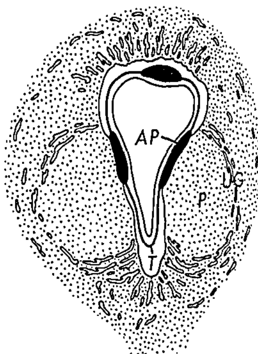


Fig. 1 Diagram of a cross section of the ute showing implantation of the bilaminar blastocyst. Implantation is bilateral and the embryonic is oriented antimesometrially.

uterine glands at the ovarian end of th horn, and a considerable amount of se tory product surrounds and is adsorbed t the surface of the blastocyst.

Implantation

The initial attachment of the bilamin blastocyst is accompanied by the intima apposition of the simple cuboidal layer c trophoblast and the epithelium of the bi lateral endometrial pads. Following this th trophoblast of the attachment site liferates to form a layer 2–6 cells thick

lthough the multilayered cytotrophoblast heres intimately to the uterine epithe-um, the apposed plasma membranes ap-ear to remain intact (fig. 10). The two cell pes may be distinguished on the basis of eir nuclear characteristics. The tropho-lastic nuclei are slightly larger and con-in one or two large, centrally located ucleoli. Scattered fine chromatin clumps ay occur in the nucleus, but its general ppearance is open or vesicular. Nuclei of terine epithelial cells are spherical and ontain small nucleoli; the nucleoplasm is elatively homogeneous and basophilic figs. 10, 11).

A specimen of *Tupaia longipes* in an arly stage of implantation bears irregular hallow invaginations or crypts over the ntire epithelial lining of the uterine horn. The crypts measure 55–120 μ in depth, and terine glands open into them except on he gland-free endometrial pads (figs. 11, 12). The attaching trophoblast has partially penetrated into the lumina of the endo-metrial pad crypts, but there is no apparent rupture of the uterine epithelium.

A slightly later stage of implantation bears prominent bilateral "attachment plaques" approximately 100 μ thick, consist-ing of the multilayered attaching tropho-blast and a nearly complete layer of uterine epithelium. Occasional trophoblastic cells have sent processes between epithelial cells toward the underlying basement mem-brane, but there is little evidence of epithe-lial degeneration (fig. 13). Trophoblastic and epithelial nuclei remain distinguishable on the basis of the previously mentioned criteria, and plasma membranes persists between the two cell types.

By the time the attachment plaques are about 130 μ thick, numerous crenated, pyknotic nuclei have appeared in them (fig. 14); these probably represent degene-rating uterine epithelial cells. The tropho-blastic and epithelial elements of the plaques are radially arranged; this prob-ably reflects the interdigitation of tropho-blast and the irregular epithelial crypts seen in an earlier phase of attachment. Some relatively normal appearing epithelial cells occur in the bases of the plaques, but it is impossible to identify some of the other plaque cells with certainty.

The early attaching blastocyst occupies an enlargement of an the antimesometrial third of the uterine lumen, and then nar-rows between its bilateral attachments to the endometrial pads (fig. 1). The em-bryonic knot of the late preimplantation stage has become an enlarged flattened disc. Nuclei of the pseudostratified epithe-lium of the disc are arranged in two ir-regular layers. There is no evidence of a primitive streak in any of these early at-taching blastocysts. A thin, homogeneous, acidophilic lamina separates the tropho-blast and endoderm in the mesometrial half of the blastocyst; it is analogous to the Reichert's membrane characteristic of rodents, shrews and some bats, but is not nearly as thick.

The endometrial pad stroma is edematous during the initial period of blastocyst at-tachment. A thin subepithelial zone of stroma is more compact, possibly due to pressure from the expanded blastocyst. In cross sections of the uterus the maternal capillaries form a characteristic radiating pattern throughout the endometrial pad (fig. 15). The stroma of the superficial fifth of the pads is more compact in later attach-ment stages, apparently due to a slight hypertrophy of the stromal cells rather than to hyperplasia (fig. 13).

Implantation in the tree shrews is there-fore bilateral and superficial, and the em-bryonic shield is oriented antimesometri-ally. There is no decidual reaction at the initial stage of attachment (fig. 1).

Primitive streak stage

The thickened attachment plaques show little or no evidence of remnants of uterine epithelium. They now consist of two dis-tinct zones (fig. 16). (1) An inner cytotro-phoblastic zone 2–5 cells thick is separated from the endoderm by Reichert's mem-brane. There are scattered mitotic figures in this zone, and the cells are similar to those of earlier implantation stages. (2) An outer zone of hypertrophied tropho-blastic cells abuts upon the maternal stroma. At least some of these radially elongated trophoblastic "giant" cells are multinucleate, but plasma membranes separate adjacent multinucleated masses. Mitotic figures were never seen in them. The intensely basophilic cytoplasm of the

giant cells contrasts sharply with the pale staining cytoplasm of the inner cytotrophoblastic cells, and the diameter of giant cell nuclei is about twice that of the cytotrophoblastic cells.

The basement membrane of the uterine epithelium which previously covered the endometrial pads is still demonstrable in the two specimens of this stage (fig. 17). It appears intact beneath the antimesometrial and mesometrial borders of the attached trophoblast, and is continuous with the basement membrane of the uterine epithelium adjacent to the attachment plaques. The basement membrane is incomplete beneath the mid-region of the plaque, where it is penetrated in places by multinucleated giant cells. At both margins of the attachment site wedge-like processes of trophoblastic giant cells seem to detach the uterine epithelium from its underlying basement membrane (fig. 18).

The endometrial pad stroma is composed predominantly of closely packed, enlarged, epithelioid decidual cells with basophilic cytoplasm (figs. 16, 18). There is a gradation from basally located fibroblast-like cells to the closely packed epithelioid cells adjacent to the attachment plaque. The decidual cells have vesicular nuclei with one or two prominent nucleoli. Those decidual cells immediately adjacent to the trophoblast are further modified; their cytoplasm is more basophilic and their nuclei contain increased amounts of clumped chromatin which tend to be located next to the nuclear membrane (fig. 19). Plasma membranes are increasingly difficult to detect around individual hypertrophied stromal cells, and the decidual cells bordering the trophoblast appear to be multinucleated masses. Mitoses are frequent in the compacted decidual cells, but are less numerous in the thin basal zone of unmodified and more edematous stroma.

There is no evidence of blood island formation in either specimen of this stage. Reichert's membrane is continuous with a slightly thinner basement membrane of identical staining affinities which underlies the antimesometrial trophoblast and the embryonic ectoderm. The fact that the extraembryonic mesoderm separates both trophoblast and Reichert's membrane from the endoderm suggests that this membrane in the tupaiid blastocyst consists largely of the basement membrane of the trophoblast, similar to the condition in the Soricidae (Brambell and Perry, '45; Wimsatt and Wislocki, '47).

Development of chorio-vitelline placentation

Avascular trilaminar omphalopleure covers the antimesometrial surface of the endometrial pads in a specimen of *Lyonogale* containing a 6-somite embryo, whereas the remainder of the surface of the pad is covered by bilaminar omphalopleure. The giant cell layer characteristic of the primitive streak stage is now considerably reduced; only scattered giant cells occur between the multilayered cytotrophoblast and the decidual cells. A few giant cells have penetrated into the decidual zone, but the cytotrophoblast shows no evidence of such penetration.

The decidual cells are densely packed and show numerous mitotic figures. Some of the decidual cells adjacent to the trophoblast form syncytial "decidual knots" consisting of a peripheral ring of nuclei and a central cytoplasmic core (fig. 21). The decidual knot nuclei contain abundant peripheral chromatin clumps. The knots are more numerous adjacent to the attached trophoblast, but they are also found in the deeper regions of the pad.

The incipient placental discs show three prominent zones by the time vascular mesoderm has extended into their antimesometrial margins (figs. 20, 21, 22). (1) The inner trophoblastic zone at the fetal surface of the disc consists of a thin layer of cytotrophoblast which commonly surrounds maternal capillaries. Densely nucleated, irregular processes extend from the cytotrophoblast into the decidua as primary villi; they appear to be syncytial in nature, but this is difficult to ascertain. Frequently one or several giant cells appear to be embedded in the core of the primary villus, but they are almost completely obscured by the surrounding smaller nuclei. Many primary villi completely or partly surround decidual knots (fig. 21).

(2) The decidual zone adjacent to and intermingled with the primary villi is characterized by numerous, more developed decidual knots (fig. 21). In a cross section

a knot the hyperchromatic nuclei are anged in a semilunar or circular pattern about the periphery, and the center nsists of a common acidophilic cytosm. The greatest diameter of the erical knots is about 30 μ, and about 9 nuclei are located in the periphery of mot in cross section. The irregular spaces the loosely arranged decidual knot zone ty be due to tissue dissolution or to ema.

(3) The relatively normal basal decidua separated from the flattened uterine that underlie the disc by a thin of typical spindle-shaped stromal lls. Mitotic figures are numerous in the sal decidua but are absent in the decidual There is a transitional zone between rmal basal decidual cells and decidual (fig. 22). Maternal capillary endo- remains intact in the decidual knot ne, despite considerable tissue dissolution this region. These capillaries are often letely surrounded by cytotrophoblast the fetal surface of the incipient placntal disc (fig. 21).

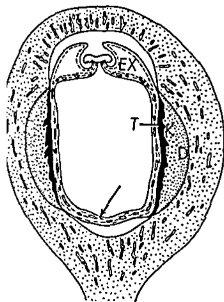


Fig. 2 Diagram of a cross section of the uterus luring the period of chorio-vitelline placentation. Trilaminar omphalopleure (arrow) extends beween the mesometrial margins of the developing lacental discs. Mesoderm, dashed lines.

In 12-somite embryos mesometrial extension of vascular extraembryonic mesoderm results in the establishment of bilateral chorio-vitelline placental discs (fig. 25). The bilaminar omphalopleure persists temporarily between the mesometrial margins of the placental discs, but the continued mesometrial extension of vascular mesoderm soon results in a vascular trilaminar omphalopleure (fig. 2).

The cytotrophoblast at the fetal surface of the chorio-vitelline placentae varies in thickness from 2–4 cells in the early stages to ten or more cells later. Mitoses are frequent in the cytotrophoblast, especially during the early period of chorio-vitelline placentation. The maternal capillaries which are surrounded by the cytotrophoblast of the chorionic plate tend to run in a mesometrial-antimesometrial plane (fig. 24). Their endothelium remains intact, and its continuity can be traced with that of the thin-walled, radial arterioles which course through the basal decidua up to the chorionic plate. In many instances maternal capillaries are separated from the fetal vessels of the chorio-vitelline placentae by a layer of cytotrophoblast only 1–2 cells thick.

Densely nucleated, primary trophoblastic villi are well developed in the chorio-vitelline placentae and grade into the cytotrophoblast of the chorionic plate (figs. 23, 24). Mitoses are absent from the multinucleated or syncytial primary villi. Trophoblastic giant cells are rare in the main portion of the chorio-vitelline placental disc. However, paraplacental trophoblastic giant cells are conspicuous throughout chorio-vitelline placentation; they continue to detach or delaminate the uterine epithelium from its basement membrane in this region, and to invade the paraplacental uterine glands (fig. 26). This paraplacental epithelial delamination appears to play a role in the marginal expansion of the placental discs, since the discs come to occupy the entire lateral walls of the uterine horn.

The zone of decidual knots becomes reduced in thickness in later stages of chorio-vitelline placentation. The knots may become surrounded by the proliferating cytotrophoblast of the chorionic plate (fig. 27), or they may be partly surrounded by the densely nucleated primary villi (figs. 24,

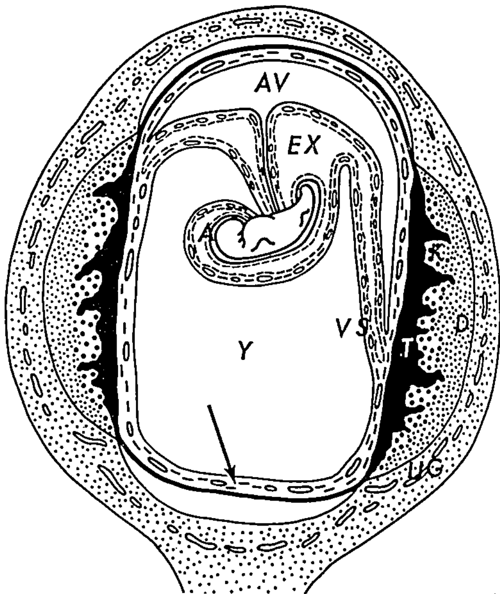


Fig. 3 Diagram of a cross section of the uterus during the transition between chorio-vitelline and chorio-allantoic placentation. The vascular splanchnopleure of the yolk sac still adheres to the mesometrial region of the placental discs, and trilaminar omphalopleure (arrow) persists between the mesometrial borders of the placental discs.

25). Later many decidual knots show increased nuclear pyknosis and a loss of the characteristic peripheral nuclear arrangement (fig. 23). Mitotic figures are abundant in the normal appearing basal decidua, but they are absent in the thinner zone of decidual knots. The basal decidua may form up to half the thickness of the placental disc.

Transition between chorio-vitelline and chorio-allantoic placentation

Two specimens were collected in the early limb bud stage. A very short vitelline duct connects the yolk sac to the midgut The patent allantoic duct leaves the ver tral body wall and runs antimesometriall to open into the large allantoic vesicle which has spread bilaterally and mesome trially (fig. 3). The vascular allantois ha displaced the vascular splanchnopleure o the yolk sac from the surface of the pla cental discs throughout the antimesome trial half of their extent. The mesometria portion of each placental disc remains covered by the vascular splanchnopleure o the yolk sac and apparently still function as a chorio-vitelline placenta (figs. 28, 29)

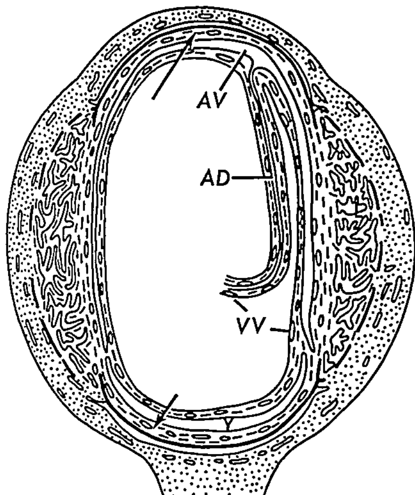


Fig. 4 Diagram of a cross section of the uterus during late pregnancy. The smooth chorion is fused with the outer layer of the allantois and of the yolk sac to form a chorio-allantoic membrane (long arrow) and a chorio-vitelline membrane (short arrow). Vitelline vessels run along the periphery of the umbilical cord and the proximal wall of the allantoic vesicle to vascularize the free yolk sac. No attempt is made to portray the detailed structure of the labyrinth in this diagram.

Short, blunt processes of vascular al-ntoic mesoderm indent the chorionic plate the antimesometrial portion of the pla-ntal discs, but these vascular buds have t yet penetrated the chorionic plate to rm an intimate relationship with the aternal capillaries. The prominent, dense-nucleated primary villi extend through out 60% of the thickness of the placen-l disc. Scattered mitoses occur in the orionic plate and basal decidua, but are cking in the pri-

Chorio-allantoic placentation

The placental disc during the progressive establishment of chorio-allantoic placentation consists of four zones which tend to interdigitate with each other: (1) chorionic plate, (2) chorio-allantoic labyrinth, (3) a mixed zone of primary villi and decidual knots, and (4) a reduced zone of basal decidua (fig. 30).

The multilayered cytotrophoblast of the ch[illegible] plate is about 80–180 μ thick [illegible]is period, and mitotic figures are

infrequent. Vascular allantoic mesenchymal villi, surrounded by a single layer of trophoblast, penetrate the chorionic plate to form an intimate relationship with maternal capillaries. These villi have carried portions of the chorionic plate with attached primary villi deep into the basal decidual zone (figs. 30, 31) as trophoblastic "cell columns."

The mesenchymal cells of the villus-like processes are hypertrophied and epithelioid in appearance. Their spherical or ovoid nuclei are vesicular and contain prominent nucleoli; their cytoplasm is lightly acidophilic (figs. 33, 35, 37, 38, 39). Transitional cell types occur between these and the spindle-shaped, unmodified mesenchymal cells of the fetal surface of the chorionic plate.

The attenuated layer of trophoblast covering the villus-like processes of the labyrinth is completely syntrophoblastic, and it is continuous with the cytotrophoblast of the chorionic plate and the multinucleated cell columns at the tips of the villus-like processes (fig. 32). Maternal capillary endothelium persists in the developing labyrinth and is considerably hypertrophied in places, especially adjacent to the chorionic plate (fig. 33). The vesicular maternal endothelial nucleus is spherical or ovoid and contains 1–2 prominent basophilic nucleoli. The flattened syntrophoblastic nuclei are filled with densely packed chromatin clumps and are intensely basophilic.

Multinucleated primary villi extend almost to the base of the placental disc, where they interdigitate with decidual knots or the reduced region of unmodified decidua basalis (fig. 31). As development proceeds, the labyrinth comes to occupy more and more of the thickness of the placental disc, while the mixed zone of cell columns and decidua becomes reduced to a thinner region of irregular, densely nucleated masses (fig. 34). These masses probably represent remnants of cell columns; their densely packed nuclei are filled with clumped chromatin, similar to the condition in the cell columns. Occasional decidual knots and small clusters of pyknotic nuclei, which probably represent degenerating decidual knots, also occur.

The major change in the placental during later stages of pregnancy is continued reduction in thickness of basal densely nucleated masses. If multinucleated masses are indeed of cell columns, they would be with the *trophospongium* of other thine placentae, that is, the trop zone that contains only maternal channels. However, the possibility ex that the basal decidua or decidual kn may also contribute to the mul masses, and therefore the non-specif "basal chromatic masses" is used to to these structures. Some of the basal matic masses exhibit two or more cell and some of these are similar to pyknotic, degenerating decidual knots earlier stages (fig. 35). Villus-like from the labyrinth spread out and flattened against the basal masses. In placentae obtained from norm term deliveries, the basal chromatic are generally less than 50 μ thick and absent from much of the bases of the p cental discs. At parturition the separate from the uterus at the between the chromatic masses and the th layer of unmodified stroma that the flatened uterine glands.

Near term placental discs are in cross section, and most of their thick consists of the labyrinth or zone of intimacy (fig. 36). The villus-like ance of the labyrinth reflects antly radiate arrangement of the allan mesenchymal villi (fig. 37). However, are no true chorio-allantoic villi, as trophoblast which surrounds the capillaries is continuous throughout labyrinth (fig. 38).

The hypertrophied maternal endothelium persists to term in the rinth of the chorio-allantoic placentae. basement membranes of both the phoblast and the maternal endothelium demonstrated by PAS, azan silver and lory's trichrome staining (figs. 39, 40). layers that separate the fetal and mater blood in the chorio-allantoic labyrinth (1) fetal endothelium, (2) the bas membrane(s) between the fetal lium and syntrophoblast, (3) blast, (4) basement membrane of maternal endothelium, and (5)

endothelium. According to Grosser's ('09) scheme, the chorio-allantoic placenta of the tupaiids is therefore endotheliochorial. This endotheliochorial condition is also evident in placentae obtained from normal term deliveries (fig. 41). Hyalinization of the basement membranes of the labyrinth at term renders them visible with routine staining procedures.

Placental circulation in tupaiids is similar to that of most species possessing a labyrinthine placenta. Maternal arterioles from the myometrium pass through the labyrinth and open into capillaries at the fetal surface of the placental disc. The modified maternal capillaries follow a predominantly radial course to the base of the placental discs, where they drain into uterine veins. Fetal arterioles penetrate deeply into the placental labyrinth before breaking up into capillaries which pursue a rather tortuous course toward the allantoic veins at the fetal surface of the placentae. The countercurrent flow of the tupaiid placental circulation follows the general scheme postulated for most, if not all, mammalian placentae (Mossman, '65).

Amniogenesis

The earliest evidence of amniogeneis among the available specimens occurs in a 6-somite embryo of *Lyonogale*. The amniotic tail fold covers the caudal tip of the embryo, but there is only a slight suggestion of the amniotic head fold. On the other hand, in a 12-somite *Tupaia javanica* embryo the amniotic head fold extends over the anterior third of the embryo; but there is no suggestion of an amniotic tail fold. In the later stages of chorio-vitelline placentation the amniotic folds extend over the anterior and posterior ends of the embryo, but the amnion remains incomplete over the midregion. The amniotic folds are not completely closed until the allantoic diverticulum fuses with the chorion to initiate chorio-allantoic placentation.

Yolk sac

The antimesometrial wall of the yolk sac is invaginated by the expanded amnion and embryo during the transitional stage between chorio-vitelline and chorio-allantoic placentation. For convenience the antimesometrial concave surface of the yolk sac may be considered the proximal layer, and the convex mesometrial surface may be considered the distal layer (fig. 3). After its detachment from the surface of the placental disc by the spreading allantois, the yolk sac becomes compressed between the amnion and the mesometrial margins of the placental discs (fig. 4).

In early stages of chorio-allantoic placentation the intestinal loop herniates into the umbilical coelom (fig. 42), but it is no longer connected with the yolk duct. In the mid-mesometrial region of the proximal wall of the yolk sac a short endodermal diverticulum about 0.5 mm long represents the distal remnant of the yolk duct (fig. 43). It is flanked by the two primary branches of the vitelline vein, and its blind end is closely associated with the vitelline artery. There is no herniation of the intestinal loop into the umbilical cord in later stages of chorio-allantoic placentation, and there is no continuity between the endoderm of the midgut and that of the yolk sac. The free yolk sac remains large although compressed throughout the remainder of pregnancy, and it extends the entire anterior-posterior length of the fetus. The single vitelline artery and vein leave the umbilical cold antimesometrially and run laterally and mesometrially between the amnion and allantois toward the yolk sac (fig. 4).

The distal splanchnopleure of the yolk sac vascularizes the overlying mesometrial smooth chorion, forming a chorio-vitelline membrane, and the proximal splanchnopleure of the yolk sac is similarly associated with the amnion to from a vascular amnio-vitelline membrane (fig. 4). During the early stages of chorio-allantoic placentation the distal endoderm of the yolk sac is generally 2–3 times as thick as the proximal layer, particularly toward the mid-mesometrial region (fig. 44). The distal splanchnopleure also possesses a richer vascular supply than does the proximal splanchnopleure. The most pronounced modification of the chorio-vitelline membrane occurs in a specimen of *Tupaia palawanensis*. The mid-mesometrial chorion has invaginated to form a pocket or sac which opens into the uterine lumen (fig. 45). Villus-like processes of the distal endoderm penetrate the folds or plicae

infrequent. Vascular allantoic mesenchymal villi, surrounded by a single layer of trophoblast, penetrate the chorionic plate to form an intimate relationship with maternal capillaries. These villi have carried portions of the chorionic plate with attached primary villi deep into the basal decidual zone (figs. 30, 31) as trophoblastic "cell columns."

The mesenchymal cells of the villus-like processes are hypertrophied and epithelioid in appearance. Their spherical or ovoid nuclei are vesicular and contain prominent nucleoli; their cytoplasm is lightly acidophilic (figs. 33, 35, 37, 38, 39). Transitional cell types occur between these and the spindle-shaped, unmodified mesenchymal cells of the fetal surface of the chorionic plate.

The attenuated layer of trophoblast covering the villus-like processes of the labyrinth is completely syntrophoblastic, and it is continuous with the cytotrophoblast of the chorionic plate and the multinucleated cell columns at the tips of the villus-like processes (fig. 32). Maternal capillary endothelium persists in the developing labyrinth and is considerably hypertrophied in places, especially adjacent to the chorionic plate (fig. 33). The vesicular maternal endothelial nucleus is spherical or ovoid and contains 1–2 prominent basophilic nucleoli. The flattened syntrophoblastic nuclei are filled with densely packed chromatin clumps and are intensely basophilic.

Multinucleated primary villi extend almost to the base of the placental disc, where they interdigitate with decidual knots or the reduced region of unmodified decidua basalis (fig. 31). As development proceeds, the labyrinth comes to occupy more and more of the thickness of the placental disc, while the mixed zone of cell columns and decidua becomes reduced to a thinner region of irregular, densely nucleated masses (fig. 34). These masses probably represent remnants of cell columns; their densely packed nuclei are filled with clumped chromatin, similar to the condition in the cell columns. Occasional decidual knots and small clusters of pyknotic nuclei, which probably represent degenerating decidual knots, also occur.

The major change in the placental discs during later stages of pregnancy is [illegible] continued reduction in thickness of [illegible] basal densely nucleated masses. If these multinucleated masses are indeed [illegible] of cell columns, they would be homologous with the *trophospongium* of other labyrinthine placentae, that is, the trophoblastic zone that contains only maternal blood channels. However, the possibility [illegible] that the basal decidua or decidual knots may also contribute to the multinucleated masses, and therefore the non-specific "basal chromatic masses" is used to refer to these structures. Some of the basal chromatic masses exhibit two or more cell types and some of these are similar to the pyknotic, degenerating decidual knots of earlier stages (fig. 35). Villus-like processes from the labyrinth spread out and become flattened against the basal chromatic masses. In placentae obtained from normal term deliveries, the basal chromatic masses are generally less than 50 µ thick and are absent from much of the bases of the placental discs. At parturition the placentae separate from the uterus at the junction between the chromatic masses and the thin layer of unmodified stroma that overlies the flatened uterine glands.

Near term placental discs are lenticular in cross section, and most of their thickness consists of the labyrinth or zone of vascular intimacy (fig. 36). The villus-like appearance of the labyrinth reflects the predominantly radiate arrangement of the allant[illegible] mesenchymal villi (fig. 37). However, there are no true chorio-allantoic villi, as the trophoblast which surrounds the maternal capillaries is continuous throughout the labyrinth (fig. 38).

The hypertrophied maternal capillary endothelium persists to term in the labyrinth of the chorio-allantoic placentae. The basement membranes of both the syntrophoblast and the maternal endothelium are demonstrated by PAS, azan silver and Mallory's trichrome staining (figs. 39, 40). The layers that separate the fetal and maternal blood in the chorio-allantoic labyrinth are: (1) fetal endothelium, (2) the basement membrane(s) between the fetal endothelium and syntrophoblast, (3) syntrophoblast, (4) basement membrane of the maternal endothelium, and (5) maternal

and Wislocki, '47). The cushions develop as the result of hypertrophy and hyperplasia of endometrial stromal cells, and implantation occurs on the thickened epithelium of these cushions. The soricid endometrial cushions differ from the endometrial pads of the tupaiids in that they contain uterine glands, and their occurrence is cyclic rather than permanent. Similar bilateral stromal enlargements also occur in uteri of American moles with preimplantation embryos (Prasad, '58). Endometrial modifications also occur in the lateral uterine walls of the insectivorous families Centetidae and Chrysochloridae (Goetz, '37a; Bluntschli, '38; de Lange, '19; Gabie, '60), but the details of their development are incompletely known.

The tupaiid endometrial pads appear to be more homologous to the bilateral mucosal pads of the Soricidae than to the endometrial caruncles of the higher ruminants. Only the highly specialized superfamilies Bovoidea and Cervoidea among the subdivisions of the Artiodactyla possess caruncles. Caruncles are absent in the more generalized suborder Suina (including pigs and the hippopotamus) and in the infraorder Tylopoda (camels and llamas) of the suborder Ruminantia (Mossman, '37; van Lennep, '61). The chevrotains (*Tragulus*) are generally classified as intermediate in position between the Tylopoda and the higher ruminants (Romer, '66), and slightly raised areas of their endometrium opposite the villous regions of the chorioallantois may be interpreted as a primitive form of caruncle development (van Lennep, '61). It appears that the functional similarities of tupaiid endometrial pads and ruminant caruncles are due to convergent evolution.

A dual endometrial arterial supply occurs in the prosimians *Galago senegalensis* (Butler, '62, '64, '67) and *Loris tardigradus* (Ramaswami and Anand Kumar, '65) and in the Old World monkeys, apes and man (Kaiser, '47a). Coiled arterioles supply the more superficial functional zone and shorter, relatively straight arterioles supply the basal glandular endometrium. It has been suggested that the coiled endometrial arterioles of Old World monkeys, apes and man regulate the amount of hemorrhage and tissue loss during menstruation (Kaiser, '47a,b). There is no definite evidence for menstruation in any of the prosimians. Menstruation is microscopic in amount in the New World monkeys *Cebus*, *Ateles* and *Alouatta*, but coiled arterioles are absent in these genera (Kaiser, '47b).

The distribution of coiled endometrial arterioles in tupaiids differs from that of *Loris*, *Galago* and the higher Primates, since coiled arterioles do not penetrate into the endometrial pads of the tupaiid endometrium. (My personal communication reported by Butler ('67) was based on incomplete observations regarding the occurrence of coiled arterioles in the tupaiid endometrium.) The tupaiid endometrial pads are analogous to the functional layer of the hominoid endometrium, since these regions are involved in implantation, placentation and possibly menstruation in both groups. There is some evidence that tree shrews exhibit a slight loss of the epithelium and subepithelial stroma of the endometrial pads following pseudopregnancy (Luckett, '63; Conaway and Sorenson, '66). This may be comparable to the small amount of menstrual loss in the New World monkeys. Pseudopregnant cycles are probably a rare occurrence in wild tree shrews, since postpartum pregnancy appears to be the usual condition and there is no clear evidence for a restricted breeding season. This would account for the nearly complete lack of evidence for menstruation in wild tupaiids.

The only non-primate in which the occurrence of menstruation is well established is the elephant shrew *Elephantulus* (Van der Horst and Gillman, '41). Van der Horst ('54) reported that enormous coiled arterioles in *Elephantulus* are localized in the small mesometrial region where menstruation and implantation normally occur. He also mentioned that uterine coiled arterioles are found in the insectivorous families Centetidae and Erinaceidae, although menstruation has not been reported in either of these families. The phenomenon of menstruation is of little taxonomic value at the present time, because of the lack of experimental or histological evidence for possible microscopic menstruation in Insectivora, prosimian Primates, or other taxa.

formed by the invaginated chorion. The vitelline vessels that supply the distal splanchnopleure and modified chorion are particularly prominent, whereas there is no noticeable specialization of the proximal splanchnopleure of the yolk sac or of its vascular supply.

Allantois

The allantoic duct decreases in diameter during chorio-allantoic placentation, but it remains patent and opens into the large, cup-shaped allantoic cavity which extends over most of the surface of both placental discs and persists until term (fig. 4). The antimesometrial region of the allantois vascularizes the overlying chorion to form a smooth chorio-allantoic membrane, and the proximal wall of the allantois contacts the antimesometrial and lateral regions of the amnion to form an amnio-allantoic membrane. There is no persisting true chorion.

The single specimen of *Dendrogale* is poorly fixed, and hemorrhaging has occurred at the margins of the placental discs. However, some of the paraplacental hemorrhaging may be a normal occurrence, as it is found to a lesser extent in well preserved specimens of the other genera. The bleeding is most noticeable at the antimesometrial margins of the discs, and the trophoblast of the smooth chorio-allantoic membrane appears to phagocytize red blood cells that adhere to its free surface (fig. 46). Nuclei are basally located in the tall columnar trophoblastic cells, and the supranuclear cytoplasm is filled with large vacuoles and fragments of red blood cells. Several pits or invaginations occur in the smooth chorio-allantoic membrane, and these are probably homologous with the chorionic vesicles or areolae seen in many other mammals.

Umbilical cord

The definitive umbilical cord of the tupaiids contains the narrow allantoic duct, paired allantoic arteries and a single allantoic vein. The reduced vitelline vessels are located in the periphery of the cord, but there is no remnant of the vitelline duct. The allantoic vessels diverge to supply both placental discs.

Uterus

The uterine glands are secretory throughout pregnancy, but this activity is most intense through the period of chorio-vitelline placentation. From the time of implantation through the period of establishment of the chorio-vitelline placentae, the secretory product accumulates about, and is adsorbed to, the trophoblast of the bilaminar or trilaminar yolk sac. Material of similar consistency and staining affinity accumulates within the lumen of the yolk sac during this period, but this is less evident in later stages of chorio-vitelline placentation.

In parous animals numerous macrophages containing hemosiderin granules occur in the basal interglandular connective tissue. In many places macrophages are intimately associated with the epithelium of the basal glands, and fine granules which resemble hemosiderin in their color and refractivity are seen in the supranuclear cytoplasm of the gland epithelial cells (fig. 47). Turnbull's blue and Hutchison's iron reaction demonstrate the presence of ferrous or ferric ions in the interglandular macrophages and in the supranuclear cytoplasm of the gland epithelial cells (fig. 48).

DISCUSSION

The bilateral endometrial pads or ridges of the tupaiid uterus appear to be analogous to the uterine caruncles found in most ruminants. Caruncles are ". . . specialised non-glandular areas to which the foetal cotyledons become attached, and are essentially local thickenings of the subepithelial dense connective tissue which elsewhere exists as a thin layer immediately beneath the epithelium." (Amoroso, '52). The tupaiid endometrial pads and the ruminant caruncles are specialized sites for placental development, and both are gland-free endometrial modifications evident in immature animals. Endometrial pads are prominently developed at the time of birth, and Mossman ('37) reported that caruncles are grossly differentiated in female cow fetuses of 40 cm CR.

Prominent bilateral mucosal folds or endometrial cushions project into the uterine lumen of shrews (family Soricidae) with free uterine blastocysts (Hubrecht, 1894a; Brambell and Perry, '45; Wimsatt

-vert signs of epithelial degeneration. A slightly later implantation stage shows degenerating epithelial cells which have been separated from their basement membrane by the invading trophoblast.

Enders ('63, '64) suggested that the initial penetration of the endometrium by the armadillo blastocyst is accomplished by the distintegration of a localized region of the uterine epithelium. After the initial penetration there is little additional epithelial loss. The remaining epithelial cells deteriorate only after the invasive trophoblast separates them from their normal vascular relationships. Preliminary observations on the fine structure of implantation in the hamster suggest that after initial attachment of the blastocyst to the endometrium, trophoblastic cells penetrate between epithelial cells and send horizontal processes which detach the epithelial cells from their underlying basement membrane (Luckett, W. P. and M. W. Orsini, unpublished observations). A similar mechanism is postulated for implantation in the tree shrew.

Mitoses are evident in the cytotrophoblast of the attachment plaques, but not in the giant cells. The latter probably arise by fusion of cytotrophoblastic cells, although there is no direct evidence for this. Neither is there any evidence to support Hubrecht's (1899) contention that giant cells are formed by amitotic division of trophoblastic nuclei. The tupaiid giant cells appear to be involved primarily with penetration of the basement membrane of the uterine epithelium and the formation of cores for the subsequent development of the multinucleated primary villi. The disappearance of giant cells from the main body of the placental discs with the establishment of the chorio-vitelline placentae accounts for the reports that giant cells do not occur in the tree shrew placenta (van der Horst, '49; Meister and Davis, '56, '58). These authors lacked the late implantation stages which are characterized by the prominent zone of giant cells.

The basement membrane of the endometrial pad epithelium appears to form a temporary barrier to trophoblastic invasion, since it persists for a longer time than does the overlying epithelium. It is penetrated only after the differentiation of the layer of multinucleated giant cells. The fine structural studies of implantation in the rabbit (Larsen, '61), rat (Enders and Schlafke, '67) and the hamster (Luckett, W. P. and M. W. Orsini, unpublished observations) also suggest the probability that the basement membrane acts as a temporary barrier to the invasive trophoblast.

Hill ('65) described decidual knot formation in an early stage of chorio-vitelline placentation in *Tupaia minor*. He noted that these masses are formed by the fusion of decidual cells and ". . . are syncytial formations, maternal in origin and destined eventually to be resorbed and so we may adopt for them the designation maternal symplasma or simply symplasma, a term suggested by Bonnet."

The term symplasma does indicate maternal tissue that loses its cellular boundaries and is destined for destruction, but in almost all cases it connotes uterine epithelium that undergoes this characteristic reaction. Symplasmic reaction of epithelium associated with implantation typically occurs in some of the Old and New World monkeys (Hill, '32). Because the syncytial modifications of the tupaiid endometrium involve decidual cells rather than epithelium, and since the reaction occurs during early chorio-vitelline placentation rather than at implantation, the term "decidual knots" is preferred for these masses.

Hubrecht (1899) used the term "trophospongia" to describe the maternal decidual tissue of *Tupaia*. He made no reference to the decidual knots, although they can be seen in several of his figures. Hubrecht (1889) first used the term trophospongia to describe ". . . the product of the further continuation of the endothelial proliferation" during the development of the hedgehog placenta. He believed that this proliferated maternal endothelium or trophospongia conducts maternal blood from the endothelial-lined decidual vessels to the trophoblast-lined lacunae at the fetal surface of the placenta. Thus, trophospongia in the hedgehog refers only to modified maternal endothelial cells and not to decidual cells. Unfortunately, the development of the hedgehog placenta has not been re-examined in detail since Hubrecht's publication.

The occurrence of an expanded bilaminar blastocyst in the ovarian end of the uterine horn in an instance of pregnancy during lactation suggests lactational delay of implantation in tree shrews. It is uncertain from most reports of delayed implantation in other species whether blastocysts are distributed throughout the uterine horn during the delay period or whether they accumulate at the ovarian end of the horn. There is some evidence that blastocysts are distributed throughout the uterine horn during the delay period in both the rat (Krehbiel, '41) and the European badger (Canivenc, '66), but information appears to be lacking for other species.

Hopefully, more information will be obtained about possible lactational delay of implantation in tree shrews from a study of these animals in captivity. It will be interesting to determine whether there is actually a delay in attachment, as appears likely in the rat, or whether there is a delay in distribution of the blastocysts to the optimal sites of implantation. The latter phenomenon is suggested in the single delayed specimen of *Tupaia* available.

The irregular epithelial crypts that develop in the surface epithelium of the endometrium, including that of the endometrial pads, are best seen in *Tupaia longipes*. A few crypts are seen just prior to implantation in *T. javanica*, and there is a suggestion of crypt formation in *T. minor* in a later stage of implantation. The crypts seen in *T. longipes* at the time of implantation appear identical to those of nulliparous *T. chinensis* treated with exogenous estrogen plus progesterone (Conaway and Sorenson, '66). Hubrecht (1899) made no mention of epithelial crypts prior to or at the time of implantation in *T. javanica*, but several of his drawings depict the radial arrangement of epithelial and trophoblastic cells seen in the implantation plaque of *T. minor*. This radal arrangement is best interpreted as the result of interdigitation of trophoblastic and epithelial elements at the time of attachment. An analogous condition of crypt formation occurs in the cat during the second week after ovulation, at the time when blastocysts lie free in the uterine lumen (Courrier and Gros, '32; Dawson and Kosters, '44).

Prominent epithelial thickenings are formed in shrews just prior to implantation on both the antimesometrial and lateral walls of the endometrium (Hubrecht, 1894a; Sansom, '37; Brambell and Perry, '45; Wimsatt and Wislocki, '47). The antimesometrial thickenings give rise to epithelial crypts which mark the site of the future development of the chorio-allantoic placenta. Implantation occurs circumferentially on the lateral epithelial thickenings, but there is no crypt development.

The formation of epithelial crypts on the uterine caruncles prior to the formation of trophoblastic villi occurs in the Bovidae (Björkman, '54) and Cervidae (Hamilton, Harrison and Young, '60). The epithelial crypts are the sites of attachment and invasion of trophoblastic villi; crypts do not form in the intercaruncular regions.

Superficial, central implantation or attachment of an expanded blastocyst which fills most or all of the uterine lumen is characteristic of the Tupaiidae, the insectivorous groups Soricidae, Talpidae and *Chrysochloris*, the order Carnivora, and all of the taxa with an epitheliochorial or syndesmochorial placenta: Lemuroidea, Lorisoidea, Artiodactyla, Perissodactyla, Pholidota and probably Cetacea (Mossman, '37). In all of these groups the initial attachment is either circumferential or bilateral, but only the Tupaiidae, Soricidae, Talpidae and *Chrysochloris* exhibit noticeable endometrial modifications in this region prior to implantation.

There is no evidence to support Hubrecht's (1899) description of the blending together of trophoblast and uterine epithelium at the time of implantation to form a common coalescing plasmodium ("einem gemeinschaftlichen Verschmelzungsplasmodium"). Plasma membranes separate trophoblastic and epithelial cells up until the time that epithelial cells begin to degenerate.

The degeneration of epithelial cells in later stages of implantation appears to be initiated by their separation from the basement membrane by the ameboid activity of the trophoblast rather than by cytolytic activity of the trophoblast. In early implantation stages cytotrophoblastic cells send processes between epithelial cells toward the basement membrane, but there are no

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Hubrecht (1899) described the proliferation of intervascular connective tissue to give rise to trophospongial connective tissue in the tree shrew placenta. His description and figures indicate that trophospongia in this case is maternal decidual tissue. In a later publication ('08) he stated that trophospongia indicates a maternal cell proliferation, but he did not specify whether it is endothelial, stromal or both. The name trophospongia is no longer used to describe maternal tissue, and it is easily confused with the term "trophospongium," which describes a trophoblastic zone that contains maternal blood channels but no fetal capillaries (Orsini, '54).

The loose zone formed by the degenerating decidual knots appears to provide a pathway for the subsequent invasion of primary villi. The reason for the fusion of decidual cells to form knots prior to their degeneration is unknown. Multinucleated primary villi appear to develop around a core of giant cells; they are first evident when avascular mesoderm separates the endoderm and trophoblast of the placental disc. There is no evidence of mitoses in the primary villi, and presumably they differentiate from the cytotrophoblast of the chorionic plate.

Hill ('65) believed that the cytotrophoblast of the chorionic plate becomes transformed into syntrophoblast at the end of the period of chorio-vitelline placentation. However, he reported the occurrence of mitotic figures in this syntrophoblast and admitted that this is an exceptional phenomenon. There are no well-documented cases for mitotic activity occurring in syntrophoblast, and evidence from autoradiography (Richart, '61; Midgley, Pierce, Deneau and Gosling, '63) and electron microscopy (Carter, '64; Pierce, Midgley and Beals, '64; Enders, '65; Boyd and Hamilton, '66) of human and rhesus monkey placentae indicates that syntrophoblast differentiates from cytotrophoblastic cells which are incorporated into the syncytium and subsequently lose their plasmalemmas.

In all stages of chorio-vitelline placentation examined in the present study, the chorionic plate consists of cytotrophoblast which is continuous with the irregular primary villi. Plasma membranes are apparent in structures labeled as "syncytiotrophoblast" in Plate VII of Hill's ('65) paper. In addition, structures labeled as "symplasma" in his Plates IIIa, Vb, and VIc are identical to the multinucleated primary villi described in the present study. One of Hill's figures shows giant cells buried in the cores of two "symplasmic masses" which are really primary villi. The more densely nucleated primary villous processes which have penetrated deeper into the decidual knot zone and can be seen partly surrounding several decidual knots are also labeled as "symplasma" by Hill.

Hubrecht (1899) and van der Horst ('49) have emphasized the difficulties involved in distinguishing between fetal and maternal tissues in the developing chorio-vitelline placenta of *Tupaia*. Nuclear morphology is quite similar in both primary villi and decidual knots, and it is frequently necessary to follow serial sections in order to determine the identity of a particular densely nucleated mass. Hill's ('65) misinterpretation of tissue relationships during chorio-vitelline placentation apparently resulted from the gap in his series of developmental stages. His earliest is an incipient stage of chorio-vitelline placentation, since the bilaminar omphalopleure still covers the major portion of the placental disc. His next stage is transitional between chorio-vitelline and chorio-allantoic placentation. This gap, which covers most of the period of chorio-vitelline placentation, is filled in the present study by nine specimens which consist of all stages of this critical period.

The vascular chorio-vitelline placentae of tupaiids appear to play an important role in fetal-maternal exchange, since they are prominently developed through the early limb bud stage. Maternal capillaries are embedded in the chorionic plate; in many places they are separated from the fetal vitelline capillaries by only 1–2 layers of cytotrophoblast.

Only a single layer of syntrophoblast covers the maternal capillary tubules of the tupaiid labyrinth, whereas other mammalian endotheliochorial or hemochorial placentae are generally characterized by having both cytotrophoblastic and syntrophoblastic layers, at least in the early stages. This cytotrophoblast may be reduced or absent near term, and it is believed to give rise to the syntrophoblast. A

noticeable exception to this pattern is the armadillo placenta, which possesses only a single layer of syntrophoblast in the labyrinth, similar to the tupaiid condition (Enders, '60a,b). Cytotrophoblastic cell columns occur at the tips of the early chorio-allantoic villi in the armadillo and apparently act as a source of proliferation for the syntrophoblast of the villi, since cytotrophoblast does not persist in the chorionic plate.

Chorionic plate cytotrophoblast and the multinucleated cell columns are the most likely sources of syntrophoblast in the tupaiid labyrinth. Mitoses occur in the chorionic plate but not in the cell columns. Both chorionic plate cytotrophoblast and cell columns have disappeared in late stages of pregnancy.

The epithelioid allantoic mesenchymal cells of the tupaiid labyrinth are strikingly similar to those of the shrew placenta (Wimsatt and Wislocki, '47; Mossman and Owers, '63). In both groups these cells differentiate early during the formation of the chorio-allantoic placenta and persist until term. Mossman and Owers ('63) believed that the trophoblast is lost in the labyrinth of the shrew placenta and they classified it as endothelio-endothelial. They suggested that the epithelioid mesenchyme of the shrew placenta is possibly an adaptation to the absence of trophoblast. However, Wimsatt and Wislocki ('47) stated that a thin layer of syntrophoblast persists in the labyrinth, and they classified the shrew placenta as endotheliochorial.

Enders ('60a) reported that the mesenchymal cells of the chorio-allantoic placenta from late pregnant armadillos are rounded, gland-like cells with pronounced cytoplasmic basophilia. The presence of vast amounts of dilated rough endoplasmic reticulum containing moderately electron-dense material within their cisternae and of a well developed Golgi complex suggests that these cells are involved in protein secretion (Enders, '60b). Modified mesenchymal cells are not seen in early stages of armadillo placentation.

The epithelioid mesenchymal cells of the shrew and tree shrew placentae do not appear to be closely related to those of the armadillo. They are prominent from the earliest stages, and their cytoplasm does not exhibit marked basophilia. Histochemical and electron microscopic examinations may provide some evidence of their function.

The persistence of maternal endothelium in the tupaiid labyrinth was previously reported by van der Horst ('49) and Hill ('65). Hubrecht (1899) believed that this endothelium degenerates after it becomes surrounded by syntrophoblast, however, his figures that illustrate this point are not convincing. Maternal capillaries are clearly surrounded by trophoblast in his figure 54; in fact, his stages illustrating "loss" of maternal endothelium are prior to the establishment of the chorio-vitelline placentae. Meister and Davis ('56, '58) believed that the endotheliochorial labyrinth of early stages is converted to the hemochorial type near term, but all of their figures show hypertrophied maternal endothelium adjacent to a thin layer of syntrophoblast. These authors did not attempt to demonstrate the presence of basement membranes by histochemical techniques. Since the three species examined by Hubrecht and Meister and Davis are represented in the present study, the possibility of species differences is ruled out.

The thin incomplete zone of chromatic masses at the base of the placental discs in late stages of pregnancy may represent remnants of both cell columns and decidual knots. The nuclei of both of these cell types become hyperchromatic, and it is impossible to determine whether one or both contribute to the chromatic masses. The basal layer of hyperchromatic and pyknotic nuclei in the placentae of American shrews has been interpreted as syntrophoblast (Wimsatt and Wislocki, '47) or as a symplasmic remnant of uterine gland epithelium (Mossman and Owers, '63). The chromatic masses are strikingly similar in these two families. The possible derivation of these masses from glandular epithelium in soricids and from uterine stroma in tupaiids does not pose a major problem in homology, since both of these tissues are mesodermal derivatives.

Placentae obtained at normal births indicate that separation from the uterus occurs between the zone of basal chromatic masses and the thin zone of unmodified stroma. The present study refutes the con-

tention that the placentae of *Tupaia* degenerate *in situ* after the delivery of the fetus (van Herwerden, '06; Hubrecht, '08). It is possible that the "puerperal" uteri examined by van Herwerden were examples of embryonic resorption. Several uteri with resorbing embryos were seen during the present study, and they show evidence of partial degeneration of the placentae *in situ*.

The appearance of the yolk sac is relatively similar during the early stages of pregnancy in most mammalian orders, but several modifications may occur during later stages of gestation. (1) The yolk sac may become completely vascularized and persist as a relatively large structure, (2) the completely vascularized yolk sac may become extremely reduced or vestigial in later stages, (3) mesoderm and vitelline vessels may never extend to separate the layers of the abembryonic or distal wall of the yolk sac, and the vascular splanchnopleure may then be closely apposed to the persisting bilaminar orphalopleure, resulting in an *incompletely inverted yolk sac*, and (4) a completely inverted yolk sac is formed when the bilaminar omphalopleure is completely lost and the endoderm of the vascular splanchnopleure is directly exposed to the endometrium (Mossman, '37).

A large, free vascular yolk sac occurs in reptiles and birds and probably represents a primitive condition; among eutherian mammals it is retained in the Tupaiidae and the Carnivora. The taxa with a reduced or vestigial vascular yolk sac include the Primates, Artiodactyla, Perissodactyla, Cetacea, Megachiroptera, the family Centetidae of the Insectivora, the family Bradypodidae of the Edentata and several families of Microchiroptera (Mossman, '37; Goetz, '37b; Moghe, '51; Gopalakrishna, '58). Inverted yolk sacs are characteristic of Rodentia, Lagomorpha, most Insectivora and several families of Microchiroptera (Mossman, '37; Wimsatt, '54; Gopalakrishna, '58).

The tupaiid yolk sac is strikingly similar to that of the most primitive family of carnivores, the Viverridae. The free yolk sac of a late stage of pregnancy of *Paradoxurus* occupies a greater cross-sectional area than is usually seen in other carnivores; this appears to be a case of primitive retention (Moghe, '56; personal observations).

Meister and Davis ('56, '58) reported that the allantoic vesicle of *Tupaia* is "large in early pregnancy, becoming small near term; the vesicle does not reach the fetal surface of the placenta at any stage of pregnancy." These authors sectioned one uterus with the fetus *in situ* from an early stage of chorio-allantoic placentation; however, in the three later stages which they studied, only the placental discs appear to have been sectioned. The structure identified as the allantoic vesicle in figure 2 of their 1958 paper bears no resemblance to the allantois seen at any stage of development during the current study. Examination of placental discs alone from later stages provides little information about the nature of the allantoic vesicle, since it is intimately fused with the chorion, amnion and the surface of the placental discs. Examination of serial sections during the present study demonstrates the continuity between urachus, allantoic duct and the *large persisting allantoic* vesicle.

The retention of a large allantoic vesicle in late stages of pregnancy is characteristic of reptiles, birds and all of the mammalian taxt that possess an epitheliochorial placenta: Artiodactyla, Perissodactyla, Pholidota, Cetacea and Lemuroidea. A large allantoic vesicle is also found in the subungulate orders Proboscidea, Sirenia and Hyracoidea, and in many, but not all, of the taxa that have an endotheliochorial placenta: Carnivora, Tubulidentata, Tupaiidae and Talpinae (Mossman, '37, '57). The only known occurrence of a large allantoic vesicle associated with a hemochorial placenta occurs in several families of Insectivora: Centetidae, Potamogalidae and Chrysochloridae (Goetz, '37b; Strauss '43; Hill, '38; Hintzsche, '40).

It has been suggested that the presence of a large allantoic vesicle is correlated with the development of a large functional mesonephric kidney (Bremer, '16). Large and apparently functional mesonephroi are found in birds, reptiles, pigs and sheep and all of these forms possess a large allantois. Both mesonephroi and allantoic vesicles are reduced or vestigial in rodents and the human. The best evidence for the correlation between the functional develop-

ment of the mesonephros and the size of the allantois is presented by Davies ('52) for the sheep. Histochemical and morphological evidence suggests that the sheep mesonephros becomes functional on about the 18th day of gestation, and this appears to be correlated with the sudden expansion of the allantoic vesicle and an accumulation of fluid within it. Although the tree shrew possesses a moderately well developed mesonephros, it is uncertain whether its large allantoic vesicle has a functional significance or whether it is another example of primitive retention.

The small depressions of the smooth chorio-allantoic membrane of *Dendrogale* are probably analogous to the variety of paraplacental absorptive areas that occur in all of the groups that possess an epitheliochorial placenta, as well as in the Carnivora, Talpidae and Tubulidentata (Amoroso, '52; Mossman, '57, '67b). Hill ('65) observed several shallow crypts in the trophoblast of the chorio-vitelline membrane in a single specimen of *Tupaia dorsalis*. He suggested that the crypts are possibly vestiges of the chorionic vesicles found in Talpidae, Lemuroidea and the ungulates. The rudimentary development and scarcity of these structures observed in the present study support this suggestion. The phagocytosis of red blood cells in *Dendrogale* by the trophoblast of the vestigial chorionic vesicles and the smooth chorio-allantoic membrane in general supports the hypothesis of Sinha and Mossman ('66) that the placental hematomes characteristic of carnivores are derived from specialized chorionic vesicles or areolae.

Uterine glands are secretory throughout pregnancy in tupaiids and in most taxa that possess an epitheliochorial or endotheliochorial placenta; this "histotrophe" appears to play an important role in fetal nutrition (Amoroso, '52).

Macrophages containing hemosiderin, an iron-containing compound related to ferritin, accumulate at the sites of previous placentae in many mammals to form so-called "placental scars" (Momberg and Conaway, '56). The similarity of the supranuclear granules in the cytoplasm of the glandular epithelium to the hemosiderin granules of the macrophages, and the positive iron test for both suggest the possibility that hemosiderin is transferred from macrophages to the glandular epithelium in tree shrews. Ferritin does not normally penetrate the cell plasmalemma, but there is good evidence that under both normal and experimental conditions it can enter cells by the process of micropinocytosis (Fawcett, '65). It is quite possible that hemosiderin enters the basal glandular epithelial cells by a similar mechanism.

The occurrence of iron in the apical cytoplasm of the basal uterine glands during pregnancy has been reported in the rat, guinea pig, pig and woman (Wislocki and Dempsey, '45). These authors believed that the presence of iron in the uterine glands is a consequence of pregnancy, since tests for iron are negative in nonpregnant specimens. In contrast, Wislocki and Dempsey were unable to detect iron in the uterine glands of the cat at any time. They attributed this to the presence of placental hematomes in cats and other carnivores as specialized sites of iron uptake from extravasated maternal red blood cells. This hypothesis should be tested on a variety of other mammals, both with and without placental hematomes.

De Lange and Nierstrasz's ('32) report that amniotic folds first appear in the 4–6 somite stage is confirmed by the present study. However, they believed that the amniotic head fold is initially a proamnion, that is, formed by ectoderm and endoderm but lacking mesoderm, similar to the condition in the early avian embryo. They noted the complete closure of the amniotic folds in the 23 somite stage (corresponding to the time when the vascular allantois has reached the wall of the chorion), although the proamnion still covered a considerable portion of the anterior trunk region. There is no evidence for a proamnion in any stage of amniogenesis in the present study. In all specimens the amniotic folds are composed of an ectodermal and mesodermal layer.

De Lange and Nierstrasz ('32) described the formation of an "archamnionic cavity" in the inner cell mass of *Tupaia javanica* prior to implantation, but similar stages were not available for the present study. This cavity in the inner cell mass is formed by a separation of cells; the subsequent

thinning of both Rauber's layer and the roof of the archamniotic cavity results in its opening into the uterine lumen and the flattening of the cup-shaped disc to form the embryonic shield. Archamniotic formation and loss occur prior to implantation, whereas the formation of the true amnion by folding occurs in the 4–6 somite stage. A similar process of archamnion formation and rupture appears to occur also in the musk shrew *Crocidura caerulea* (= *Suncus murinus*) (Sansom, '37), moles (Goetz, '38; Prasad, '58) and in ungulates and carnivores (Goetz, '38). In all of these forms the true amnion is formed by folding.

Although experimental evidence is lacking, the fetal membrane morphogenesis of the tupaiids suggests the possibility of a variety of regions for feto-maternal exchange that frequently overlap temporally. Uterine gland secretion appears to play an important nutritive role from the preimplantation through the early chorio-vitelline placental phases. Secretory activity is reduced during chorio-allantoic placentation, but the increased vascularity and modification of the mid-mesometrial region of the smooth chorio-vitelline membrane suggest that it is involved in the uptake of uterine secretion. Iron appears to be supplied to the fetus in part by uterine gland secretion. The antimesometrial smooth chorio-allantoic membrane is capable of phagocytizing RBC's and is probably involved in absorption of uterine gland secretion. The chorio-vitelline placentae are well developed through the early limb bud stage, and the proximity of fetal and maternal capillaries suggests an efficient exchange mechanism. The degenerating decidual knot zone may provide material for phagocytosis by the trophoblast during the period of chorio-vitelline placentation. The chorio-allantoic placentae replace the chorio-vitelline placentae both topographically and functionally.

The renewed interest in the phylogenetic relationships of the Tupaiidae warrants a detailed re-evaluation of the phylogenetic implications of the tupaiid fetal membranes on a broad comparative basis, and this is the subject of a paper now in preparation.

Synopsis of Basic Data on the Fetal Membranes of the Tree Shrews *Tupaia, Lyonogale, Urogale, Dendrogale, Anathana*

Implantation:
- Orientation (disc): Antimesometrial
- Orientation (first attachment): Bilateral
- Depth: Superficial

Decidua: Basalis forms "decidual knots," no capsularis

Amniogenesis: "Archamnion" formed by cavitation prior to implantation; true amnion formed by folding

Yolk sac:
- Bilaminar omphalopleure: Temporary
- Chorio-vitelline placentae: Bilateral; well developed until early limb bud stage
- Vascular splanchnopleure: Large, free vascular yolk sac; no inversion of germ layers

Chorio-allantoic Placentae:
- Shape: Discoidal
- Type: Labyrinthine
- Finer morphology: Endotheliochorial
- Location: Bilateral

Allantoic vesicle: Large: permanent

Authorities: Hubrecht, 1899; van der Horst, 1949; Hill, 1965; Verma, 1965; Luckett, 1968

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The mesometrium is toward the bottom of all figures unless otherwise indicated.

PLATE 1

EXPLANATION OF FIGURES

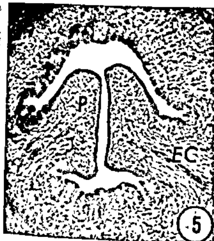
5 Cross section of uterine horn at 12 days postnatum. Note the prominent bilateral endometrial pads and the early glandular development in the mesometrial and antimesometrial regions. *T. longipes* 43. × 100.

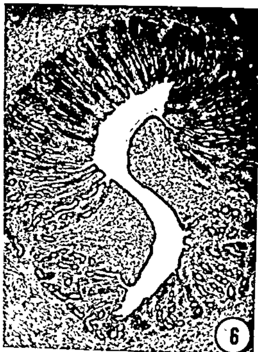
6 Cross section of uterine horn of immature animal. The increased thickness of the glandular endometrium causes the gland-free pads to appear less prominent. *T. minor* 46. × 63.

7 Cross section of uterine horn during tubal blastocyst stage. Gomori's basic fuchsin stain shows the coiled arterioles beneath the pads. The endometrial pad, delimited by arrows, contains no coiled arterioles. *U. everetti* 3. × 63.

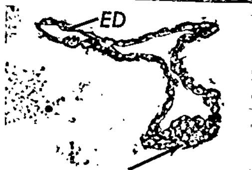
8 Expanding bilaminar blastocyst unoriented in the uterine lumen. Rauber's layer (arrow) is complete over the embryonic knot. *T. glis* 78. × 250.

9 Expanded bilaminar blastocyst near tubo-uterine junction. The delay in distribution of the blastocyst is presumably due to concurrent lactation. The mesometrium is toward the right. *T. longipes* 60. × 100.

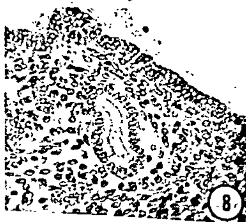

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The mesometrium is toward the bottom of all figures unless otherwise indicated.

PLATE 1

EXPLANATION OF FIGURES

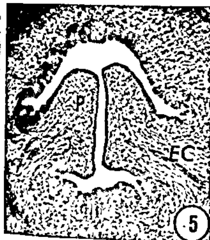
5 Cross section of uterine horn at 12 days postnatum. Note the prominent bilateral endometrial pads and the early glandular development in the mesometrial and antimesometrial regions. *T. longipes* 43. × 100.

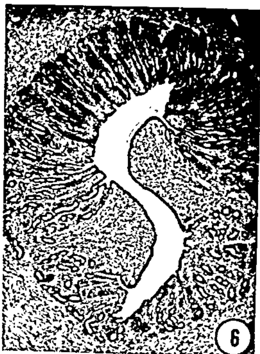
6 Cross section of uterine horn of immature animal. The increased thickness of the glandular endometrium causes the gland-free pads to appear less prominent. *T. minor* 46. × 63.

7 Cross section of uterine horn during tubal blastocyst stage. Gomori's basic fuchsin stain shows the coiled arterioles beneath the pads. The endometrial pad, delimited by arrows, contains no coiled arterioles. *U. everetti* 3. × 63.

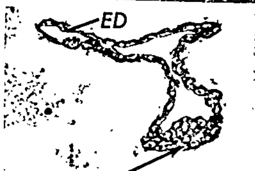
8 Expanding bilaminar blastocyst unoriented in the uterine lumen. Rauber's layer (arrow) is complete over the embryonic knot. *T. glis* 78. × 250.

9 Expanded bilaminar blastocyst near tubo-uterine junction. The delay in distribution of the blastocyst is presumably due to concurrent lactation. The mesometrium is toward the right. *T. longipes* 60. × 100.


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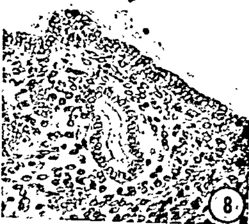

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PLATE 2

EXPLANATION OF FIGURES

10 Early attachment phase of implantation. The intact uterine epithelium is closely apposed to the trophoblast, and the two cell types may be distinguished by their nuclear morphology. *T. picta* 53. × 660.

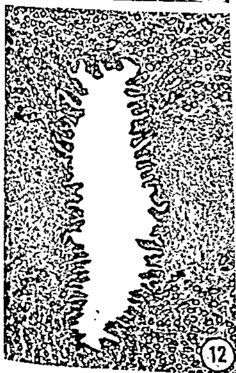
11 Early stage of implantation. Epithelial crypts occur on the endometrial pad and they are partially invaded by the attached trophoblast. *T. longipes* 82. × 400.

12 Uterine horn of early implantation stage, cranial to the attachment site. Epithelial crypts are well developed on both the glandular endometrium and the endometrial pads. The uterine glands are maximally coiled at this period. *T. longipes* 82. × 63.

13 Attachment plaque during later stage of implantation. Some trophoblastic cells have insinuated between epithelial cells to reach the underlying basement membrane. Endometrial stromal cells are enlarging to form decidual cells (arrow). *T. minor* 50. × 560.


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PLATE 3

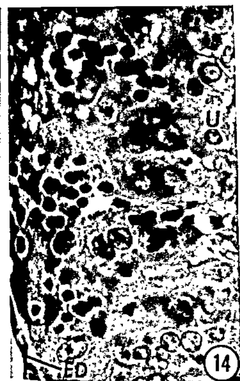
EXPLANATION OF FIGURES

14 Further development of the attachment plaque. The numerous pyknotic nuclei are thought to be remnants of degenerating epithelial cells. Occasional apparently normal epithelial cells are evident. The endodermal lining of the bilaminar yolk sac overlies the attachment plaque. *T. minor* 51. × 560.

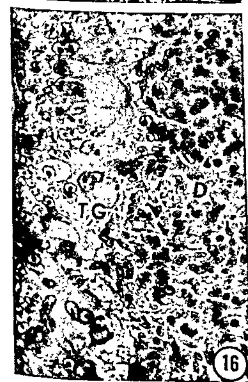
15 Cross section of endometrial pad during early attachment phase of implantation. Note the radiation of capillaries from the arteriole at the base of the pad. The mesometrium is toward the left. *T. picta* 53. × 100.

16 Period of differentiation of the attachment plaque. The trophoblast has differentiated into an inner cytotrophoblastic layer and an outer zone of multinucleated giant cells. The underlying stroma has differentiated into typical decidual tissue. *U. everetti* 4. × 250.

17 Differentiated attachment plaque. Silver azan staining demonstrates the continuity of the basement membrane that underlies the uterine epithelium with the basement membrane that separates the trophoblastic giant cells from the decidua. *U. everetti* 4. ×250.


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PLATE 4

EXPLANATION OF FIGURES

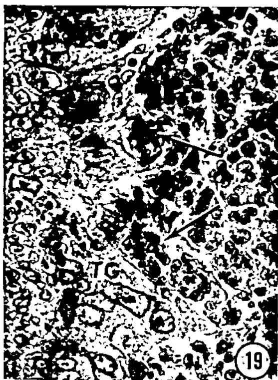
18 Marginal zone of attachment plaque. Trophoblastic giant cells have detached the uterine epithelium from its basement membrane. *U. everetti* 4. × 250.

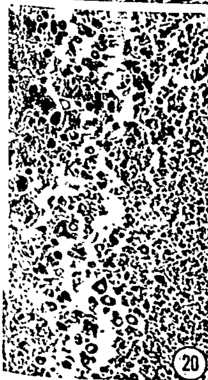
19 Deciduo-trophoblastic junction of late stage of implantation. The decidual cells (arrows) adjacent to the giant cells show increased cytoplasmic basophilia and hyperchromatic nuclei. *U. everetti* 4. × 400.

20 Incipient placental disc covered by avascular trilaminar omphalopleure. Three characteristic regions are seen in cross section: (1) trophoblastic zone, (2) the zone of degenerating decidual knots, and (3) the basal decidua. *T. palawanensis* 46. × 110.

21 Higher magnification near fetal surface of incipient placental disc. Irregular primary villi penetrate into the loose zone of necrotic decidual knots. Note the intact maternal capillaries surrounded by trophoblast. *T. palawanensis*. 46. × 250.


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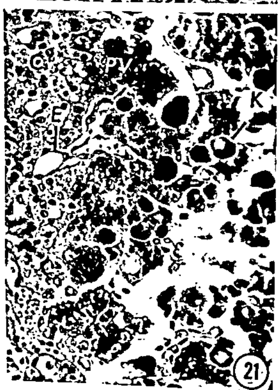

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PLATE 5

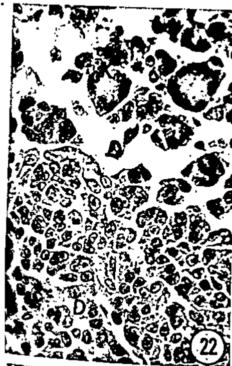
EXPLANATION OF FIGURES

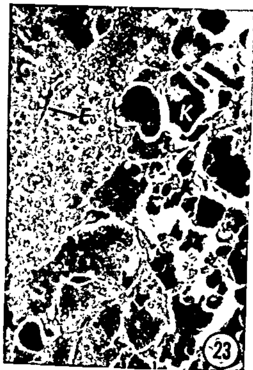
22 Junctional zone of decidua basalis and decidual knots in incipient placental disc. The decidual knots in cross section are characterized by a peripheral ring of nuclei and a central, common cytoplasmic core. Transitional stages occur between the basal decidual cells and the decidual knots. The mesometrium is toward the left. *T. palawanensis* 46. × 400.

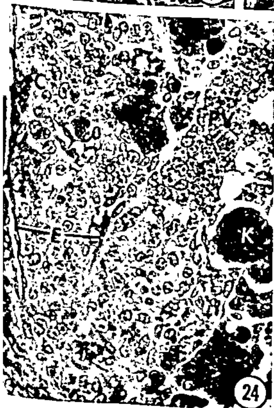
23 Near fetal surface of chorio-vitelline placenta. Primary villi are continuous with the cytotrophoblast. Some of the decidual knots show signs of degeneration. *T. glis* 76. × 250.

24 Higher magnification of a primary villus. Note the intact endothelium of the maternal capillaries surrounded by cytotrophoblast. *T. glis* 76. × 400.

25 Vascular chorio-vitelline placental disc. The interdigitated zone of primary villi and the decidual knots occupies about a third of the thickness of the disc. *T. javanica* 2. × 38.


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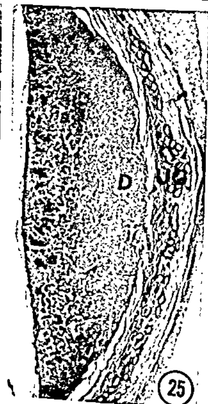

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PLATE 6

EXPLANATION OF FIGURES

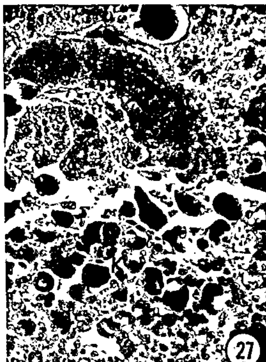
26 Paraplacental region of chorio-vitelline placental disc. Prominent trophoblastic giant cells detach the uterine epithelium in this region. *T. glis* 76. × 325.

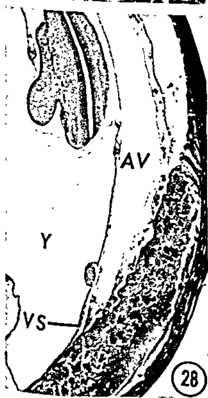
27 Deciduo-trophoblastic junction of chorio-vitelline placental disc. The densely nucleated primary villi are closely associated with degenerating decidual knots. The mesometrium is toward the left. *T. glis* 76. × 275.

28 Initiation of chorio-allantoic placentation. The vascular allantois is detaching the antimesometrial portion of the vascular splanchnopleure of the yolk sac from the surface of the placental disc. *T. javanica* 6. × 28.

29 Higher magnification of end of allantoic vesicle shown in fig. 28. Yolk sac splanchnopleure still covers the mesometrial portion of the placental disc. Vascular allantoic mesenchyme is beginning to indent the cytotrophoblast of the surface of the placental disc. *T. javanica* 6. × 110.


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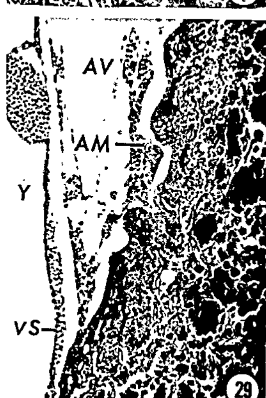

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PLATE 7

EXPLANATION OF FIGURES

30 Early stage of chorio-allantoic placentation. Four zones are evident at this stage: (1) chorionic plate, (2) labyrinth, (3) the dense zone of primary villi and decidual knots, and (4) the decidua basalis. *T. javanica* 7. × 100.

31 Base of the placental disc during early chorio-allantoic placentation. Densely nucleated primary villi interdigitate with the decidua basalis. Scattered decidual knots are evident. *T. javanica* 7. × 250.

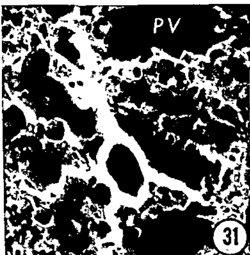
32 Fetal surface of the placental disc during early chorio-allantoic placentation. Note the continuity between the cytotrophoblast of the chorionic plate and the syntrophoblast of the labyrinth. Shrinkage has separated the syntrophoblast from the allantoic mesenchyme. The mesometrium is toward the left. *T. javanica* 7. × 350.

33 Fetal surface of disc during early chorio-allantoic placentation. Note the persistence and hypertrophy of the maternal capillary endothelium adjacent to the chorionic plate cytotrophoblast and to the syntrophoblast of the labyrinth. The mesometrium is toward the left. *T. javanica* 161. × 400.

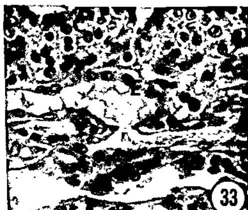
34 Base of the placental disc of a stage later than that of figure 31. The densely multinucleated masses at the base of the disc are probably remnants of the trophoblastic cell columns. Note the accumulation of secretory product in the uterine glands beneath the disc. *T. javanica* 5. × 100.

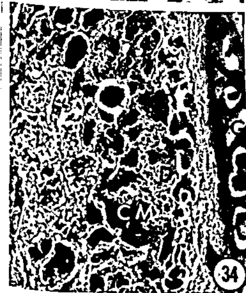
35 Base of placental disc from definitive chorio-allantoic placentation. Basal chromatic masses may represent remnants of cell columns or decidual knots. *T. montana* 62. × 300.


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PLATE 8

EXPLANATION OF FIGURES

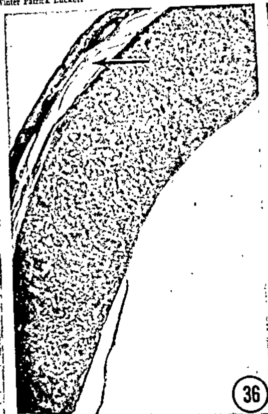
36 Cross section of placental disc from near term pregnancy. Most of the disc is occupied by the labyrinth. Note that the disc separates easily from the underlying stroma immediately beneath the incomplete layer of basal chromatic masses. *T. montana* 62. × 25.

37 Labyrinth of near term chorio-allantoic placenta. Note the epithelioid appearance of the allantoic mesenchymal cells. A thin layer of syntrophoblast surrounds the modified endothelium of the maternal capillaries. *T. palawanensis* 68. × 250.

38 Labyrinth of near term chorio-allantoic placenta. The interconnection of the trophoblastic lamellae demonstrates the true labyrinthine nature of the tupaiid placenta. *T. montana* 62. × 250.

39 Labyrinth of near term chorio-allantoic placenta. A PAS-positive basement membrane separates the syntrophoblast of the labyrinth from the maternal capillary endothelium. *T. gracilis* 26. × 672.

PLACENTATION OF THE TREE SHREWS
Winter Patrick Luckett



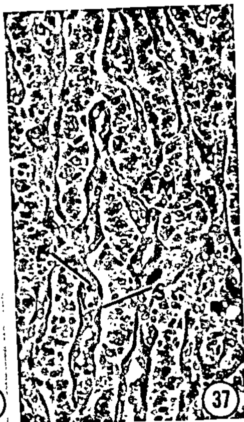






PLATE 8

EXPLANATION OF FIGURES

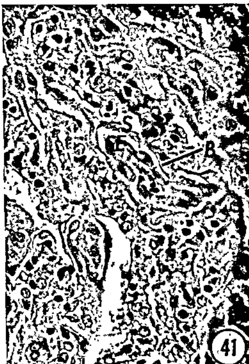
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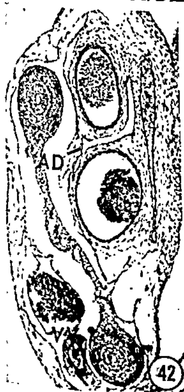
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PLATE 9

EXPLANATION OF FIGURES

40 Labyrinth of near term chorio-allantoic placenta. Azan silver staining demonstrates a thin, incomplete layer of reticular fibers separating syntrophoblast and maternal endothelium. More prominent reticular fibers separate allantoic mesenchyme and syntrophoblast. *U. everetti* 1. × 440.

41 Labyrinth of placenta obtained from normal term birth. The hypertrophied maternal endothelium is separated from the syntrophoblast by a hyalinized basement membrane. *T. chinensis* 45. × 400.

42 Umbilical cord during early chorio-allantoic placentation. The allantoic arteries flank the compressed allantoic duct. The intestinal loop and vitelline vessels lie in the umbilical coelom. *T. javanica* 5. × 63.

43 Distal remnant of vitelline duct during early chorio-allantoic placentation. The vitelline duct (arrow) is flanked by the primary branches of the vitelline vein. *T. javanica* 7. × 76.

PLATE 10

EXPLANATION OF FIGURES

44 Mid-mesometrial region of smooth chorio-vitelline membrane during chorio-allantoic placentation. Note the increased thickness and vascularity of the distal endoderm as compared to the proximal endoderm. The thickened distal wall of the yolk sac is fused with the overlying chorion to form the chorio-vitelline membrane. The mesometrium is toward the right. *U. everetti* 90. × 130.

45 Mid-mesometrial modification of the chorio-vitelline membrane. The trophoblast of the invaginated chorion surrounds a pocket that is continuous (arrow) with the uterine lumen. The mesometrium is toward the right. *T. palawanensis* 68. × 100.

46 Smooth chorio-allantoic membrane. The trophoblast forms several shallow invaginations (arrow), and it is phagocytizing maternal RBC's that have escaped into the uterine lumen. The mesometrium is toward the left. *D. murinus* 93. × 400.

47 Uterine glands during implantation stage. Macrophages containing hemosiderin are scattered in the interglandular connective tissue. The supranuclear cytoplasm of the glandular epithelium contains numerous fine granules that resemble hemosiderin in their color and refractivity. *U. everetti* 4. × 1000.

48 Uterine glands during early chorio-allantoic placentation. Turnbull's blue stain demonstrates the presence of iron in the interglandular macrophages and in the cytoplasm of the glandular epithelium. *U. everetti* 90. × 400.

Cortisone-induced Hypertension and Cardiovascular Lesions in Mice [1]

THOMAS D. CLARKE,[2] ALLEN D. ASHBURN AND W. LANE WILLIAMS
Department of Anatomy, University of Mississippi School of Medicine, Jackson, Mississippi

ABSTRACT Daily subcutaneous injections of cortisone acetate (0.5, 1.5 or 2.5 mg) were given to three groups of mice for seven consecutive days. Daily systolic blood pressures of the anesthetized mice were obtained by adapting the method of Friedman and Freed ('49). The maximal arterial pressure increase for the 0.5, 1.5 and 2.5 mg groups was 22%, 31% and 41% respectively. This supports the conclusion that cortisone produces hypertension in mice when administered in large doses. Mural hyalinization, vacuolization and cellular proliferation of coronary arteries were greatest in the 0.5 mg group. The highest incidence of myocardial necrosis, 56%, was in the group receiving 2.5 mg of cortisone daily. The frequency and severity of myocardial and renal cortical necrosis were directly related to the size of the cortisone dose. Adrenal medullary vacuolization and lipid infiltration of the liver were common in all experimental groups.

In mice which received large amounts (2.5 mg) of cortisone for seven days the arterial walls were normal and there were no periarterial reactions except in arteries adjacent to myocardial abscesses (Ashburn, Williams and Arlander, '62). The coronary arteries were normal and periarteritis was absent when formation of such abscesses was prevented by treatment with penicillin (Ashburn and Williams, '66).

In a study of sequential responses to a single large dose of cortisone medial hyalinization of coronary arteries occurred within 24 hours and exudative and proliferative changes in the arterial walls and perivascular spaces followed hyalinization (Ashburn, Williams and Smith-Vaniz, '68).

The above results indicate that the initial injection of cortisone produces an arterial lesion which is masked when cortisone is continued but is allowed to develop fully if administration of cortisone is terminated after the initial injection.

Other studies show that in rats cortisone may increase blood pressure with (Andreichev, '64) or without demonstrable cardiovascular injury (Crane and Ingle, '64). In this study the actions of a range of cortisone doses upon blood pressure in mice were determined and correlated with the amount and type of cardiovascular injury.

MATERIALS AND METHODS

Animals. A total of 245 adult nulliparous albino mice of the Taconic Swiss (TS) strain were randomly assigned to seven experimental groups. Animals were three to four months old and weighed 28–32 gm. Water and a nutritionally balanced commercial diet (Purina Laboratory Chow) were available *ad libitum*. The regimen included caudal blood pressure measurements for ten consecutive days at 24 hour intervals. Four experimental groups received daily subcutaneous injections of cortisone acetate (0.5, 1.5 or 2.5 mg) or isotonic saline solution (0.1 ml) immediately following pressure measurements on days three through nine. The final injection of cortisone or saline occurred 24 hours before autopsy. Weights were recorded daily immediately prior to blood pressure measurement.

To correlate the onset, development and incidence of lesions with levels of blood pressure an additional three groups (105 mice) were injected daily with cortisone (0.5, 1.5 or 2.5 mg). Fifteen of these ani-

[1] Supported by grants from the National Institute of Health, U.S.P.H.S., HE 04052.
[2] Predoctoral Trainee in the Anatomical Sciences (5T1 GM 287).

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mals were killed daily on days 4 through 10. Blood pressure determinations were monitored only from those mice killed on a given day.

Blood pressure determination. An auscultatory monitor (Carter Electronics) was employed for the caudal determination of systolic blood pressure. The procedure is based upon microphonic detection of low frequency vibrations which first occur when a pneumatic occlusion cuff is deflated to a pressure less than that of the caudal artery. The mice were deeply anesthetized with intraperitoneal sodium pentobarbital, 2.5 mg per mouse. To obviate hypervolemic-induced hypertension, the volume of the injection was 0.05 ml. The anesthetized mice were placed for ten minutes in a wooden 22 × 22 × 36 cm warming box at 37°C to produce caudal vasodilation. Mice were subsequently transferred to a larger container (ambient temperature 36–37°C) containing the occlusion cuff and microphone. The 6 mm circular occlusion cuff containing a latex diaphragm was placed to invest the base of the tail but not abut upon the body of the mouse. The rubber diaphragm liner could occlude the lumen of the cuff at 20 mm Hg and yet sustain inflation pressures over 250 mm Hg. The cuff design followed specifications of human sphygmomanometry. The carbon microphone was placed distal to the cuff but not touching it (about 2 cm from the tail root) in order to eliminate noise caused by respiratory and adventitious body movements. Several minutes were allowed to elapse until the mouse was sufficiently immobile. With the cuff completely deflated pulsations of uniform regularity were detected on an oscilloscope. By an inflation bulb the pressure in the occlusion cuff was increased manually to approximately 50 mm Hg higher than the recorded systolic pressure of the previous day. When pressure in the pneumatic cuff decreased (about 2 mm Hg per second) to the systolic level, a persistent pulsatile contour began abruptly (fig. 1). The respective systolic pressure was read with a Tycos aneroid manometer. A permanent record was charted by an oscillograph (Grass 50 with a PIK preamplifier). A precise determination of diastolic pressure was not feasib[le] because of the gradual intensity change o[f] the recorded wave. For each mouse thre[e] consecutive pressure determinations wer[e] made three to five minutes apart. Whe[n] they agreed within a range of 10 mm Hg the average of the three was taken as th[e] systolic pressure.

To attest the adequacy of the indirec[t] pressure recording system 12 mice (thre[e] from each experimental group) were can[-]nulized in the carotid artery with a 2[?] gauge needle which was connected wit[h] 0.55 mm I.D. polypropylene tubing to [a] low volume displacement P23Dc Statham gauge previously calibrated with a one le[g] mercury manometer. The results wer[e] charted so that 1 mm scale deflectio[n] equalled 5 mm Hg pressure (fig. 1).

Statistics. Blood pressures and weight[s] of all mice were normalized to day one. [A] regression line showing the correlation b[e]tween the indirect caudal and the di[rect] carotid arterial pressure was fitted by th[e] method of least squares. Seven mice die[d] during the experiment. These were ex[-]cluded from the statistical analyses b[e]cause of their abnormally low blood [pres]sure and precipitous weight loss prior t[o] death.

Histological technics. These w[ere] standard procedures that have been de[-]scribed before (Ashburn, Williams an[d] Arlander, '62). Paraffin-embedded speci[-]mens of heart, liver, kidneys, adrena[ls,] lungs and aorta were sectioned (minimu[m] of 12 sections) at 6 μ. Sections [were] stained with hematoxylin and eosin (H & E) and periodic acid-Schiff plus h[ema]toxylin (PASH), with or without prior di[a]stase hydrolysis (PASD). Selected section[s] were also stained by Masson's trichrom[e] (McManus and Mowry, '60).

RESULTS

Blood pressure

Prior to injection with cortisone or [sa]line the mice had a three day mean sy[s]tolic blood pressure of 102 mm Hg. Wh[en] systolic pressures of all mice were norma[l]ized to day one, variable gains in exce[ss] of the control group were noted in thr[ee] groups of mice administered cort[isone] (fig. 2). The first statistically signifi[cant]

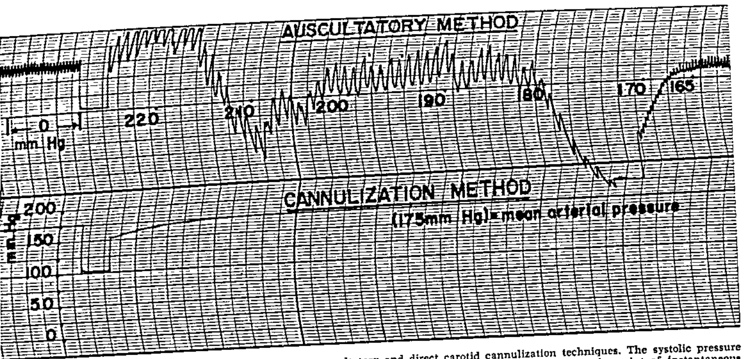


Fig. 1 Comparison of blood pressure values by indirect auscultatory and direct carotid cannulization techniques. The systolic pressure by auscultatory determination is 171 mm Hg. The pressure is equilibrated with the occlusion cuff pressure at the point of instantaneous increase in the frequency and intensity of amplified vibrations. Pulsations preceding the systolic level are due to respiratory and adventitious sounds. Pressure value for the cannulization method represents mean arterial pressure. Paper speed 10 mm/sec.

difference ($p < 0.01$) in blood pressure between the cortisone and saline groups occurred 24 hours after cortisone administration. The most dramatic response was the 41% rise in arterial pressure of mice injected daily with 2.5 mg of cortisone. After the peak on day four the mean systolic blood pressure of the 2.5 mg group decreased significantly below the saline control group ($p < 0.05$) but not below the norm. There was a sustained plateau of blood pressure in the 1.5 mg group; however, the 0.5 mg animals did not differ significantly from those of the controls ($p > 0.05$) for the last three days of the regimen. Blood pressures of the 105 mice killed daily did not differ significantly from the corresponding group receiving the same dose of cortisone.

When the auscultatory technique was validated by direct arterial measurement, a scatter diagram, with the indirect auscultatory pressure as abscissa and the direct carotid cannulization pressure as ordinate, showed a linear relationship between the two variables. The calculated regression line was $y = 112.2 + 1.14x$. The standard error of estimate (SEE) was 18.2. Correlation of pressures in excess of 190 mm Hg were excluded because of incompatibility.

Changes in body weight. Progressive decreases in mean weight after seven days of cortisone or isotonic saline administration were −16% (2.5 mg), −15% (0.5 mg), −11% (1.5 mg) and −4% (saline control). A statistically significant difference in weight loss between all cortisone and saline control groups ($p < 0.01$) existed at least from day seven the end of the experiment.

Changes in organs

No lesions were observed in the saline injected control mice. This stock (Taconi

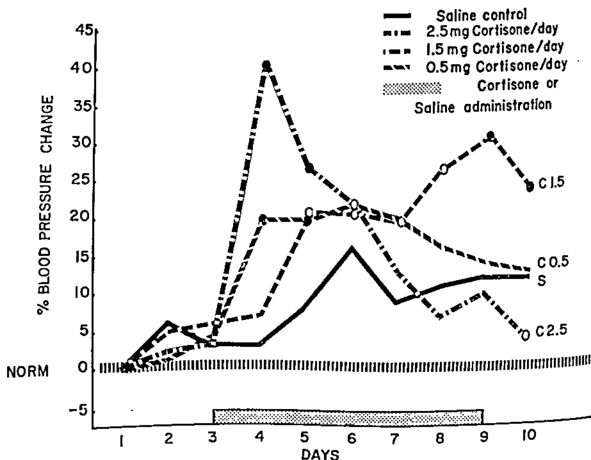


Fig. 2 Percentage shift in mean systolic blood pressure from norm. —●— $p < 0.01$ as compared to saline controls (t-test). —O— $p < 0.05$ as compared to saline controls (t-test).

TABLE 1

Incidence (%) of lesions in mice receiving 0.5, 1.5 or 2.5 mg of cortisone daily for seven days

Types of lesions	0.5	1.5	2.5
	mg/day	*mg/day*	*mg/day*
Myocardial necrosis	20	55	56
(a) Right ventricle	20	55	53
(b) Left ventricle	0	25	28
(c) Atrial	0	0	8
Hyalinization of coronary arteries	55	30	2
Adrenal medullary vacuolization	59	31	30

iss) of mice is free of spontaneous dis- (Ashburn, Williams and Arlander,).

earts. Foci of myocardial necrosis observed in all cortisone-injected (table 1). The right ventricular wall, ventricular septum and subendocar- layers of both ventricles were major of lesions. Atrial necrosis was infre-

Myocardial necrosis was characterized primary degeneration of the sarcoplasm condensation and intense pyknosis of lei. The fibers were shrunken and in- ly PAS-positive (fig. 3). Use of dia- se demonstrated that the PAS-positivity s not due to polysaccharide. Occasion- leukocytes invaded the necrotic areas. ocardial necrosis was first observed 24, and 72 hours respectively after the first ly injection of 2.5, 1.5 or 0.5 mg of cor- one. The highest incidence was on the th day.

Coronary arteries. In mice receiving mg of cortisone arterial lesions were after the second injection. Nu- rous arteries, inconsistently adjacent to eas of myocardial necrosis, contained a non-granular, homogeneous sub- which was extremely PAS-positive stained red with Masson's trichrome. alinized arteries were more frequent in e right ventricle. In the proximal por- ns of arteries hyalinization was often (fig. 4), involving only a part of the sel wall. Hyalin was primarily within The condition resembled "acute ve arteritis" (Karsner, '47). In arently more advanced lesions and in al portions of arteries the entire media frequently hyalinized. Arterial walls often thickened by the hyalin even though the vessels were dilated. Endothelial cells when present appeared swollen and nuclei were often pyknotic.

Adventitial and periadventitial fibrosis of small arteries (fig. 5) occurred within 72 hours after cortisone administration (0.5 mg). A *perivascular response* consisting of fibroblasts, macrophages and mononuclear inflammatory cells was frequent. These proliferative changes involved the media by the sixth day. The sarcoplasm of smooth muscle fibers was swollen giving an impression of medial vacuolization. The smooth muscle nuclei were pyknotic and the cellular membranes were more PAS-positive. These changes appeared to be sequels to hyalinization.

Beginning on the fourth day in the groups receiving 2.5 mg of cortisone a significant number of small coronary arteries showed desquamation of endothelial cells, fragmentation of the elastica, atrophy and necrosis of medial smooth muscle *plus initial* thrombosis (fig. 6). These arterial lesions lacked a mural or perivascular inflammatory response. Hyalinization was infrequent in these severely damaged arteries. Myocardial veins were normal except for dilatation.

Kidneys. Many kidneys had a mottled appearance. Necrosis of the cortex was more prevalent than in the medulla. Medullary hemorrhage and tubular ectasia with degeneration of the epithelial lining developed in conjunction with widespread cortical necrosis. A plasma exudate (fig. 7) occurred within Bowman's space and in the convoluted tubules in those mice which sustained for three days or more a blood pressure in excess of 190 mm Hg. Characteristically in all *cortisone-injected mice* glomerular capillaries showed moderate

distention and the mesangium appeared hypercellular. Tuft adhesion or partial to complete glomerular hyalinization was observed in 11% of the 2.5 mg group.

Adrenal glands. Lipid vacuolization of the outer perimeter of the adrenal medulla (fig. 8) was observed in all cortisone groups but was more prevalent in mice receiving the minimal dose (table 1). The cells of the medullary core appeared normal. There was no necrosis.

Livers. The incidence of necrosis was directly related to the cortisone dose, i.e., 32% (2.5 mg), 30% (1.5 mg) and 15% (0.5 mg). Lipid infiltration of the liver was common to at least 45% of all groups. The incidence and severity of liposis increased directly with the size dose of cortisone (84% in 2.5 mg group).

Lungs. Pulmonary congestion, brochiectasis and diffuse pneumonia were observed in approximately half of the animals but could not be correlated with the dose of cortisone or to changes in other organs.

Other organs. Thymic and splenic involution were frequent. There were no changes in skeletal muscle other than a slightly increased glycogen deposition. Aortas, mesenteric and pancreatic arteries were normal. Splenic vessels, especially the penicillar arterioles, were frequently hyalinized in control and cortisone-injected animals.

DISCUSSION

The reported actions of cortisone on blood pressure are contradictory. Immediate changes of blood pressure in rats, dogs and man are insignificant, but hypertension develops following prolonged administration of large amounts of glucocorticoids (Ingle and Young, '62; Dahl, Heine and Tassinari, '65; Ogawa, Tadokoro, Kirihara, Kawakami and Shibata, '65). In rats daily injections of cortisone produced arterial hypertension within two weeks (Sala, Benati, Amira, Ciceri and Cavallero, '51; Bertazzoli, Cavallero and Sala, '52;



Figures 3–4

Fig. 3 Myocardial necrosis. Dark fibers are necrotic and PAS-positive. (2.5 mg of cortisone for 7 days) PAS plus hematoxylin. × 400.

Fig. 4 Incomplete medial hyalinization of dilated coronary artery. Adjacent myocardial fibers are normal. (0.5 mg of cortisone for 2 days) PAS plus hematoxylin. × 500.

owlton, Loeb, Stoerk, White and Heffern, '52; Andreichev, '65) or elevated hy-ensive pressures to normal (Freed and George, '59; Freed, '61).

Here cortisone produced hypertension in ice very rapidly. The acute rise in blood was approximately reciprocal to dose. Prolonged administration does appear to be a requisite as in rats and Young, '62; Ogawa, Tadokoro, ra, Kawakami and Shibata, '65). The is obscure. Steroid-induced hy- in other animals usually is ited to alterations in electrolytes or increased peripheral resistance, cardiac or blood viscosity (Cosgriff, Diefen- Vogt, '50; Freed, '61; Andreichev, ; Sambhi, Weil and Udhoji, '65).

The blood pressures of control mice in study are in close agreement with reported by other investigators. Di- and inconsistent results may be ned, however, if the occlusion cuff , ambient temperature, level of hesia, caudal pressure gradient, en- and population pressure or age d strain of mice are varied (McMaster, Henry, Meehan, Stephens and Santiban, '65). The dose of pentobarbital large but it has been used before with- t complications (Wu and Visscher, '48). under deep anesthesia have less sys-ic pressure variation than those lightly unanesthetized (McMaster, '41). Although the anesthesia may have lowered stolic blood pressures on a given day by least 10 mm Hg (McMaster, '41; Edrds and Reinecke, '53; Henry, Meehan, ephens and Santisteban, '65), there was gradual rise of pressures in control animals. This may be attributed to an agressive response of the mice to the experimental conditions. The consistently higher pressures by direct carotid cannulation than by the indirect method probly is indicative of the pressure gradient , the length of the mouse tail (Ed- and Reinecke, '53).

In rats focal myocardial necrosis and pertension followed daily cortisone doses 5 mg/100 grams of body weight (Anichev, '64, '65). When the dose was to 2.5 mg, hypertension devel- but there was no histological evidence renal or cardiovascular damage (Knowlton, Loeb, Stoerk, White and Heffernan, '52). Doses of 2.0 mg daily failed to produce hypertension (Friedman, Friedman and Nakashima, '50). In mice DOCA plus salt loading produced chronic hypertension (Rosenblum, Donnenfeld and Aleu, '66). Cardiac hypertrophy and fibrinoid deposits within glomeruli and renal arterioles were found in these animals.

Mural hyalinization of small arteries occurs frequently in animals with hypertension (Goldblatt, Lynch, Hanzal and Sommerville, '34; Selye, '46; Wiener, Spiro and Lattes, '65), following immune responses (Karsner, '37), feeding of special diets (Dustin, '62) or following large doses of cortisone (Ashburn, Williams and Smith-Vaniz, '68). Hyalin may be degenerated smooth muscle (Montgomery and Muirhead, '56), plasma proteins (Dustin, '62; Adams, Bayliss and Orton, '67) or basement membrane material (Wiener, Spiro and Lattes, '65).

In this study the smallest dose of cortisone (0.5 mg) produced medial hyalinization of coronary arteries. The hyalinization was followed by a limited perivascular accumulation of fibroblasts and inflammatory cells. The inflammatory response may have been suppressed by the cortisone (Frenkel, '60). The mural and perivascular changes of coronary vessels may be reparative processes similar to those observed during recovery from adrenal enucleation or cortisone administration (Crane and Ingle, '65; Ashburn, Williams and Smith-Vaniz, '68).

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All photographs are of tissue stained with PAS plus hematoxylin.

PLATE 1

EXPLANATION OF FIGURES

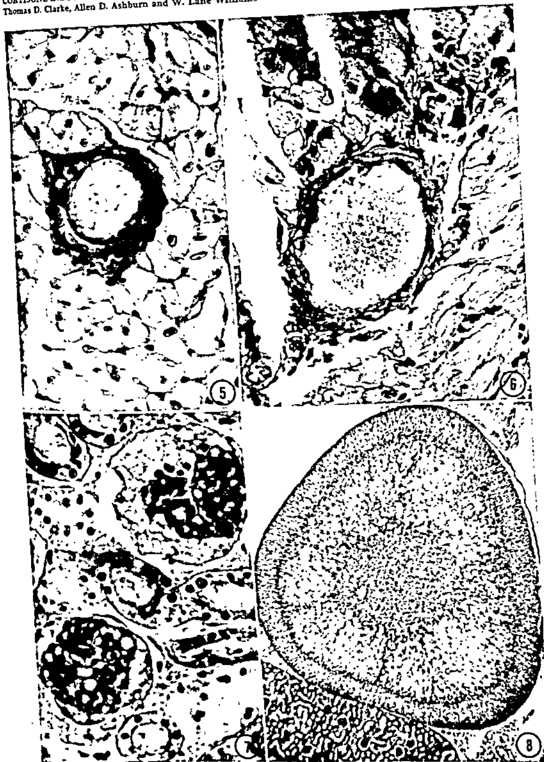
5 Mural and perivascular infiltration of fibroblasts and mononuclear cells. (0.5 mg of cortisone for 3 days) × 400.

6 Desquamation of endothelial cells and degeneration of the smooth muscle in the media. Myocardial necrosis is evident above the artery. (2.5 mg of cortisone for 7 days) × 400.

7 Plasma exudate within Bowman's space and convoluted tubules. The mouse had a blood pressure in excess of 210 mm Hg for five consecutive days. (2.5 mg of cortisone for 7 days) × 400.

8 Adrenal medullary vacuolization. (0.5 mg of cortisone for 7 days) × 40.

CORTISONE-INDUCED HYPERTENSION
Thomas D. Clarke, Allen D. Ashburn and W. Lane Williams



All photographs are of tissue stained with PAS plus hematoxylin.

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Studies on the Fine Structure of Ovarian Steroid-secreting Cells in the Rabbit

I. THE NORMAL INTERSTITIAL CELLS [1]

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ABSTRACT *In terms of their light microscopic appearance and fine structure the ovarian interstitial cells of the rabbit are typical steroid-secreting cells. They are* characterized by an abundance of agranular endoplasmic reticulum, spherical mitochondria with closely packed lamellar cristae, lipid droplets which appear to arise independently of the endoplasmic reticulum, conspicuous Golgi areas, a cytoplasm containing ribosomes and variable numbers of glycogen granules. A feature of the differentiation of the cells from the theca interna of atretic follicles or the stroma is the enlargement of the multiple Golgi areas and the progressive accumulation of agranular endoplasmic reticulum, possibly by "budding" from the Golgi cisternae. "Light" and "dark" cells are observed, the latter being characterized by a more closely packed agranular endoplasmic reticulum which tends to be tubular in type, that of the "light" cell being vesicular. Electron dense material (lipid?) is found in the vesicles and tubules of the agranular endoplasmic reticulum and in the Golgi cisternae; it may indicate a role of these structures in the biosynthesis of steroidal hormone. No fine structural changes specifically associated with pregnancy were observed. Degenerative changes are common and are described. The role of the interstitial cells, especially in relation to the production of 20 alpha-hydroxyprogesterone, is discussed.

The similarities between cells involved in the production of steroidal hormones, long observed by histologists (see Brambell, '62 and Corner, '63 for extensive reviews), have been confirmed in recent years by electron microscopy in a variety of tissues and species, for example the adrenal cortex (Luse, '67; Ross et al., '58; Lever, '59; Zelander, '59; Sabatini and de Robertis, '61), the interstitial cells of the testis (Fawcett and Burgos, '60; Christensen and Fawcett, '61; Carr and Carr, '63; Crabo, '63; Leeson, '63; Christensen, '65; Christensen and Fawcett, '66), the granulosa cells of the Graafian follicle (Wartenberg and Stegner, '60; Björkman, '62; Blanchette, '66), the thecal and interstitial cells of the ovary (Belt and Pearse, '56; Mota, '58), and the corpus luteum (Lever, '56; Enders, '62; Carsten, '65; Blanchette, '66, Flaks and Bresloff, '66; Rennels, '66). These studies and others under experimental conditions after hypophysectomy and treatment with trophic hormones (e.g. those of Sabatini et al., '62; Enders and Lyons, '64; Blanchette, '66; Schwarz and Merker, '67) have resulted in a substantial body of knowledge on the normal fine structure of these cells and on their subcellular responses to trophic hormones.

The steroid-secreting cells are characterized by an abundance of agranular endoplasmic reticulum, often arranged in concentric whorls around lipid inclusions. The mitochondria vary in form from organ to organ and from one species to another, having conventional lamellar cristae in some, tubular "cristae" in others. The Golgi areas are discrete in some cells; in others they have been described as "diffuse." Foldings and villous modifications of the plasma membranes are common, especially in the luteal cells. Free ribosomes, *areas of granular endoplasmic reticulum,* granules of glycogen and electron dense "microbodies" of uncertain origin have been described in the cytoplasm. The form of the agranular reticulum is a subject of

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This confirms the conclusions of others (Guraya and Greenwald, '64). The follicular origin of much of the interstitial tissue is indicated by the frequent occurrence of thickened and hyalinized remnants of the basement membrane of the stratum granulosum ("glassy membranes") embedded in the center of lobular masses of partially or completely differentiated cells. These remnants are especially common in pregnancy. The interstitial tissue of the immature animal is pink and translucent. In adult animals, especially after numerous episodes of pseudopregnancy or pregnancy, it becomes very compact and creamy in appearance.

Interstitial cells at an early stage in their "luteal-like" transformation (fig. 1) are epithelial in appearance. The nucleus is small and compact with indistinct nucleoli. The cytoplasm is granular and markedly acidophilic throughout. In later stages of their differentiation there is hypertrophy involving both nucleus and cytoplasm. Mitoses are rare and even when observed may be related to the stromal elements rather than to the interstitial cells. The nucleus becomes less compact and contains one or more conspicuous nucleoli. The cytoplasm becomes differentiated into a perinuclear acidophilic zone and a peripheral vacuolated zone (fig. 2). The perinuclear zone, which may be shown by a study of sections stained with borax-toluidine blue to contain many of the mitochondria, is a characteristic feature of the normal interstitial cell and its disappearance is an early and significant indication of degeneration. The loss of this acidophilic zone has been found to be a useful criterion for the assessment of functional maintenance of both interstitial and luteal cells in the rabbit, e.g. after hypophysectomy. The peripheral vacuolation is due to the removal of lipid droplets during preparation of the sections. These droplets are lost in the preparation of paraffin-embedded sections but are normally preserved after adequate fixation with osmic acid (figs. 5, 6).

In pseudopregnancy and pregnancy there is a marked hypertrophy of the ovarian interstitial cell (Lane-Claypon, '05) involving both nucleus and cytoplasm (figs. 3, 4), but without any significant increase in number of cells by mitosis (Claesson, '54). There also appears to be a significant accretion of new interstitial tissue by transformation of cells of the theca interna of atretic follicles. The "tempo" of follicular atresia seems to be increased during pregnancy (Halliday, '59) with a new formation of "corpora lutea atretica" or "pesudo corpora lutea" which, after a period of differentiation, become indistinguishable from the main interstitial cell mass. This irregular but progressive accretion of interstitial tissue associated with repeated episodes of pregnancy and pseudopregnancy (fig. 4), probably accounts for the extreme variability in the mass of interstitial tissue in adult rabbits irrespective to their body weight (Davies, '62). In old or senile rabbits the ovaries contain a predominance of interstitial tissue with few residual follicular elements. In a significant percentage (up to 30%) of rabbits obtained from local dealers, from whom no reliable reproductive history of the animal was available, a markedly hypertrophy of the ovarian interstitial tissue was observed over a period of years (Davies, '62). This was accompanied by atrophy of the corpora lutea, which were usually present, massive estrogenic hypertrophy of the uterus, cervix and vagina, hypertrophy of the liver of a type commonly observed after prolonged estrogen administration, severe degeneration of the inner zones of the adrenal cortex, and hypertrophy of the anterior lobe of the pituitary gland. This association of findings in "syndromic pattern" was such that the existence of the other features could be predicted from observation of any one at laparotomy. This "syndrome" appears to represent a spontaneous imbalance of pituitary-adrenal-ovarian function. It is observed so frequently in our experience that it must be considered carefully in all studies of reproductive function in the rabbit.

The vascularity of the ovarian interstitial tissue differs from that of the corpus luteum in the rabbit in that groups of cells rather than individual cells are in immediate contact with blood vessels. The interstitial tissue does not bleed freely on cutting, in contrast to the corpus luteum. Injections with India ink followed by re-

controversy, being described as tubular by some, as combined vesicular and tubular by others. Problems of fixation have greatly complicated the analysis of the probable exact form of the organelles, especially the endoplasmic reticulum and mitochondria. There are many unresolved problems such as the developmental origin of the lipid droplets in relation to the endoplasmic reticulum, and the significance of the droplets and organelles in relation to steroidal hormone biosynthesis.

The ovarian interstitial cells studied in this paper are a less well studied population of steroid-secreting cells. They present, however, unique features which make them especially suitable for the experimental analysis of fine structure and function. They are well developed in the rabbit, forming in many adult animals a major proportion of the total ovarian weight even in the presence of corpora lutea, and were described as forming an "interstitial gland" by Limon ('02). Interest in the cells has revived in recent years, chiefly as a result of the studies of Hilliard et al. ('63, '67) who have implicated them in the preovulatory synthesis and release of 20 alpha-hydroxyprogesterone following mating or gonadotrophic stimulation.

The ovarian interstitial cells as described in this paper have been shown to be typical steroid-secreting cells as far as their fine structure is concerned. They have been studied in non-pregnant ("estrous") and in pseudopregnant and pregnant rabbits. Though hypertrophy of the interstitial cells, involving apparently all cytoplasmic elements, takes place in pseudopregnancy and pregnancy, the changes have been judged to be quantitative and not qualitative in nature, and no specific changes associated with these states of enhanced gonadotrophic function could be determined. The studies reported here are intended as a basis for experimental studies on the interstitial cells of the rabbit after hypophysectomy and treatment with gonadotrophic hormone, to be published later.

MATERIALS AND METHODS

Ovarian interstitial tissue was obtained from New Zealand white rabbits in the following periods and stages of their reproductive life: immature (less than 5 months), mature (non-pregnant or "estrous"), pseudopregnant and pregnant, at term and in the postpartum period. Tissues for light microscopy were fixed in 10% neutral formalin or Bouin's solution. Tissues for electron microscopy were fixed for 40 minutes in buffered osmic acid in which the final concentration of osmic acid was 2%. The fixative was adjusted to a final osmolality of about 350 m.osmols by the addition of 0.3 M phosphate buffer (final concentration of buffer about 0.15 M) at a pH of 7.2–7.4. Osmolality was determined by the method of freezing point depression, using a Fisk osmometer. Increasing the final concentration of osmic acid from 1% to 2% resulted in markedly better and consistent fixation. The use of sucrose for the adjustment of osmolality was found to be disadvantageous, resulting in shrinkage of the cells, and disruption of plasma membranes and those of the organelles. Fixation was also carried out in glutaraldehyde (1.25%) with phosphate buffer at a final osmolality of about 350 m.osmols, followed by osmication in 2% osmic acid-phosphate buffer. Tissues were rapidly dehydrated in ethanol (70%, 80%, 90%, 100%) and in absolute butanol (one hour) followed by clearing in toluene for one hour. They were then embedded in English or American araldite and sectioned on a Porter-Blum MT-2 ultramicrotome. Phase contrast sections were studied for purposes of orientation before final trimming of the face of the block and only those areas showing good preservation of lipid (usually peripheral) were used for thin sectioning. Some were stained with borax-toluidine blue. Thin sections were mounted on 400-mesh uncoated copper grids and stained with uranyl acetate (saturated solution in 50% alcohol) and lead citrate (Reynolds, '63). The sections were examined in a Hitachi HU-11B electron microscope.

OBSERVATIONS

Summary of light microscopic observations

The development of the ovarian interstitial cells is clearly by a "luteal-like" transformation of fibroblastic elements of the theca interna of follicles, both atretic and recently ovulated, and also of the stroma.

ternae and vesicles of the Golgi areas usually contain a small amount of homogeneous material of moderate electron density. The cytoplasm of the interstitial cells at this stage (fig. 9) is densely crowded with small vesicles, presumably of the agranular endoplasmic reticulum. They are predominantly small at first, the largest ones approaching in size that of the dilated vesicles of the Golgi areas. Like the latter they contain electron dense material which completely fills the vesicles in the case of the smallest ones, while in the larger ones it forms a peripheral condensation surrounding an empty central area. The nucleus is surrounded by a continuous outer membrane with multiple areas of contact with the inner nuclear membrane at typical nuclear pores. The cytoplasm is finely granular and contains in addition many free ribosomes arranged singly or in small clusters. There are occasional areas of granular endoplasmic reticulum.

At the stage of differentiation described above and before the deposition of significant amounts of lipid, the interstitial cells show a uniformly granular acidophilic cytoplasm (as in fig. 1). This acidophilia presumably must be attributed to predominantly acidophilic proteins which may include not only those of the mitochondria but also of other elements of the cytoplasm, notably the endoplasmic reticulum. The first indication of the deposition of lipid in the interstitial cell is the appearance of irregular spaces within the peripheral cytoplasm. The spaces frequently appear empty, suggesting that any lipid they may contain at this stage is easily dissolved out during the preparation of the tissues, or that it is of extremely low electron density. It may usually be demonstrated in phase contrast sections as a delicate stippling of droplets not easily distinguished from the mitochondria which are also osmiophilic and of comparable size. It has been found impossible to detect a unit membrane circumscribing individual droplets, though there is usually a peripheral condensation of coarsely granular and electron dense material (figs. 9, 12). At this stage in the differentiation of the interstitial cells, therefore, the lipid droplets appear to arise *de novo* within the cytoplasm and independently of the endoplasmic reticulum, often in areas rich in free ribosomes. Though some of the early spaces appear empty, other spaces in the same cell or in adjacent cells frequently contain homogeneous electron dense lipid, suggesting marked variation in solubility or osmiophilia at this stage.

As the interstitial cells approach the stage of full differentiation (figs. 10, 12) the lipid droplets become more numerous and the cytoplasm becomes crowded with membranes of the Golgi systems and endoplasmic reticulum. The Golgi areas are conspicuous in extent and size and appear in some places in direct continuity with vesicles of the agranular endoplasmic reticulum (fig. 12). The cisterns and vesicles contain electron dense material as described in earlier stages (figs. 10, 12). The mitochondria are uniformly spherical or slightly oval and contain internal membranes of greater complexity. These membranes are arranged in groups of parallel cristae, one group being parallel to another or to the surface of the mitochondrion. Tubular forms of the internal mitochondrial membranes cannot be ruled out, but could be artifactual and result from tangential sections of groups of lamellar cristae. The mitochondrial matrix is finely granular and of considerable electron density. Free ribosomes occur singly or in small clusters but are more scattered, representing either an actual decrease in their numbers compared with earlier stages or a wider distribution throughout the now more extensive cytoplasm. Electron dense granules which are larger and more variable in size than the ribosomes and which are stained selectively with lead salts probably represent glycogen (fig. 12). Electron dense bodies are often observed, particularly in the vicinity of the Golgi areas (fig. 12). Evidence for their possible origin from lipid droplets will be presented later.

Degeneration of interstitial cells occurs frequently even in immature animals. It is characterized by a loss of the formed elements of the cytoplasm (Golgi, mitochondria, endoplasmic reticulum), and by the appearance of "myelin figures" and crystalline structures (cholesterol?). The contents of the crystalline structures, except

construction of the vascular pattern in serial sections indicate that the vascular beds of the interstitial tissue and corpus luteum are independent and no portal system of vessels linking them exists.[3]

The lipid droplets in the vacuolated areas of the interstitial cell cytoplasm are illustrated following fixation in osmic acid (figs. 5, 6). The droplets are uniform in size (estimated at 1–2 μ) within a single cell but vary considerably from one cell to another. In non-pregnant animals the droplets appear more crowded and relatively numerous. In pregnant animals the droplets appear less densely packed. "Light" and "dark" cells appear in the ovaries of both the non-pregnant and pregnant rabbit (fig. 5). The varying density of the cells, as studied by phase contrast microscopy, is due not only to the degree of crowding of the droplets but also to an increase in the density of the intervening cytoplasm in the "dark" cells.

Degeneration of interstitial cells is common in the ovaries of all mature rabbits whether pregnant or not. It is usually unicellular in type (fig. 7) but is occasionally "lobular" (fig. 8). In each instance the cytological appearance of degenerating cell is the same. The cell is increased in size. There is nuclear pycnosis and a loss of the perinuclear acidophilic zone. Vacuolation of the cytoplasm is complete, and in paraffin-embedded sections the spaces appear larger and the septa between them correspondingly attenuated. Study of frozen sections and sections observed after fixation in osmic acid shows that the large spaces contain lipid droplets which are, however, larger and more varied in size than in the normal cell. They are also less soluble in organic solvents such as acetone than are the droplets of normal cells but are eventually dissolved by prolonged treatment. Late stages in the degeneration of the cells (fig. 7) are associated with swelling, pycnosis, and eventual loss of the nucleus, and apparent disappearance of cytoplasmic material the former site of which is marked by irregular empty spaces.

Electron microscopic observations

The fine structure of the interstitial cells will be described in relation to three principal stages of their presumed life cycle: (1) immature differentiating cells, (2) mature cells, and (3) degenerating forms.

(1) *Immature differentiating cells.* These are easily studied in young rabbits of less than five months of age. At this time, in our experience, these cells are becoming able to respond to mating with ovulation; there is much variation, however. The interstitial cell mass during this period is pink and rather translucent to the naked eye in contrast to its creamy appearance in the adult animal. Immature interstitial cells, however, may be observed in all stages of their "luteal-like" transformation from fibroblasts of the theca interna of atretic follicles and of the stroma in adult animals (fig. 4), especially during pregnancy.

Interstitial cells in the early stages of differentiation are notable for their relative simplicity of cytoplasmic fine structure. They contain many free ribosomes and occasional elements of the granular endoplasmic reticulum but few of the agranular endoplasmic reticulum. This stage is presumably associated more with protein synthesis and growth. The Golgi areas are discrete but inconspicuous. The plasma membranes are uncomplicated and in simple apposition with each other with occasional areas of contact, forming typical desmosomes. The mitochondria are few and are elongated in form with simple lamellar cristae.

Further differentiation of the interstitial cells is accompanied by increased complexity of cytoplasmic fine structure, in addition to the nuclear changes and cytoplasmic growth observed in the light microscope (fig. 1). The mitochondria are increased in number and, at an early stage, become spherical or oval in form with more complex internal membranes. These membranes are arranged predominantly in lamellar form, rarely in tubular form, and lie in a finely granular matrix. The Golgi areas are now conspicuous and usually multiple. They consist of parallel tubules or cisterns each of which has a dilated vesicular extremity. The cis-

[3] We are indebted to Miss Jane Davy of St. Thomas's Hospital Medical School, London, England, who carried out these studies during the tenure of a summer fellowship.

16). A thin discontinuous basement membrane envelops the interstitial cells and the cytoplasmic processes and extends around groups of cells but not between individual cells. The basement membrane of the blood vessels in the connective tissue septa is complete.

In interstitial tissue fixed in glutaraldehyde the cytoplasm shows that there is an additional feature not seen after fixation in osmic acid, namely "microtubules" (fig. 17). These microtubules are approximately 270–300 Angstroms in diameter and are rather uniform in caliber. They are easily observed in "light" cells in which there are extensive areas of cytoplasm between the other organelles and inclusions; they are undetectable in "dark" cells.

The mitochondria in both "light" and "dark" cells are similar, and it has not been possible to determine whether they are more numerous in one or another type of cell. They are almost invariably round or slightly oval, rarely elongated, and contain characteristic internal membranes. As previously described, these membranes are arranged in groups of lamellae, one group being at an angle to another or parallel to the surface of the mitochondrion. Tubular "cristae" are rare. The mitochondrial matrix is finely granular and of considerable electron density. It has not been observed to contain large granules at any stage. Ballooning of the outer membranes, vesiculation of the matrix, discontinuities in the outer membranes, and other features described by others in steroid-secreting cells have not been observed.

Free ribosomes become relatively rare in fully differentiated interstitial cells, especially in the so-called "dark" cells. Glycogen granules are also very variable in number. They are generally sparse or absent in apparently well differentiated cells. They are more common in "light" than "dark" cells and, as shown previously (fig. 12), more common in undifferentiated than in differentiated cells. Glycogen is often observed in the endothelial cells of the blood vessels (fig. 18), and this appears to be more common in late pregnancy than at any other time. In late pregnancy the thickening and reduplication of the basement membrane of the interstitial cells, noted in late pregnancy may represent an involutional phenomenon (fig. 18); it has not been observed in non-pregnant animals.

A feature of many interstitial cells at all stages in the mature rabbit is the presence of variable numbers of extremely electron dense bodies. These are often observed in the perinuclear zone associated with the Golgi area (figs. 12, 15). A study of interstitial cells which show other indications of diminishing function or regression (e.g. fig. 19) provides evidence for the origin of these structures from existing lipid droplets. The transformed droplets appear uniformly round, in contrast to the highly irregular homogeneous lipid of fully differentiated cells in young rabbits (c.p. fig. 19 with figs. 13, 14, 16). They also show a peripheral condensation of extremely electron dense material which may encircle the droplet evenly or in the form of a crescent. The absence of an observable limiting membrane around these droplets, and the less intimate relationship between them and the endoplasmic reticulum, confirms the impressions gained from a study of early differentiated cells (see fig. 9) that they exist independently of the endoplasmic reticulum. The transformed droplets also show a gradation in electron density from the periphery to the center (fig. 19). There is frequently a central empty area from which the lipid has apparently been dissolved, suggesting poor penetration of the osmic acid into the droplets during fixation. Other droplets are completely transformed into electron dense bodies similar to those observed previously in the Golgi area (fig. 14); they are considerably reduced in size when compared with the original lipid droplets.

(3) *Degenerating cells.* Degeneration of interstitial cells, as shown by light microscopy (figs. 7, 8) usually involves single isolated cells, but is occasionally lobular in extent. Regression of the interstitial cells is observed in both "light" and "dark" cells. Regression is associated with changes in the mitochondria which show a simplification of the cristae (fig. 19), later gross shrinkage and a marked increase in the electron density of the matrix (fig. 20). The endoplasmic reticulum appears to show marked resistance to degeneration and persists in some form in degenerated cells for an undetermined but probably

for a peripheral condensation of electron dense material, are usually dissolved out.

(2) *Mature interstitial cells.* These are described irrespective of the stage of the reproductive life of the rabbit for the ensuing reasons. The changes in the cells during pregnancy and pseudopregnancy, involving the nuclear and cytoplasmic hypertrophy observable in the light microscope (fig. 4), are seen by electron microscopy to be accompanied by an increase in the number and extent of existing organelles and inclusions (mitochondria, endoplasmic reticulum, lipid droplets, etc.) without, however, detectable qualitative difference in their organizations and interrelationships. The changes then are assumed to be quantitative rather than qualitative. Moreover, the extreme variability of fine structure of the interstitial cells in mature rabbits, complicated by the continuous new-formation of cells from atretic follicles, the widespread occurrence of degenerative changes, and the notorious sampling errors inherent in electron microscopy, make the detection of systematic fine structural alterations specifically related to pregnancy and pseudopregnancy impossible to define. This is in marked contrast to the corpus luteum in which systematic changes in the luteal cells occur in pregnancy and pseudopregnancy.

A feature of the interstitial cells as the rabbit approaches maturity (figs. 13, 14) is the presence of many lipid droplets crowding the cytoplasm. The droplets appear highly irregular in form and contain homogeneous material of a uniform electron density throughout the cytoplasm. Irregular stellate depots of lipid occur in some areas rich in free ribosomes but without membranes of the endoplasmic reticulum (fig. 14), suggesting that here they arise independently. In other areas (figs. 13, 14) the droplets of lipid are enveloped on all sides by vesicular and tubular elements of the endoplasmic reticulum, and here it is more difficult to make a case for their independent origin. In many regions the droplets are so close together that the intervening cytoplasm, containing small vesicles, ribosomes, glycogen, etc., is compressed and often attenuated, forming delicate septa. The irregularity of these droplets and contours of the cytoplasmic septa between them impart to the cytoplasm a characteristically "lacy" appearance (fig. 13). Golgi areas are invariably present but less easily distinguished from the all-pervading network of endoplasmic membrane than in undifferentiated cells (c.p. fig. 12). The cells lie in intimate contact with each other except in the region of the perivascular space where folding of the plasma membranes occurs with the formation of slender processes (fig. 13). These processes are often devoid of lipid inclusions but are especially rich in free ribosomes and glycogen.

"Light" and "dark" cells are regularly but variably observed in the interstitial cell mass of the mature rabbit (see fig. 5). The differences between these two types of cells are seen in the electron microscope to be determined by the amount of agranular endoplasmic reticulum and to the resultant compression of the intervening cytoplasmic structures (fig. 16). In a typical "light" cell (fig. 15) the cytoplasmic structures are not compressed, and discrete Golgi areas are easily detected. The endoplasmic reticulum, moreover, is predominantly vesicular in such "light" cells and the electron dense contents of the vesicles are easily observed. The cytoplasm between the vesicles contains scattered ribosomes and occasional larger granules of glycogen Small areas of granular endoplasmic reticulum occur, and in such areas continuity between it and the vesicles of the agranular endoplasmic reticulum is occasionally observed (fig. 15). In "dark" cells the agranular endoplasmic reticulum is extremely complex and appears to be made up of short tubules (fig. 16). In these cells the electron dense contents of the tubules of endoplasmic reticulum are less conspicuous or absent. All variations in cell ty[illegible] between "light" and "dark" cells exist. Th[illegible] differences are accentuated by fixation in hypertonic solutions, probably as a result of shrinkage and compression. The [illegible]chondria are similar in both types of cells They are numerous in mature cells [illegible] have the characteristic oval or spherica[illegible] forms previously described. The folding [illegible] the plasma membrane and the formation of slender cytoplasmic processes which [illegible]tend into the perivascular space are al[illegible] more conspicuous in mature animals (fig

and nucleolus, the organelles (Golgi, mitochondria, endoplasmic reticulum), a deposition of lipid droplets, and modifications of the plasma membrane. As the rabbit approaches maturity (about 5 months) the cells are crowded with lipid droplets and the agranular reticulum is abundant. Ribosomes appear to decrease relatively or absolutely, and there is an appearance of glycogen in monoparticulate form, recognized by its relative size and variability in comparison to ribosomes and its selective affinity for lead salts (Revel et al., '60). Blanchette ('66) observed glycogen during the differentiation of the granulosa cells under normal and experimental conditions in which luteinizing hormone was injected into the follicle of the rabbit; it was monoparticulate in the early stages, mostly in rosette-form in the later stages of luteal transformation.

The conformation of the organelles in fixed steroid-producing cells has been a subject of controversy. The form of the endoplasmic reticulum, vesicular or tubular, has been a particular problem, and several writers have concluded that the vesicular form is an indication of poor fixation, and that the tubular form more closely approximates to its condition in the living state (Enders, '62; Christensen, '65). Both types of agranular endoplasmic reticulum have been observed, however, by many investigators and there are undoubtedly marked variations in its form from one cell to another, from one zone to another (as in the adrenal cortex), and from one species to another. Blanchette ('66) observed both vesicular and tubular forms of the endoplasmic reticulum in the corpus luteum of the rabbit with all kinds of fixation and with all variations in osmolality within reasonable limits. Hypertonic fixatives, especially those containing sucrose, in our experience caused marked shrinkage of the cells with compression of the endoplasmic reticulum, which then appears predominantly tubular. However, in normal rabbits in which the ovarian interstitial tissue was fixed at a controlled osmolality (350 m.osmols) both vesicular and tubular forms of the agranular endoplasmic reticulum were observed and the cells in all other respects showed evidence of good fixation.[3] "Light" and "dark" cells were observed, similar to those described by Enders ('62) in the corpus luteum, the endoplasmic reticulum being sparse and vesicular in the former, densely packed and tubular in the latter. Since proliferation of the agranular reticulum has been shown to be a significant subcellular response of the interstitial cells to luteinizing hormone resulting in an almost complete transformation of all interstitial cells into "dark" cells (Davies, '66), we have tentatively concluded that "dark" cells may represent cells at a higher level of gonadotrophic stimulation than are "dark" cells.

A feature of the vesicular elements of the agranular endoplasmic reticulum in the interstitial cells, especially the "light" cells, was their content of electron dense material, possibly representing lipid. Similar material was observed in the dilated extremities of the Golgi cisterns and in the cisterns themselves. This material may have some relevance to the unsolved problem of how and where the steroidal hormones are synthesized or conveyed out of the cell. Hamilton et al. ('67) in our laboratory has shown that perfusion of the liver with free fatty acids results in the accumulation of electron dense bodies comparable in size to those observed in the interstitial cells of the rabbit. The Golgi cisterns and vesicles of the hepatic cell are also apparently involved in the synthesis or transport of this material which appears to represent "very low density" lipoprotein. It is possible that the application of methods and concepts widely developed in the field of hepatic lipid transport to the unsolved problems of steroidal hormone biosynthesis at the subcellular level may be of great value. The electron microscopic evidence is not inconsistent with the view that the vesicles of the agranular endoplasmic reticulum may be formed by "budding" from the Golgi area, a suggestion made by Mercer ('62) for cells in general and by Schwarz and Merker ('67) for

[3] We have recently had the opportunity to study the interstitial cells of the opossum testis fixed in the same manner as described in this paper (2% osmic acid, approximately 0.15 phosphate buffer, 350 m.osmols). Tubular endoplasmic reticulum identical to that described by Christensen and Fawcett ('61) predominated but there were extensive areas of vesicular endoplasmic reticulum not only in different cells but also in the same cell. A similar conclusion was reached by de Kretzer ('67) from a study of the human testicular interstitial cells.

prolonged period (fig. 21). There is a disorganization of the regular pattern of organization of the endoplasmic reticulum (fig. 20). The vesicles, formerly occupied by electron dense material, are more often empty. In many areas the endoplasmic reticulum forms closely packed systems of parallel membranes, superficially resembling Golgi areas (fig. 20) but differing in essential particulars from them (c.p. fig. 15). Similar Golgi-like areas may be of mitochondrial origin (fig. 20). The formation of concentric whorls of membranes, having the unit-membrane structure when appropriately resolved, increase throughout the cytoplasm in degenerating cells and may ultimately become the predominant feature (fig. 21); these are the so-called "myelin" figures, and may result from the hydration of existing protein-phospholipid material.

The lipid droplets also show marked resistance to degeneration and may persist long after the formed elements of the cytoplasm are unrecognizable (figs. 20, 21). Aggregations of very electron dense granules are also observed in degenerating cells (fig. 21). Their nature cannot be ascertained. The granules appear too large and uniform and, in addition, are not selectively stained with lead salts, suggesting that they are not glycogen. They may represent pigmentary inclusions.

DISCUSSION

The interstitial cells of the rabbit ovary, as judged by light and electron microscopic criteria, are typical steroid-producing cells. Unlike the luteal cells, which are formed only as a result of an ovulatory stimulus, the interstitial cells are a permanent element of the ovary and constitute a significant mass of steroid-producing cells. Like the cells of the adrenal cortex and the interstitial cells of the testis, they form a stable population of cells, in contrast to the luteal cells which appear to have an inherently limited life span under normal conditions. The luteal cells, moreover, regress and degenerate after hypophysectomy, whereas the interstitial cells revert under these conditions to a resting or "basal" state in which they are capable of further response to gonadotrophins (Velardo, '59; Davies, '66). In terms of pituitary-dependence the luteal cells are the most dependent; the ovarian interstitial cells, the interstitial cells of the testis and the outer zones of the adrenal cortex are the least dependent; and the cells of the inner zones of the adrenal cortex appear to be intermediate in dependence. The luteal cells appear to function normally at a maximal level of gonadotrophic stimulation and steroidal output, whereas the ovarian interstitial cells function at a lower level which can be raised by suitable gonadotrophic stimulation. Preliminary studies in our laboratory on the effects of luteinizing hormone on the interstitial cells of hypophysectomized rabbits (Davies, '66) indicate that under these circumstances the interstitial cells become fine structurally indistinguishable from luteal cells, with respect to the rich development of the endoplasmic reticulum and the complex folding of the plasma membrane. Though pregnancy appears to be associated with increased levels of gonadotrophic hormone (luteinizing hormone?, Davies et al. ('65)), it is of interest that no fine structural changes typical of pregnancy could be detected. The rate of turnover of the interstitial cells is unknown (Brambell, '62). The increase in size of the interstitial tissue in pregnancy is due to hypertrophy of individual cells and not to mitotic division (Claesson, '54).

The progressive differentiation of interstitial cells throughout the first five months of life in the rabbit ensures that a large mass of competent steroid-secreting cells is available to meet the demands of reproductive life. It is not known if this differentiation is governed wholly or in part by luteinizing hormone, as it appears in the luteal transformation of granulosa cells of the follicle. If so, luteinizing hormone or whatever other hormones are involved, must be regarded as morphogenetic hormones as well as trophic hormones. There is some evidence (Kilpatrick et al., '64) that luteinizing hormone is the trophic hormone of the ovarian interstitial cells in the rabbit.

In the differentiation of interstitial cells from cells of the theca interna of follicles and the stroma, all parts of the cells appear to change *pari passu*. The changes, in addition to growth, involve the nucleus

(minus corpora lutea) of the rabbit. They have demonstrated activity with 3 beta-, 17 beta-, and 20 alpha-hydroxysteroids. Preferences of one enzyme system for DPN or TPN were observed, and some were observed in the microsomal fraction and others in the supernatant. The role of the interstitial cells in the production of androgen or estrogen has not been substantiated (Mericard, '53; Eckstein, '62). Falck's observations on microtransplants of ovarian tissue in rats ('59) that these do not function independently but in synergy in the production of estrogen (e.g. granulosa cells plus theca interna cells, granulosa cells plus interstitial cells) may underlie the failure to ascribe a specific role to the ovarian interstitial tissue.

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steroid-secreting cells in particular. Areas of granular endoplasmic reticulum are also commonly observed in steroid-producing cells, and direct continuity between it and vesicles or tubules of the agranular endoplasmic reticulum are also common, as shown in this study, and by others (Ross et al., '58; Enders, '62; Christensen, '65; Flaks and Bresloff, '66; Zelander, '59). These areas may be associated with regions of protein, or possibly lipoprotein, synthesis.

The mitochondria of steroid-secreting cells are also extremely varied in fine structure (Sheridan and Belt, '64), being of giant size in some cases, e.g. the fetal human adrenal cortex (Luse, '67), and in all cells often confused in phase-contrast microscopy with lipid droplets. The mitochondria of the ovarian interstitial cells become ovoid or spherical at an early stage in their differentiation and have an extremely complex system of internal cristae which are predominantly lamellar. Tubular or "villiform" cristae, though a regular feature of the zona fasciculata of the adrenal cortex and human fetal adrenal cortex, do not seem to be typical of all steroid-secreting cells. Internal membranes of this type are also seen in other organs (e.g. liver, placenta) not exclusively concerned with steroidal hormone production. Shrinkage and increased density of the mitochondrial matrix has been found to be an early indication of degeneration in interstitial cells. These shrunken forms resemble the "condensed" forms described in liver by Hackenbroeck ('66) and shown by him to be potentially reversible.

Information on the correlation of fine structure and function in the ovarian interstitial cells, as in all steroid-secreting cells, is fragmentary. Deane ('58) has emphasized that the number of lipid droplets is not necessarily a good criterion of functional activity. Isolation of the lipid droplets from the rabbit ovary (Claesson, '54) has shown that they are extremely stable over a wide range of time and physical conditions. About 75% of their lipid is esterified cholesterol, the remaining lipid being fatty acid (triglyceride, 23%), free cholesterol (1%), and phospholipid (1%). Following acute gonadotrophic stimulation the droplets shrank slightly in size and showed a decrease in esterified cholesterol to about 25%, a rise in fatty acid, and a rise in phospholipid to 5%. These results, and those derived from histochemical studies (Rennels, '51; Falck, '53; Jacoby, '62) are consistent with the view that the droplets contain a depot of cholesterol to serve the needs of steroidal hormone biosynthesis. The depletion studies of Claesson are also consistent with the histochemical observation of a depletion of birefringent Schiff-positive and Schultz-positive material in the interstitial cells (not necessarily in the droplets) following gonadotrophic stimulation, and an accumulation after hypophysectomy. Boström and Odelblad ('52) observed an inverse relationship between the uptake of P^{32} and the amount of birefringent material in the interstitial tissue of the rabbit. Gonadotrophic stimulation, as after mating or the injection of luteinizing hormone, is also accompanied in the rabbit ovary by a depletion of ascorbic acid (Claesson, '54), by an almost total conversion of inactive to active phosphorylase (Davenport and Longley, '67), and by an outpouring of 20 alpha-hydroxyprogesterone into the ovarian venous blood (Hilliard et al., '63). This steroid appears to be the principal progestational steroid of the rabbit during the preovulatory and possibly the later stages of pregnancy. Though it has only about one-eighth the progestational activity of progesterone, it is produced in about ten times the amount (Rennie and Davies, '65). Rennie et al. ('65) have studied some of the enzymes of the glycolytic cycle in rabbit ovarian tissue for 8 days after mating, viz. glucose-6-phosphate dehydrogenase, phosphoglucomutase, and phosphohexose isomerase. The activity of these enzymes showed no change that could be related to pregnancy but in those cases where variation was observed seemed to be related to the mass of interstitial tissue. Histochemically the rabbit's ovarian tissue shows marked hydroxysteroid dehydrogenase activity (Rubin et al., '63; Davies et al., '66), especially 3 beta-ol dehydrogenase activity, essential for the conversion of pregnenolone to progesterone. Davenport and Mallette ('66) have made a study of hydroxysteroid dehydrogenase activity in the ovary

(minus corpora lutea) of the rabbit. They have demonstrated activity with 3 beta-, 17 beta-, and 20 alpha-hydroxysteroids. Preferences of one enzyme system for DPN or TPN were observed, and some were observed in the microsomal fraction and others in the supernatant. The role of the interstitial cells in the production of androgen or estrogen has not been substantiated (Mcricard, '53; Eckstein, '62). Falck's observations on microtransplants of ovarian tissue in rats ('59) that these do not function independently but in synergy in the production of estrogen (e.g. granulosa cells plus theca interna cells, granulosa cells plus interstitial cells) may underlie the failure to ascribe a specific role to the ovarian interstitial tissue.

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PLATE 1

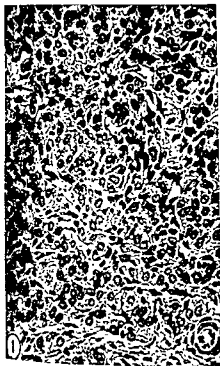
EXPLANATION OF FIGURES

1 Ovarian interstitial tissue of immature virgin rabbit (2 kilos). The cells are small with a relatively small area of cytoplasm which is uniformly acidophilic. The cells are arranged in clusters with thick septa of connective tissue between them. At the top left (arrow) is the remains of an atretic follicle. H and E, × 335.

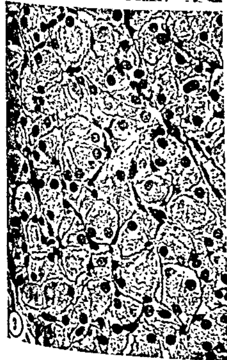
2 Interstitial tissue of non-pregnant ("estrous") rabbit (3 kilos). The cells are larger (c.p. fig. 1) and show a much greater ratio in area between cytoplasm and nucleus. The cytoplasm is vacuolated except for a small perinuclear zone of acidophilic material. The cells are in close mutual contact with only occasional capillaries which lie predominantly in the septa of connective tissue between clusters or lobules of interstitial tissue. H and E, × 335.

3 Interstitial tissue of rabbit at the ninth day of pregnancy. The cells are hypertrophied (c.p. fig. 2). The perinuclear and acidophilic zone is particularly conspicuous. The peripheral cytoplasm is vacuolated. H and E, × 335.

4 Interstitial tissue of rabbit at the twenty-fifth day of pregnancy. This area illustrates the lack of homogeneity in the cellular population of the interstitial tissue but also illustrates its essential development and final arrangement in clusters or lobules. To the right is an area of immature cells (c.p. fig. 1), arising in relation to the theca interna of an atretic follicle. To the left is a group of interstitial cells of mature type comparable to those in figure 3. H and E, × 335.


1


2


3

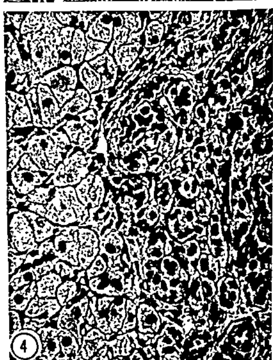

4

PLATE 2

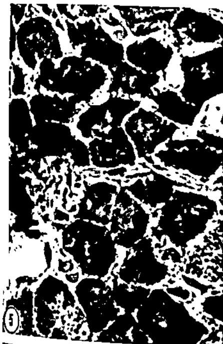
EXPLANATION OF FIGURES

5 Phase-contrast photograph of ovarian interstitial tissue of a non-pregnant ("estrous") rabbit. The cells are packed with small droplets of osmiophilic material (lipid). "Dark" and "light" cells are present, the difference being due to the density of the droplets and to the *greater density of the background cytoplasm. The droplets* vary in size but are constant in size within any one cell. Fixed in 2% osmic acid and buffered phosphate (350 m.osmols, pH 7.4), × 1,000.

6 *Phase-contrast photograph of ovarian interstitial tissue* of pregnant rabbit at the sixteenth day. The cells are hypertrophied and there is less distinction between "dark" and "light" cells. The lipid droplets are numerous but more widely spaced. Fixation as in figure 6, × 1,000.

7 Area of ovarian interstitial tissue of rabbit at the twenty-third day of pregnancy showing the occurrence of individual degenerating cells (at arrows). These cells are enlarged, showing nuclear pycnosis and *an "empty" or "stippled" cytoplasm. The perinuclear acidophilic* zone is absent in the degenerated cells (N.B. loss of this zone is a significant early sign of loss of function or of early degeneration). H and E, × 355.

8 A different and rarer type of degeneration of interstitial cells in a non-pregnant ("estrous") rabbit. In this case the degeneration is "lobular" in pattern rather than "individual" and involves a large *group of cells, presumably one once derived from the wall* of an atretic follicle. The cellular appearance of the degenerating cells is as in figure 7. H and E, × 100.


5


6


7


8

Abbreviations

b, electron dense body
c, crystal
d, dilatation of Golgi
e, endothelium
f, myelin figure
g, Golgi
l, lipid
m, mitochondrion
p, cell processes
n, nucleus
nl, nucleolus
r, rough endoplasmic reticulum
v, vesicle

All succeeding figures are electron micrographs. They are all of ovarian interstitial tissue unless otherwise indicated, fixed in 2% osmic acid adjusted with 0.3 M phosphate buffer to a final osmolarity of about 350 m.osmols (final buffer concentration about 0.15 M) and at a pH of about 7.2. Tissues were embedded in English araldite and stained with uranyl acetate and lead citrate.

PLATE 3

EXPLANATION OF FIGURES

9 Interstitial cell from ovary of immature rabbit of 7.5 weeks. The appearance of such a cell in the light microscope is shown in figure 1. There are a few mitochondria, elongated or oval in shape, containing lamellar cristae. The Golgi areas are prominent. The cytoplasm contains vesicular bodies assumed to be representative of the smooth endoplasmic reticulum, which contain electron dense material (lipid?). An irregular and larger vesicle (at arrow) appears empty; it may have contained lipid before fixation. There are many cytoplasmic granules, one population being small and probably representing ribosomes, and a second population of larger size which is possibly glycogen. × 70,000.

10 Details of immature interstitial cells from prepubertal rabbit of about five months. The cytoplasm in this area contains mitochondria, small vesicles of the endoplasmic reticulum which contain electron dense material and granules (probably ribosomes). × 65,000.

11 Degenerating interstitial cell from immature rabbit of five months showing the loss of cytoplasmic organization (c.p. fig. 10). In addition there are spaces whose shape suggests that they were formerly occupied by crystals (cholesterol?). There are also "myelin figures" and other electron dense masses which may be lysosomes. × 33,000.

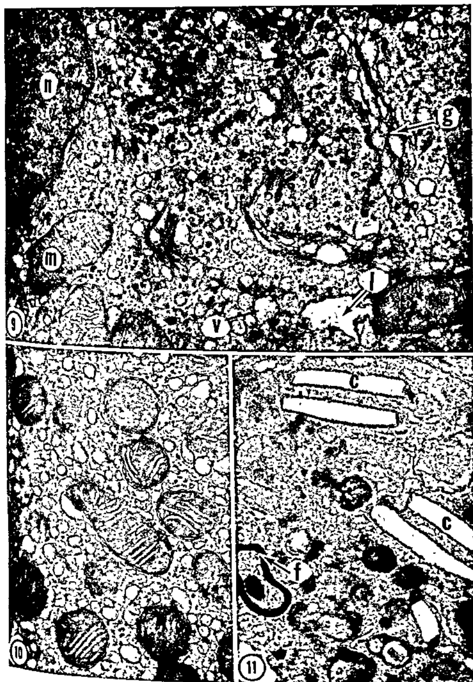

n
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m
l
9
v
c
c
f
10
11

PLATE 4

EXPLANATION OF FIGURE

12 Partially differentiated (immature) interstitial cell of prepubertal rabbit of about 5.5 months. The mitochondria are predominantly spherical with the irregular cristae typical of mature cells (c.p. fig. 15). Large lipid droplets, partially or completely emptied of their contents, are observed at top right and bottom right. The cytoplasm contains a complex system of smooth membranes. Some are arranged in parallel rows enclosing cisternal spaces containing homogeneous electron dense material, and they are assumed to be Golgi areas. Regional dilatations of the Golgi cisterns are observed at d. Elsewhere the membranes are disposed in the form of roughly spherical vesicles or short tubules containing electron dense material. There are a few electron dense bodies within the Golgi area. Cytoplasmic granules probably represent ribosomes and glycogen. × 40,000.


l
m
g
d
b
l
g
12

PLATE 5

EXPLANATION OF FIGURE

13 Almost fully differentiated interstitial cells from a young non-pregnant rabbit of about 5.5 months. Two cells are observed with their plasma membranes in close contact. There is an extensive extra-cellular space in relation to the blood vessels at the lower end of the figure, containing wisps of collagen, processes of other interstitial cells and traces of basement membrane in relation to the processes and to the endothelium of the vessels. The mitochondria are typical of mature interstitial cells. There are large lipid droplets, highly irregular in shape and containing homogeneous electron dense material. There are, in addition, small vesicles, also containing electron dense material. Ribosomes and glycogen are rare. × 45,000.

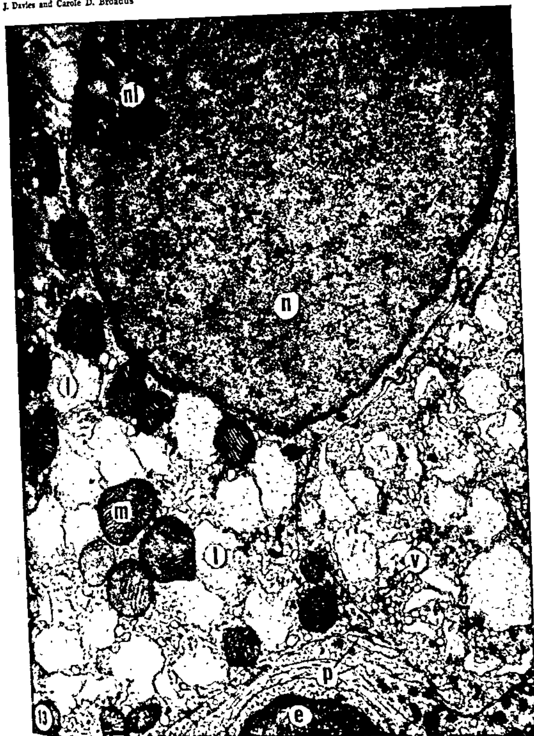

nl
n
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m
l
v
p
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13

PLATE 5

EXPLANATION OF FIGURE

13 Almost fully differentiated interstitial cells from a young non-pregnant rabbit of about 5.5 months. Two cells are observed with their plasma membranes in close contact. There is an extensive extracellular space in relation to the blood vessels at the lower end of the figure, containing wisps of collagen, processes of other interstitial cells and traces of basement membrane in relation to the processes and to the endothelium of the vessels. The mitochondria are typical of mature interstitial cells. There are large lipid droplets, highly irregular in shape and containing homogeneous electron dense material. There are, in addition, small vesicles, also containing electron dense material. Ribosomes and glycogen are rare. × 45,000.

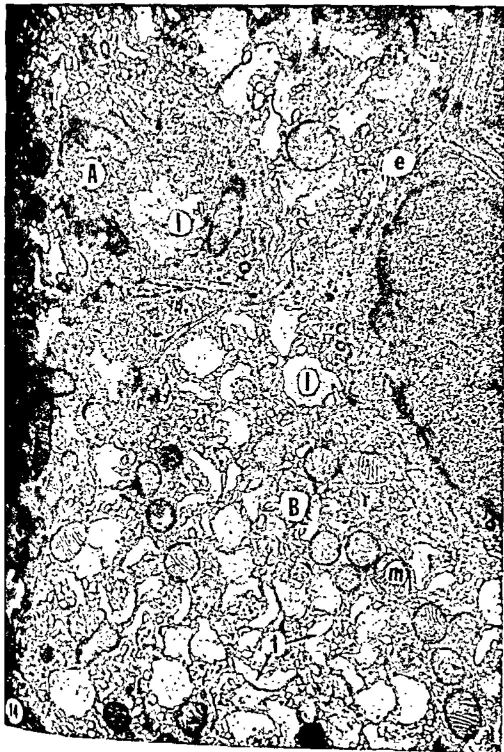

A
l
e
l
B
m
f
14

PLATE 6

EXPLANATION OF FIGURE

14 Cluster of interstitial cells from an immature rabbit of about five months in various stages of differentiation. Less differentiated cells (e.g. A) are characterized by the relative abundance of ribosomes and a paucity of lipid droplets and vesicles and by elongated mitochondria with simple cristae. More advanced cells (e.g. B) are characterized by an abundance of irregular lipid droplets, by many small endoplasmic vesicles also containing electron dense material, by spherical mitochondria with complex cristae, and by widely scattered and less dense accumulations of ribosomes and glycogen. The irregularity of the lipid droplets, presumably recently formed droplets, suggests that they may arise within dilated cisternae of the endoplasmic reticulum; a continuous and distinct limiting membrane is suggested here but is not always easy to define. In less differentiated cells (as A) in which there is little endoplasmic reticulum the lipid droplets are highly irregular and appear to arise freely within the cytoplasm with no limiting membrane. × 45,000.

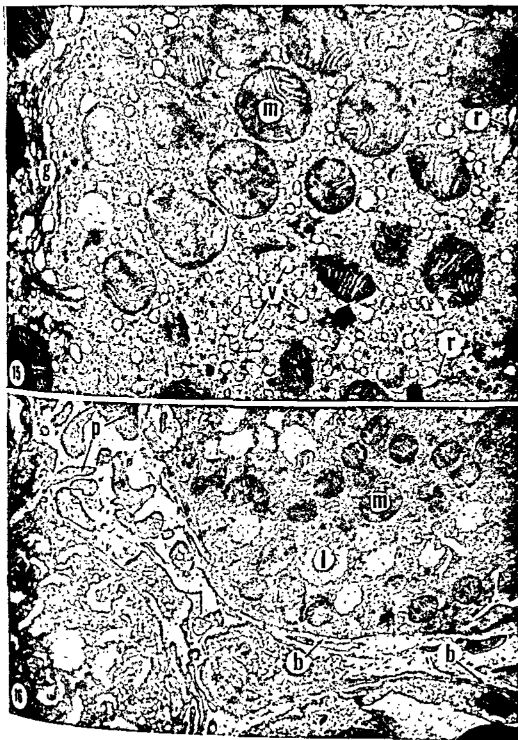

m
r
g
v
r
15
p
e
m
l
b
b
16

PLATE 7

EXPLANATION OF FIGURES

15 Mature interstitial cell of non-pregnant ("estrous") rabbit. This is typical "light" cell, as shown in figure 5. The Golgi area is prominent. Its component cisternae are dilated peripherally, forming vesicles with electron dense contents similar to those found elsewhere within the cytoplasm. The latter vary in size from about 500 to 1200 Angstroms. In some areas (e.g. r) there is direct continuity between rough endoplasmic reticulum and the smooth-walled vesicles. The mitochondria are typical of mature interstitial cells. The cristae are disposed in groups of parallel membranes, one group often being at an angle to another. There is a dense finely granular matrix. No large lipid droplets are shown in this figure though they are invariably present in mature interstitial cells. × 65,000.

16 Two "dark" cells from the "interstitial gland" of a rabbit at the tenth day of pseudopregnancy. Similar "dark" cells are found in non-pregnant animals and so far are not to be interpreted as typical of pregnancy or pseudopregnancy. They are characterized by a closely packed network of smooth-walled endoplasmic reticulum in the form of compressed vesicles or short tubules. Irregular lipid droplets and mitochondria are scattered throughout the cytoplasm. An extensive extracellular space with collagen and processes of other interstitial cells is also shown. A thin discontinuous basement membrane (b) enveloped groups of interstitial cells and their processes. This is a continuous basement membrane in relation to the endothelium of the blood vessels. × 25,000.

PLATE 8

EXPLANATION OF FIGURES

17 Portion of an interstitial cell from a rabbit at the sixth day of pregnancy showing the "microtubules" of the cytoplasm. They are only seen in material fixed in glutaraldehyde. Fixation, 1.25% glutaraldehyde with phosphate buffer, osmolarity about 350 m.osmols. × 90,000.

18 *Blood vessel of the ovarian interstitial tissue and adjacent cells* of a rabbit at the thirtieth day of pregnancy. The lumen of the vessel is collapsed. The endothelial cells contain lipid droplets, endoplasmic reticulum and accumulations of monoparticulate glycogen. There is an extensive extracellular space containing wavy strands of material similar to that of basement membranes and also wisps of collagen. × 21,000.

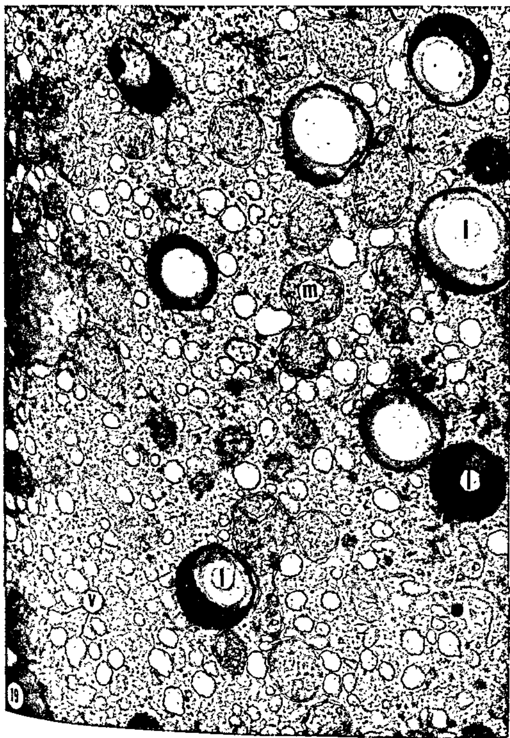

l
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l
l
v
19

PLATE 9

EXPLANATION OF FIGURE

19 Interstitial cell of rabbit seven days after parturition. The cell depicted may be degenerating and was selected to show stages suggesting the transformation of lipid droplets into electron dense bodies. A general regression of the interstitial tissue in the post-partum period is not implied by the selection of this figure. Lipid droplets are shown in which electron dense material accumulates peripherally and gradually extends deeply into the center of the droplet which also shrinks. The empty area in the center of such droplets suggests poor penetration of the fixative. Mitochondria here are somewhat simpler in their pattern of cristae than in fully active cells and there is a suggestion that they may be tubular in type. Granules, probably of glycogen, are common. × 37,500.


l
f
l'
m
l
20
f
X
m
21
X

PLATE 10

EXPLANATION OF FIGURES

20 Degenerating interstitial cell of a rabbit at the thirtieth day of pregnancy. The mitochondria are shrunken and show simplification of their pattern of cristae and increased electron density of the matrix. The formation of parallel arrays of membranes superficially resembling Golgi areas may result from the degeneration of mitochondria. Some of these areas may be early stages in the formation of "myelin figures" (v. fig. 21). The lipid droplets vary in form from irregular masses of slight electron density (lower right) to rounded droplets having a peripheral condensation of very electron dense material (upper left). The small vesicles of the endoplasmic reticulum appear empty for the most part. × 21,000.

21 Degenerating interstitial cell of the ovary of a non-pregnant ("estrous") rabbit. Such cells may be observed throughout the ovary of mature rabbits at any stage of the reproductive life (see figs. 7, 8). Here there is almost complete disorganization of the fine structure of the cell. Remnants of mitochondria may be recognized. "Myelin figures," apparently formed at the expense of the endoplasmic reticulum (f), are common and varied in morphology. Accumulations of extremely electron dense granules, larger than those assumed in previous figures to represent glycogen, are frequent; they lie free in the cytoplasm without limiting membranes (x). × 24,000.

Fluid Flow in the Liver Demonstrated with Horse Radish Peroxidase

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ABSTRACT Using horse radish peroxidase as a tracer for fluid flow, rapid diffusion was demonstrated between sinusoidal lumina and the interhepatocytic spaces of the murine liver. The marker passed rapidly between adjacent sinusoidal cells and filled the space of Disse. Appreciable amounts of peroxidase were also seen in pinocytic and phagocytic vacuoles within sinusoidal cells. Many junctions in the wall of bile canaliculi impeded the passage of the marker, but in some sites this hindrance did not occur. This provides a path for the direct exchange of material between vascular and biliary contents. Such a mechanism may be physiologically controlled. Junctions in bile ductules prevent diffusion of horse radish peroxidase.

For over a century the attention of morphologists and physiologists has been directed to the peculiarities of blood and fluid flow in the vertebrate liver. With the development of modern experimental techniques some of these problems are slowly being resolved. In this report ultrastructural histochemical techniques have been employed to trace the flow of fluid in the liver of the albino mouse; horse radish peroxidase (HRP) was used as the marker. This small haemoprotein has a molecular weight of approximately 40,000 (Keilin and Hartree, '65) which, in comparison with other plasma proteins, provides a more realistic picture of fluid flow than the use of larger, electron-dense markers such as ferritin. Horse radish peroxidase was first used as a marker by Straus ('57); it was later developed for ultrastructural-histochemical techniques by Karnovsky ('65a).

Horse radish peroxidase activity can be demonstrated by its interaction with 3 3′ amino-benzidine and H_2O_2. The product of this reaction combines with osmium tetroxide and is deposited as electron-dense, easily seen material.

MATERIALS AND METHODS

Prince Henry albino mice weighing between 30–35 gm were injected intravenously with 3.0 mg of horse radish peroxidase dissolved in 0.5 ml of 0.9% saline. Hepatic biopsies were taken at 1, 2, 5, 10, 15 and 30 minute intervals. Specimens from at least three mice were removed at each time interval. Small blocks of tissue were immersed in cold 6.25% glutaraldehyde in 0.1 M phosphate buffer (pH 7.5) and allowed to fix for four hours; they were then washed overnight in 0.1 M phosphate buffer containing 5% sucrose.

Sections 40 μ in thickness were cut on a freezing microtome and the tissue processed by the method of Graham and Karnovsky ('66). The sections were incubated for five to ten minutes in a saturated solution of 3, 3′ diaminobenzidine in 0.05 M Tris HCl buffer (pH 7.6) containing 0.01% H_2O_2. After washing in distilled water the sections were fixed in 1% osmium tetroxide in phosphate buffer (pH 7.3) for 90 minutes.

At the time of biopsy other portions of liver were immersed directly in osmium tetroxide in phosphate buffer (pH 7.3) and allowed to fix for 90 minutes. These were used as control examples of normal ultrastructure. After osmication the tissues were dehydrated in graded solutions of ethanol and embedded in Araldite according to the method of Luft ('61). Sections were cut with an LKB ultramicrotome and examined on a JEM T6 electron microscope. Mice injected with HRP served as controls.

OBSERVATIONS

A. *Livers from uninjected mice.* The sinusoidal cell layer is discontinuous, the gaps between adjacent cells measuring

passed (Farquhar and Palade, '63). It would appear that most of the intercellular space in the liver is available for rapid diffusion and exchange of substances. It has been suggested that this intercellular space may serve as a channel for conveying a protein-free filtrate of plasma from the hepatic sinusoids to the bile canaliculi (Ashworth and Sanders, '60). The findings in this study add some support to this belief. An intercellular aqueous pathway has been suggested for other tissues such as the choroid plexus (Maxwell and Pease, '56), ciliary epithelium of the eye (Pappas et al., '59), the kidney (Pease, '56; Sjöstrand and Rhodin, '53), and the cornea (Kaye et al., '62).

Most bile canaliculi were free of the tracer. This may be due to the terminal bars of the interhepatocytic space situated along the length of the canaliculi (Carruthers and Steiner, '62); these may be similar to the zonulae occludentes described by Farquhar and Palade ('63). Such structures are said to be the sites of maximal constraint to the progress of diffusion in epithelial tissues. A few bile canaliculi, however, did exhibit horse radish peroxidase activity in their lumina, indicating that in some instances diffusion beyond the junction (terminal bar) can occur. Such a phenomenon may be due to faults in junctional regions and may depend on the physiological state of the liver and be temporary rather than permanent. Karnovsky ('65) has demonstrated the passage of the protein tracer through the junctions of vascular endothelium in myocardium. Cerebral capillaries, however, withstood the passage of peroxidase (Reese and Karnovsky, '67). Florey ('64) showed the entry of saccharated iron oxide into similar junctions in vascular endothelium, although it was uncommon. These findings indicate that such junctions may be patent under some physiological condition; factors, as yet unknown, probably govern the patency of these junctions. On the other hand, morphologically similar structures may perhaps be functionally dissimilar in different cell types; this is suggested by the differences between the junctions of cerebral capillaries and those of myocardium and striated muscle. Moreover, Graham and Karnovsky ('66) failed to demonstrate entry of HRP into the luminal aspect of the intercellular space of renal tubular epithelium; such entry was probably impeded by the apical junction. Essentially similar results were obtained by Becker et al. ('67) for the choroid plexus. Thus the weight of evidence is that, with the exception of vascular endothelium in heart and skeletal muscle, zonular junctions generally impede diffusion of molecules of the size of horse radish peroxidase; small molecules such as water and electrolytes need not necessarily be hindered, this still remaining unproven. The apical junctions of ductular epithelium also obstruct the passage of HRP. We could not conclusively demonstrate the type of junctions in the walls of bile canaliculi, and a differentiation between zonula adherens and a zonula occludens could not be made.

The possibility of some dissolution of the intercellular material by the enzymatic effect of HRP needs also to be considered. Information on this is, however, lacking but, considering the data cited above, any such reaction would appear to be dependent on the type of junction encountered.

The disappearance of HRP from the extracellular space within 30 minutes indicates a pathway for rapid clearance. This may be accomplished by lymphatic drainage (Hampton, '58) and by drainage into a few selected bile canaliculi in which the patency of some junctions provides a means of direct exchange between the vascular and biliary compartments. This pathway may be under physiological control and may be disturbed in disease. The existence of connections between the space of Disse and bile canaliculi has been suggested by some electron microscopists (Rouiller, '54, '56; Cossel, '62; Schaffner and Popper, '59a,b). Rouiller and Simon ('62) and more recently Yodaiken ('66) have reaffirmed this, but state that the occurrence of such channels was rare. Other workers deny their presence. (Steiner, '61; Steiner et al., '61; Novikoff and Essner, '60; Goldfischer et al., '62). Although retrograde injection of colloidal particles did not demonstrate such a pathway (Hampton, '58), the size of the particles used was probably too great when compared with the relatively small molecular weight of

0.01–1 μ (fig. 1). In the space of Disse discontinuous strands of osmiophilic material reminiscent of the basement membrane of vascular endothelium are found (fig. 1). A few cells lining the sinusoidal wall also exhibit tight junctions of the type seen in vascular endothelium (fig. 2). Occasional fenestrae occur in some of the sinusoidal cells (fig. 2). These measure approximately 0.1 μ in diameter; those cells having them are unsupported by a basement membrane.

The ultrastructural morphology of hepatocytes resembles closely that described by various authors (Dalton et al., '50; Daems, '61; Novikoff and Essner, '60; Bruni and Porter, '65).

B. Livers from mice injected with horse radish peroxidase. Within one minute the reaction product is present in hepatic sinusoids and between adjacent sinusoidal cells. Sites of HRP activity are easily distinguished by their electron density. By two minutes activity occurs in pinocytic and phagocytic vacuoles within the cytoplasm of Kupffer cells. Small amounts of reaction product are also present in the space of Disse (figs. 3, 4).

After five minutes the marker completely fills the space of Disse and outlines the hepatocytic microvilli. At the sinusoidal border of the hepatocyte small pinocytic vesicles containing the tracer are forming (figs. 3, 5).

Reaction product is also present in the inter-cellular space between adjoining hepatocytes and is directly continuous with the material in the subsinusoidal space of Disse (fig. 5). The tracer is not uniformly distributed in the interhepatocytic space; thin strands measuring 40–50 Å in thickness and displaying marker activity are seen.

Despite the widespread distribution of reaction product in the interhepatocytic space, many of the bile canaliculi do not contain the tracer which appears to be impeded by the junctional region (terminal bar) of the canaliculus (fig. 6). This, however, is not invariable and several bile canaliculi do contain the marker which is directly continuous with that in the interhepatocytic space (figs. 7, 8).

After ten minutes the tracer lies within the periductular space between bile ductular cells and hepatocytes; once again it oc curs in the intercellular space betweer adjacent ductular epithelial cells (fig. 9) The basement membrane at the base o the bile ductules does not impede diffusior from the periductular space into the inter epithelial space. However, there is n tracer in the lumen of the duct.

At 30 minutes the marker is absent fron the intercellular spaces of the liver an the reaction product is confined solely t phagocytic vacuoles in Kupffer cells an to a few hepatocytic lysosomes.

DISCUSSION

On structural grounds it is unlikely tha the discontinuous sinusoidal wall offer any resistance to the flow of fluid fror within the sinusoidal space to the spac of Disse. It is also unlikely that the fra mented basement membrane in the spac of Disse is a source of impedence. This i demonstrated by the rapidity of appea ance of HRP activity in the intercellula areas between the sinusoidal cells and i the space of Disse. Data from isotopi studies also confirm that the walls of th sinusoids are freely permeable to plasm protein (McCarrel and Drinker, '41).

Although peroxidase passed rapidly be tween adjacent sinusoidal cells we coul not conclusively ascertain whether th sparse intercellular tight junctions in th sinusoidal wall impede the diffusion o HRP. Fenestrae have been previously de scribed in the vascular endothelium of th endocrine glands, in structures engage in rapid exchange of fluids (such as th renal glomerulus and the choroid plexus and also in retia mirabilia (Majno, '65 but have also been reported in the liver of normal rats (Burkel and Low, '66) an in the connective tissue proliferation du ing extra-hepatic cholestasis (Carruther et al., '62). Whether fenestrae in hepati sinusoids are permeable to the flow o fluid could not be established.

Diffusion of marker into the interhepa tocytic space is rapid and unimpeded. Thi is not unexpected, for, apart from bil canaliculi, ultrastructural studies hav shown only maculae adherens or desmo somes between adjoining hepatocyte These are circumscribed areas of intercel lular attachment and can be easily by

assed (Farquhar and Palade, '63). It would appear that most of the intercellular space in the liver is available for rapid diffusion and exchange of substances. It has been suggested that this intercellular space may serve as a channel for conveying a protein-free filtrate of plasma from the hepatic sinusoids to the bile canaliculi (Ashworth and Sanders, '60). The findings in this study add some support to this belief. An intercellular aqueous pathway has been suggested for other tissues such as the choroid plexus (Maxwell and Pease, '56), ciliary epithelium of the eye (Pappas et al., '59), the kidney (Pease, '56; Sjöstrand and Rhodin, '53), and the cornea (Kaye et al., '62).

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horse radish peroxidase. The present results would favour the existence of these intercommunicating channels.

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PLATE 1

EXPLANATION OF FIGURES

1 Liver from uninjected control. Cells lining the sinusoidal lumen (SIN) exhibit the presence of intercellular gaps (blank arrows). In the space of Disse fibrillar material (X) is seen. The hepatocyte (HEP) does not show any abnormalities. Mag. × 28,000.

2 Liver from uninjected control. The cells lining the sinusoidal lumen (SIN) show in some instances tight junctions (J) and fenestrae (black arrows). Mag. × 24,000.

SIN

X

HEP

1

SIN

J

J

2

PLATE 1

EXPLANATION OF FIGURES

1 Liver from uninjected control. Cells lining the sinusoidal lumen (SIN) exhibit the presence of intercellular gaps (blank arrows). In the space of Disse fibrillar material (X) is seen. The hepatocyte (HEP) does not show any abnormalities. Mag. × 28,000.

2 Liver from uninjected control. The cells lining the sinusoidal lumen (SIN) show in some instances tight junctions (J) and fenestrae (black arrows). Mag. × 24,000.

KUP
HEP
3

4

PLATE 2

EXPLANATION OF FIGURES

3 Liver five minutes after the injection of horse radish peroxidase. The electron dense deposits indicate areas of peroxidase activity. Reaction product is seen in micropinocytic vacuoles (black arrows) of both Kupffer cells (KUP) and hepatocytes (HEP). It is present in the junction (J) between adjacent sinusoidal cells and has entered the space of Disse (blank arrow). Mag. × 51,000.

4 Kupffer cell two minutes after injection of horse radish peroxidase. Reaction product is present in small micropinocytic vacuoles (blank arrows) and in larger vacuoles (V). Mag. × 40,500.

KUPC
HEP
5

BC
6

PLATE 3

EXPLANATION OF FIGURES

5 Liver five minutes after injection of horse radish peroxidase. The marker has entered into the space of Disse (blank arrow) and passed into the interhepatocytic space (black arrows) the reaction product is also present in a vacuole (V) of a Kupffer cell (KUP). Mag. × 41,000.

6 Bile canaliculus (BC) showing terminal bars (blank arrows) and interhepatocytic spaces (black arrows). Mag. × 48,000.





LU

PLATE 4

EXPLANATION OF FIGURES

7 Bile canaliculus five minutes after injection of horse radish peroxidase. Reaction product is seen in the bile canaliculus (blank arrows) between adjacent hepatocytes (HEP). This is continuous with the marker in the interhepatocytic space (black arrows). Mag. × 24,000.

8 Bile canaliculus five minutes after injection of horse radish peroxidase. The bile canaliculus (blank arrow) between adjacent hepatocytes (HEP) contains reaction product which is continuous with similar material in the interhepatocytic space (black arrows). Mag. × 40,000.

9 Bile ductule ten minutes after injection of horse radish peroxidase. The marker is seen in the periductular space (black arrow) and passing between adjacent ductular cells (blank arrows). It does not, however, enter the lumen of the dutcule (LU). Mag. × 28,000.

Marginal Zone and Germinal Center Development in the Spleens of Neonatally Thymectomized and Nonthymectomized Young Rats [1]

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ABSTRACT Spleens from neonatally thymectomized and nonthymectomized young rats were studied histologically and histochemically to elucidate the development of the splenic immune system with and without thymus.

In intact animals primitive germinal center activity could be elicited with antigen as early as 13 days of age. More definitive germinal centers lacking tingible body macrophages were observed at 18 days of age. Germinal centers containing tingible body macrophages did not develop until 35 days of age in response to antigenic stimulation. This coincided with maximal development of the marginal zone of medium-sized lymphocytes and the mature development of nodular macrophages possessing strong acid phosphatase activity.

Neonatally thymectomized rats developed marginal zones and germinal centers similar to control littermates when the young animals were maintained on tetracycline. Thymectomized animals not given tetracycline showed disturbances in splenic development. These are discussed.

The results suggest that the thymus may be critical to the immune system in rats from birth to about 30 days of age but is not essential to its function beyond this period. Marginal zone lymphocytes and germinal center cells proliferate normally and mature to the plasma cell stage in the absence of a thymus if the animals are maintained on tetracycline beyond this critical age.

Histological and histochemical evidence from our laboratory implicates the medium-sized, marginal zone lymphocytes as a major source of germinal center cells following antigenic stimulation (Pettersen, Borgen and Graupner, '67).

The present study was undertaken to determine when marginal zone lymphocytes migrate to the spleen, when they respond to antigenic stimulation and whether the thymus influences their development or maintenance.

MATERIALS AND METHODS

Fifty young Holtzman rats were used to study the development of the marginal zone and the time of appearance of germinal center activity in the spleen. Animals ranged from one hour to 64 days of age at autopsy and include some which received lateral tail vein injections of typhoid-paratyphoid vaccine (Eli Lilly and Co.) or of 055:B5 E. *coli* endotoxin (Difco) at representative ages from 13 to 60 days. Intraperitoneal or intramediastinal injections were given to animals younger than 13 days of age.

Another series of rats was thymectomized on the day of birth. They were anesthetized by cooling in a freezer, the sternums were split with small surgical scissors and the thymic lobes were removed with forceps using clean but not sterile technique. Thirty percent of the animals died during or shortly following surgery. Forty-four animals survived the initial procedure.

At autopsy 12 animals were free of thymus and 13 had portions remaining. Nineteen died of runt disease or were victims of maternal cannabalism. Tetracycline hydrochloride (Pfizer Corp.) in the drinking water eliminated runting and was used routinely following a high mortality in our first four litters. Histological material was obtained from only three thymectomized animals not receiving the antibiotic. Two of these were runted animals 27 days of

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mals were not available in the 30 to 60 day age group.

Thymectomized animals. Neonatal thymectomy retarded the development of the marginal zone in the spleens of animals not maintained on tetracycline. Figure 7 illustrates this in a spleen section stained for acid phosphatase activity from a 25 day old, non-runted, thymectomized animal. Figure 8 is a section from a littermate control. Runted animals lacked marginal zones (fig. 9).

Small lymphocytes were deficient but not entirely lacking in these spleens. Runted animals also showed decreased numbers of medium-sized lymphocytes (fig. 12). These latter cells were present in normal numbers in the spleen of the non-runted animal (fig. 11).

Spleens from thymectomized animals maintained on tetracycline developed marginal zones almost as rapidly as control animals and contained significant although reduced numbers of small lymphocytes. Cellular responses to antigenic stimulation were indistinguishable from control responses except for decreased numbers of mantle layer small lymphocytes. Recovery of the marginal zone population of medium-sized lymphocytes following initial depletion in response to antigenic stimulation was normal in thymectomized animals. Plasma cell proliferation was unaffected.

The 11 month old thymectomized animal responded like its control to antigen administered five hours before autopsy. Figure 16 shows a partial collapse of the marginal zone on a section from this spleen stained for macrophages. An additional thymectomized animal of this age was not available to test whether repopulation of the marginal zone occurs by four days following stimulation.

DISCUSSION

Two types of lymphocytes can be distinguished in the young lymphoid areas of the rat spleen. The first of these to enter the spleen is a medium-sized, slightly pyroninophilic lymphocyte that initially occupies a position adjacent to the arterial vessels (fig. 2). These are later located more peripherally as the second type, a small non-pyroninophilic lymphocyte, replaces them around the vessesl (fig. 4). There is some intermingling of the two types in spleen sections from animals less than ten days of age. By 13 days of age there is a definitive marginal zone (figs. 3, 4) of medium-sized lymphocytes enveloping an inner core of small lymphocytes. This relationship is disturbed within one day following the intravenous administration of antigen with suggestions of immature germinal center formation. Medium-sized lymphocytes invade the small lymphocyte mass and differentiation into hemocytoblasts occurs. Figures 5 and 6 illustrate splenic nodules from a control and an experimental animal 20 days of age. No plasma cell formation occurred at this age when animals were killed at later stages following antigenic stimulation.

Halliday ('56) reported an abrupt cessation of the ability of young rats to absorb antibodies through the gut wall at 20 days of age. This phenomenon was unrelated to the uptake of solid food. It was also shown (Halliday, '57) that young rats could produce antibodies in response to active immunization prior to the loss in absorptive capacity. Evidence indicated that these antibodies were located in the β-globulin fraction. Our results in young animals show that cellular changes occur in rats younger than 20 days of age following active immunization. These changes are similar to the response in adult spleens in certain respects. Marginal zone lymphocytes are the source of germinal center cells in both young and adult spleens and mitotic activity and hemocytoblast formation occur in both. Germinal centers in young spleens, however, did not contain tingible body macrophages and did not produce plasma cells. Additional studies in the 10–30 day age group employing immunochemical techniques would be desirable to determine what cell type produces antibodies during this stage of development. Our sections stained histochemically for macrophage distribution suggest that a follicular antigen trapping mechanism as described by Miller and Nossal ('64) in rat popliteal lymph nodes and by Hunter ('66) in the rat spleen may not be functional in young rats because of the paucity of macrophages in the white pulp at this age. MacFadden ('68) has studied the

age and the third was a 25 day old non-runted animal.

The 12 totally thymectomized animals were grouped with littermate controls and partially thymectomized animals of the same age. Some of the animals received typhoid-paratyphoid vaccine via the lateral tail vein. The injections were generally given five days before autopsy to allow a maximum cellular response to develop. Groups of animals were killed at 25, 27, 35, 41 and 64 days of age. One thymectomized animal was killed at 11 months of age five hours following antigenic stimulation. Histological material was obtained from 40 animals in this portion of the study.

All animals were killed with ether, spleens were rapidly removed and pieces fixed in Carnoy's fluid, in Helly's fluid and in 15% cold, neutral formalin. All suspicious mediastinal tissue was removed from the surgical animals and fixed in 15% formalin. The thorax was then held under running tap water to wash it free of blood for further inspection under a hand lens.

Tissue fixed in formalin was stained for acid phosphatase activity as previously described (Pettersen, '64). These sections were studied for macrophage distribution and general splenic topography. Mediastinal tissue stained in this manner was used to identify thymic remnants.

Tissue fixed in Helly's fluid was sectioned in paraffin at 5 μ and stained with hematoxylin-eosin-azure II or with hematoxylin and eosin. Carnoy-fixed tissue was sectioned in paraffin at 5 μ and stained with methyl green-pyronin.

RESULTS

Splenic development in nonthymectomized animals. Spleens of newborns were largely myeloid with an occasional small sheath of lymphocytes surrounding the arterial vessels. Macrophages were dispersed throughout the myelopoietic areas but were not present in the small lymphoid areas (fig. 1). At three days of age several tiers of lymphocytes surrounded the arterioles (fig. 2). These cells resembled the medium-sized lymphocytes found in the marginal zones of adult rats. Suggestions of a marginal zone were present at nine days of age and it was definitive by 13 days of ag (fig. 3). Small lymphocytes were then pres ent in a position adjacent to the arteriole with medium-sized lymphocytes peripher to them (fig. 4). A disturbance in this re lationship occurred one day following travenous injections of antigen in 13 da old animals. The two types of lymphocyte were commingled rather than segregate in these spleens. This phenomenon wa suggested in animals eight or nine day of age following intraperitoneal injection but was difficult to assess because segreg tion was not complete in control spleens o this age. Figures 5 and 6 illustrate spleni nodules from a control and an injecte animal 20 days of age.

Germinal centers developed in 18 da old rats following intravenous injection of 0.25 ml typhoid-paratyphoid vaccine a 14 days of age. They consisted of smal nests of pyroninophilic cells surrounded b mantle layers of non-pyroninophilic smal lymphocytes. Tingible body macrophage were not present and no plasma cells coul be identified. About 30% of the lymphoi nodules contained such germinal center.

The marginal zone was the source o cells for early germinal centers. The widtl of this rim of cells reached adult size (8 100 μ) by 35 days of age at which tim tingible body macrophages were present i germinal centers of those spleens expose to antigen for five days. About 20% of th nodules contained germinal centers witl these macrophages (fig. 15). Up to thi age tingible bodies were not present in re sponse to antigen but there were increase in numbers of macrophages possessin acid phosphatase activity in the lymphoi areas of both stimulated and non-stimu lated spleens. In 60 day old animals nearl 100% of the lymphoid nodules responded to similar antigenic stimulation by the formation of germinal centers containin tingible body macrophages.

Our data were not complete enough t establish the age of appearance of plasm cells. Sixty day old rats contained larg numbers of plasma cells in the splenic red pulp five days following stimulation Efforts to induce a similar response in 2 30 day old rats were negative. Sufficien numbers of antigenically stimulated an

mals were not available in the 30 to 60 day age group.

Thymectomized animals. Neonatal thymectomy retarded the development of the marginal zone in the spleens of animals not maintained on tetracycline. Figure 7 illustrates this in a spleen section stained for acid phosphatase activity from a 25 day old, non-runted, thymectomized animal. Figure 8 is a section from a littermate control. Runted animals lacked marginal zones (fig. 9).

Small lymphocytes were deficient but not entirely lacking in these spleens. Runted animals also showed decreased numbers of medium-sized lymphocytes (fig. 12). These latter cells were present in normal numbers in the spleen of the nonrunted animal (fig. 11).

Spleens from thymectomized animals maintained on tetracycline developed marginal zones almost as rapidly as control animals and contained significant although reduced numbers of small lymphocytes. Cellular responses to antigenic stimulation were indistinguishable from control responses except for decreased numbers of mantle layer small lymphocytes. Recovery of the marginal zone population of medium-sized lymphocytes following initial depletion in response to antigenic stimulation was normal in thymectomized animals. Plasma cell proliferation was unaffected.

The 11 month old thymectomized animal responded like its control to antigen administered five hours before autopsy. Figure 16 shows a partial collapse of the marginal zone on a section from this spleen stained for macrophages. An additional thymectomized animal of this age was not available to test whether repopulation of the marginal zone occurs by four days following stimulation.

DISCUSSION

Two types of lymphocytes can be distinguished in the young lymphoid areas of the rat spleen. The first of these to enter the spleen is a medium-sized, slightly pyroninophilic lymphocyte that initially occupies a position adjacent to the arterial vessels (fig. 2). These are later located more peripherally as the second type, a small non-pyroninophilic lymphocyte, replaces them around the vessesl (fig. 4). There is some intermingling of the two types in spleen sections from animals less than ten days of age. By 13 days of age there is a definitive marginal zone (figs. 3, 4) of medium-sized lymphocytes enveloping an inner core of small lymphocytes. This relationship is disturbed within one day following the intravenous administration of antigen with suggestions of immature germinal center formation. Medium-sized lymphocytes invade the small lymphocyte mass and differentiation into hemocytoblasts occurs. Figures 5 and 6 illustrate splenic nodules from a control and an experimental animal 20 days of age. No plasma cell formation occurred at this age when animals were killed at later stages following antigenic stimulation.

Halliday ('56) reported an abrupt cessation of the ability of young rats to absorb antibodies through the gut wall at 20 days of age. This phenomenon was unrelated to the uptake of solid food. It was also shown (Halliday, '57) that young rats could produce antibodies in response to active immunization prior to the loss in absorptive capacity. Evidence indicated that these antibodies were located in the β-globulin fraction. Our results in young animals show that cellular changes occur in rats younger than 20 days of age following active immunization. These changes are similar to the response in adult spleens in certain respects. Marginal zone lymphocytes are the source of germinal center cells in both young and adult spleens and mitotic activity and hemocytoblast formation occur in both. Germinal centers in young spleens, however, did not contain tingible body macrophages and did not produce plasma cells. Additional studies in the 10–30 day age group employing immunochemical techniques would be desirable to determine what cell type produces antibodies during this stage of development. Our sections stained histochemically for macrophage distribution suggest that a follicular antigen trapping mechanism as described by Miller and Nossal ('64) in rat popliteal lymph nodes and by Hunter ('66) in the rat spleen may not be functional in young rats because of the paucity of macrophages in the white pulp at this age. MacFadden ('68) has studied the

phagocytic function of macrophages in the spleens of late fetal (17–18 days of gestation) to old (475 days) rats. Increased numbers of white pulp macrophages possessing phagocytic ability and hydrolytic enzyme activity were reported as the animals aged. Our results support his conclusions but also suggest that antigenic stimulation enhances the maturation of white pulp macrophages. Bauer et al. ('66) reported that the intracellular digestion of antigen was slower in macrophages of germfree versus conventional mice. Spleen sections from several germfree rats were stained for acid phosphatase activity in our laboratory. The white pulp contained relatively few cells exhibiting enzyme activity compared to spleen sections from untreated adult control rats. The red pulp did not differ from that of the controls. This suggests that ageing is not necessarily the variable in the maturation of white pulp macrophages but that some exogenous stimulation is required. Red pulp macrophages possess hydrolytic enzyme activity in the late fetal stages of development (MacFadden, '68) before such stimulation is likely to occur.

Williams and Nossal ('66) studied the antigen-trapping ability of the lymphoid tissues in young rats following footpad injections of I^{125}-labelled polymerized flagellin from *Salmonella adalaide*. Autoradiographs showed initial signs of cortical localization in lymph nodes from animals between 10–14 days of age. True follicular retention did not occur until animals were four to six days older at the time of injection. Antigen was not retained by the lymphoid nodules of the spleen six days following an injection given at four weeks of age. An adult pattern of retention was observed in the spleen in animals injected at six weeks of age. The ability to retain antigen increased five fold per unit weight of lymphoid tissue between two and six weeks of age. Williams ('66) demonstrated a parallel development of the antigen-trapping mechanism and the ability to form antibodies.

White pulp macrophages may mature from reticular cells already present in the splenic anlage at the time lymphoid immigration begins. Their specialization into antigen-processing cells may be influenced by their lymphoid environment and triggered upon initial exposure to antigen. Pettersen ('64) reported that the number of cells in the white pulp reactive to Marshall's silver impregnation stain for macrophages does not increase following antigenic stimulation but the number of cells with active hydrolytic enzyme systems does increase following such treatment. This indicates that increases in macrophages represent maturation of existing cells. It is difficult to explain how macrophages get into the developing lymphoid areas. They may be incorporated from the red pulp as the lymphoid cells migrate into the spleen. This explanation is inconsistent with the following observations: (1) during development macrophages pile up at the marginal zone-red pulp junction but are rarely found in the marginal zone (fig. 3); (2) the marginal metalophils (Snook, '64) appear to migrate outward as the small lymphocytes move into the periarteriolar regions; (3) white pulp macrophages in newborns are not selectively localized near the periphery of the lymphoid masses; (4) macrophages in the red pulp possess hydrolytic enzyme activity at or before birth, whereas white pulp macrophages acquire this activity later in development.

Alternatively, one can postulate that the white pulp macrophages represent a breed that is different from those in the red pulp. They may migrate in with the lymphoid cells during development or may be products of maturation of certain of these cells. The data available suggest some form of this alternative hypothesis.

The retarded development of the marginal zone in young neonatally thymectomized rats not maintained on tetracycline may be related to deficiencies of small lymphocytes of thymic origin or to infectious processes that keep the marginal zone lymphocytes in a state of disorganization. There were no germinal centers and no hemocytoblasts in the white pulp of these animals to suggest a cellular response to invading organisms. Azar ('64) substantially reduced the incidence of runting in neonatally thymectomized rats by adding oxytetracycline HCl (Pfizer Corp.) to the drinking water. He believed that bronchopulmonary infections were the cause of

death in runted animals. Non-hemolytic streptococcus, a Hemophilus organism and diphtheroids were cultured from bronchopulmonary tissue. The spleens were atrophic, lacked well-formed follicles and contained no plasma cells. Plasma cells were present in the bronchopulmonary lesions and the serum gamma globulin level was increased. This suggests that the spleen was not involved in an immunological process in runted animals but that cells that normally would develop within the spleen were employed at the site of infection. This may be a possible explanation for the deficiency of medium-sized lymphocytes in spleens of our runted animals.

Schriever, Hsu and Azar ('67) found normal plasma cell formation and normal numbers of cells containing gamma globulin in the mesenteric lymph nodes of neonatally thymectomized animals. They concluded that, in their experience, "should neonatally thymectomized rats survive the first critical eight to ten weeks of life without succumbing to infection, they will then adequately cope with subsequent infections and compensate for the loss of thymus function."

Waksman, Arnason and Jankovic ('64) reported a complete absence of small lymphocytes in spleens of neonatally thymectomized rats. None of our spleens, with the exception of those from the two runted animals, showed small lymphocyte deficiencies of this magnitude. Tetracycline may have potentiated the development of small lymphocytes or may have permitted their development under less antigenically stressful conditions.

It is clear that the thymus is not essential in maintaining a marginal zone cell population in young adult rats once the splenic immune system has developed. It cannot be concluded from our data that the thymus is not the stem source of marginal zone medium-sized lymphocytes. The following observations are consistent with a thymic origin for these cells: (1) medium-sized lymphocytes appear in the spleen before small lymphocytes and can be found in spleens of one hour old rats corresponding to the time of our earliest thymectomies; (2) a 25 day old non-runted thymectomized animal showed retarded development of the marginal zone and two 27 day old runted animals showed near absence of this region; (3) 35 day old neonatally thymectomized animals maintained on tetracycline showed subtle but distinguishable differences in the ability to repopulate the marginal zone following antigenic stimulation; (4) small lymphocytes entered the spleen in significant numbers in neonatally thymectomized animals maintained on tetracycline suggesting a potentiating influence.

The following are more consistent with a nonthymic origin: (1) a spleen from a non-runted thymectomized animal not maintained on tetracycline contained a sizable population of medium-sized lymphocytes although the deficiency of small lymphocytes disturbed the usual organization of the marginal zone; (2) marginal zone lymphocytes were present in normal numbers in the neonatally thymectomized animals maintained on tetracycline; (3) the marginal zone was reconstituted after the usual depletion following antigenic stimulation.

Moore and Owen ('67) recently presented evidence that the yolk sac in the chick furnishes blood-borne stem cells which enter the thymic rudiment. A sex chromosome marker and histological techniques used to study parabiosed chick embryos showed that a sizeable proportion of thymocytes came from the partner. Evidence suggested that the same type of cell migrates into the thymic rudiment of the mouse. The cell type shown on their illustrations closely resembles those we see in young germinal centers one day following antigenic stimulation (fig. 6, arrow). We have called this cell a hemocytoblast but it is believed by us to be a transitional form of the medium-sized marginal zone lymphocytes which migrate into the germinal centers in response to antigenic stimulation.

Several conclusions seem reasonable from our data: 1. The thymus plays some essential role in survival in young conventional rats. 2. Tetracycline permits normal development of the lymphoid structure in spleens of neonatally thymectomized rats except for relative deficiencies of small lymphocytes. 3. An intact thymus is not essential in maintaining these lymphoid structures once adult development has

phagocytic function of macrophages in the spleens of late fetal (17–18 days of gestation) to old (475 days) rats. Increased numbers of white pulp macrophages possessing phagocytic ability and hydrolytic enzyme activity were reported as the animals aged. Our results support his conclusions but also suggest that antigenic stimulation enhances the maturation of white pulp macrophages. Bauer et al. ('66) reported that the intracellular digestion of antigen was slower in macrophages of germfree versus conventional mice. Spleen sections from several germfree rats were stained for acid phosphatase activity in our laboratory. The white pulp contained relatively few cells exhibiting enzyme activity compared to spleen sections from untreated adult control rats. The red pulp did not differ from that of the controls. This suggests that ageing is not necessarily the variable in the maturation of white pulp macrophages but that some exogenous stimulation is required. Red pulp macrophages possess hydrolytic enzyme activity in the late fetal stages of development (MacFadden, '68) before such stimulation is likely to occur.

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been attained. 4. Two types of lymphocytes migrate to the spleen during development of the lymphoid structures neither of which is entirely thymus dependent following birth.

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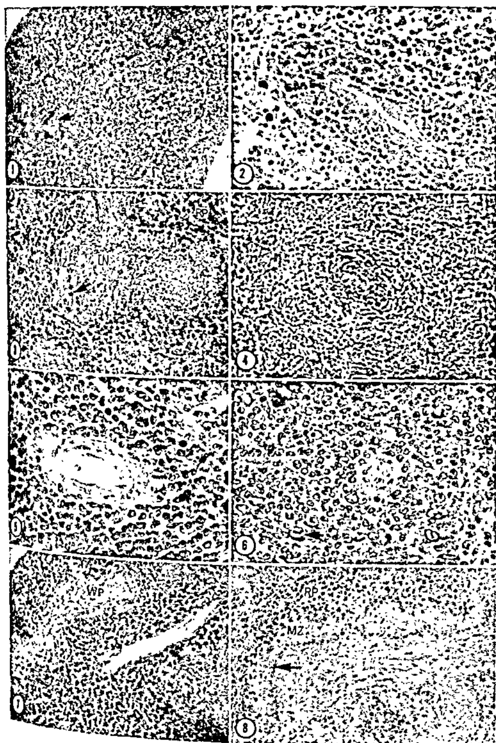

RP
MZ
LN
MZ
WP
RP
MZ

PLATE 1

EXPLANATION OF FIGURES

1 Section of spleen stained for acid phosphatase activity from a rat killed one hour after birth. Macrophages are dispersed throughout the red pulp which is entirely myeloid. Arrow identifies a small lymphoid area free of macrophages. × 70.

2 Section of spleen from a three day old rat showing a sheath of medium-sized lymphocytes surrounding an obliquely sectioned arteriole. Methyl green-pyronin. × 270.

3 Spleen section from a 13-day old rat showing developing marginal zone, MZ. A few macrophages are present in the nodules. RP, red pulp; LN, lymphoid nodule; MZ, marginal zone. Arrow identifies marginal metalophils. Acid phosphatase. × 70.

4 Spleen section from a 16 day old rat showing an inner core of small lymphocytes surrounding an arteriole and a peripheral rim of medium-sized lymphocytes occupying the marginal zone. MZ, marginal zone. Methyl green-pyronin. × 170.

5 A nodule in a spleen section from a 20 day old rat showing a core of small lymphocytes surrounding an arteriole. Methyl green-pyronin. × 400.

6 A nodule in a spleen section of a 20 day old rat one day following an intravenous injection of 0.25 ml typhoid-paratyphoid vaccine. Medium-sized lymphocytes have invaded the mass of small lymphocytes. Arrow identifies a hemocytblast. Methyl green-pyronin. × 400.

7 Section of spleen from a 25 day old, neonatally thymectomized, nonrunted rat showing retarded development of the marginal zone. WP, white pulp. Acid phosphatase. × 70.

8 Spleen section from a 25 day old littermate control. MZ, marginal zone; RP, red pulp. Arrow identifies marginal metalophils. Acid phosphatase. × 70.

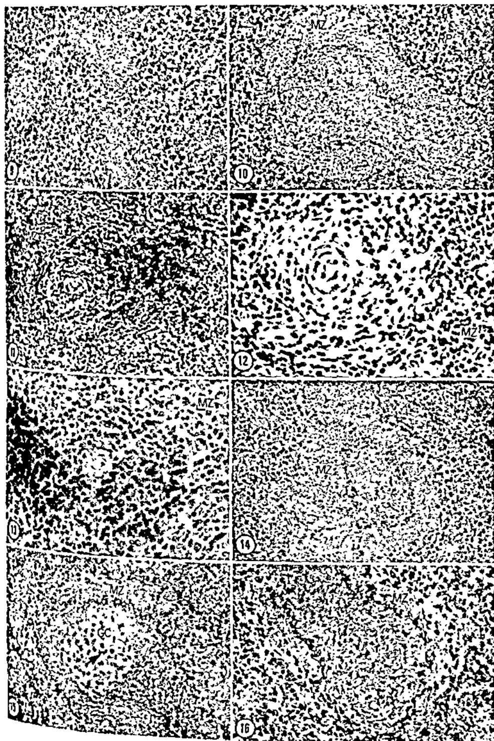

MZ
RP
10
11
12
MZ
MZ
13
MZ
14
MZ
GC
MZ
16

PLATE 2

EXPLANATION OF FIGURES

9 Spleen section from a 27 day old, thymectomized, runted rat showing poor development of lymphoid structures. Acid phosphatase. × 70.

10 Spleen section from a 27 day old littermate control. MZ, marginal zone; RP, red pulp. Acid phosphatase. × 70.

11 Splenic nodule from a 25 day old, thymectomized rat showing deficiency of small lymphocytes. Medium-sized lymphocytes show a normal population density in the marginal zone, MZ. H. and E. × 170.

12 Splenic nodule from a 27 day old, thymectomized, runted rat showing marked depletion of small lymphocytes and a marginal zone, MZ, with a low population density of medium-sized lymphocytes. H. and E. × 330.

13 Splenic nodule from a 27 day old control rat. MZ, marginal zone. Methyl green-pronin. × 330.

14 Spleen section from a 32 day old, neonatally thymectomized rat maintained on tetracycline. Marginal zone, MZ, is well developed although slightly smaller than in controls. Compare to figure 10. Acid phophatase. × 70.

15 Spleen section from a 35 day old, neonatally thymectomized rat killed five days following an intravenous injection of typhoid-paratyphoid vaccine. The marginal zone, MZ, is reconstituted, a germinal center, GC, is present in the nodule and tingible body macrophages (arrow) have developed. Acid phosphatase. × 70.

16 Ammoniacal silver-stained section of spleen from a neonatally thymectomized, 11 month old rat killed five hours after an intravenous injection of 1.0 ml typhoid-paratyphoid vaccine. The marginal zone, MZ, is constricted which is a characteristic response observed in spleens of nonthymectomized animals within one day following antigenic stimulation. × 70.

Cell Division in Injured Spinal Cord [1]

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ABSTRACT Thymidine-H^3 radioautography was used to study the proliferative response to penetrating wounds of the mouse spinal cord. In one group of animals mononuclear leukocytes which infiltrate nervous tissue wounds were labeled by injecting thymidine-H^3 prior to injury. In two other experimental groups the cells which synthesized DNA in the nervous tissue following the injury were labeled by giving either a single injection of isotope shortly before sacrifice or by giving four injections during the 24 hours prior to sacrifice. The animals were sacrificed over a five day period following spinal cord injury. Although the labeled nuclei in all three groups were similar in appearance, their distribution about the lesion was very different. The labeled blood cells were greatly concentrated at the wound, while the cells that responded to injury by DNA synthesis were much more evenly spread throughout the tissue. When these distributions were converted to straight lines and compared statistically, there was a very low probability that the group of cells labeled before injury and the two groups labeled after injury were samples from the same population. Although mononuclear leukocytes do proliferate in and around nervous tissue wounds, other cells originally present in the nervous tissue must also proliferate.

Although the cells in normal adult mammalian nervous tissue were long considered to be mitotically inert (Allen, '12; Ngowyang, '30; Ries, '38; Leblond and Walker, '56), radioautography with tritiated thymidine has provided abundant evidence for the existence in normal nervous tissue of a small population of cells which undergo DNA synthesis and mitotic division (Messier et al., '58; Hain et al., '60; Messier and Leblond, '60; Schultze and Oehlert, '60; Smart, '61; Smart and Leblond, '61; Adrian and Walker, '62; Noetzel, '62; Altman, '63; Gavrilova, '64; Noetzel and Rox, '64; Hommes and Leblond, '67). Following injury the number of such cells is greatly increased, and it has been generally assumed that these cells are glial cells (Koenig, '60; Koenig et al., '62; Altman, '62; Adrian and Walker, '62; Samorajski et al., '64; Hassler, '66). A glial proliferative response has for many years been considered to be one of the characteristic reactions of nervous tissue to injury (Adams, '58). However, by labeling hemopoietic tissues with tritiated thymidine prior to injuring the nervous tissue, it has been possible to demonstrate that many labeled cells appear around the wound. These cells were taken to consist of large numbers of mononuclear leukocytes labeled before invading the injured tissue (Adrian and Walker, '62), even though these invading cells could not be distinguished from glial cells on the basis of their appearance in paraffin sectioned material (Walker, '63). Since these invading cells have been shown to actively proliferate after entering the nervous tissue (Konigsmark and Sidman, '63; Kosunen et al., '63), the question is raised as to whether or not they can account for all the DNA synthesis that occurs in nervous tissue following injury.

Several lines of evidence have been presented which would support a position that most, if not all, of the observed DNA synthesis following local nervous tissue injury could be occurring in leukocytes. Konigsmark and Sidman ('63) demonstrated the leukocytic origin of a large percentage of the macrophages which appear at a site of nervous tissue injury. Most of the visible mitoses in injured nervous tissue occur in these macrophages, long thought to be derived from microglia (Penfield, '25; Del Rio-Hortega and Penfield, '27; Del Rio-Hortega, '32). Walker ('63) showed that although the number of labeled cells in injured nervous tissue increased with time following tritiated thymidine injection, there was no corresponding decrease in

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mersion by moving back and fourth from side to side across the section covering consecutive bands or rows of tissue. The width of each row was the same as the diameter of the oil immersion objective, 0.18 mm. The area of each row was estimated by multiplying the number of oil immersion fields in the row by $(0.18\ mm)^2$. A map was made for each section which showed the position of all radioactive nuclei, except those belonging to polymorphonuclear leukocytes. (A nucleus was considered to be radioactive if it had four or more silver grains superimposed over it.) The map was made initially on graph paper (10 × 10 per inch) by equating each oil immersion field to one 5 × 5 square. The corners of the square were observed as the field of view was changed. The borders of the section, the position of the wound, and other distinguishing features were included on each map.

To permit easier comparison of the distribution of radioactive cells in the three groups of animals, curves showing the number of radioactive cells in a band or row of tissue 0.18 mm wide extending across the width of the section were plotted as a function of the distance of the tissue band from the center of the lesion. A composite curve determined by averaging the values obtained from all the animals in the group was prepared for each day after injury.

The number of labeled cells in each two rows equidistant from and on either side of the lesion was then expressed as a ratio of the number of labeled cells in the two rows passing nearest the center of the lesion. Straight lines were fitted by the method of least squares to the ratios determined for the first five rows beyond those passing through the center of the lesion. A value for y-intercept and slope was determined for each animal. The hypothesis was made that the distribution of labeled cells (as represented by the straight lines) in the three groups of injured animals represented samples from the same population. A rank order analysis as described by Kruskal and Wallis ('52) was applied to the y-intercepts and to the slopes of the straight lines to test this hypothesis.

In a standard area of $1.25\ mm^2$ with the lesion at its center, the cell population of each spinal cord was analyzed. All nuclei of cells other than neurons, ependymal cells, and polymorphonuclear leukocytes were counted. All radioactive nuclei were classified according to their morphology. The nuclei of glial cells and of mononuclear blood cells which could not be distinguished from glial cells were classified in the following categories: (1) dark-staining round, ovoid, or irregular, (2) light-staining round, ovoid, or irregular, and (3) elongated. The light-staining nuclei were generally quite regular in shape and were larger than those of the other types. Nuclei were classified as elongated if their length was more than three times their width. Endothelial nuclei could be distinguished with considerable certainty in the material stained by the periodic-acid Schiff hematoxylin technique, and for this reason all of the nuclear classifications were done on slides stained by this method.

Sections from five of the animals in group I were chosen for mitotic counts. That part of each one of these sections lying within 3 mm from the lesion was scanned carefully under oil immersion for mitotic figures. Each mitotic figure observed was classified as to whether or not it was labeled, and its position was noted. The number of labeled mitotic figures lying within 0.7 mm from the center of the lesion and the total number of labeled cells within that same area were recorded. Likewise, the number of labeled mitotic figures between 0.7 mm and 3.0 mm from the center of the lesion and the total number of labeled cells in that area were recorded. Also, within these same two regions, the total number of mitotic figures was recorded. The χ^2 statistic as described by Dixon and Massey ('57) was used to test the hypothesis that in group I the ratio of the number of labeled mitotic figures to the total number of labeled cells within the same area is independent of their distance from the lesion. This same test was applied to the hypothesis that within a given area the ratio of the number of labeled mitotic figures to the total number of mitotic figures is independent of the distance from the lesion.

grain count, indicating an infiltration of labeled cells. Also, there was little difference in the number or in the grain count of the labeled cells regardless of whether the thymidine was given before or after injury. Since leukocytes must be virtually the only cells labeled before injury, it seemed likely that most of the cells incorporating the label after injury were also leukocytes. Finally, an experiment in which glial cells were labeled in new born mice gave no evidence for division of these cells two months later when the brain was injured (Smith and Walker, '67).

On the other hand, the observation had been made that the distribution of cells assumed to be mononuclear leukocytes around spinal cord wounds did not seem to be the same as the distribution of cells synthesizing DNA in response to injury (Adrian and Walker, '62). If these two distributions were in fact different from each other, it would seem unlikely that the two cell populations were identical. The present experiment has been designed to allowed an extensive comparison of the spatial distribution of labeled cells in injured spinal cords when labeled thymidine is injected before or after the injury. This has been done to determine whether or not the two groups of labeled cells could be samples from the same population.

MATERIALS AND METHODS

Fifty-three mice of known age were divided into three experimental groups. Group I consisted of four C57Bl adult females and seven A/J females. These animals were given three subcutaneous injections of 50 μc tritiated thymidine (specific activity 1.9 c/mm) at four hour intervals starting 24 hours before spinal cord injury. Group II consisted of 20 A/J females, each of which was given a single injection of 50 μc tritiated thymidine subcutaneously one hour before sacrifice. Group III consisted of 22 A/J females, of which each was given four subcutaneous injections of 50 μc tritiated thymidine at six hour intervals starting 24 hours before sacrifice. The animals in all three experimental groups were sacrificed at either one, two, three, four, or five days after spinal cord injury.

All of the animals were subjected to the following operative procedure. After administration of ether anesthesia beginning between 9 A.M. and 11 A.M., the dorsal skin was shaved and washed. A midline dorsal incision was made in the region of the lower thoracic vertebral column. A sterile 30 gauge hypodermic needle was carefully inserted between two of the spinous processes of the lower thoracic vertebrae and pushed ventrally with a slight rotary motion through intervening structures and the spinal cord until it was felt to encounter the resistance of the vertebral body. The needle was then quickly removed, and the skin incision was closed with 6–0 silk sutures. After the animals recovered from anesthesia, they were observed for paralysis and at later times for wound infection. No wound infection was ever noticeable, and although two animals in group III showed some weakness of their hind legs, it did not interfere with their eating or drinking.

In group II four animals and in group III five animals, which will be considered as sham operated controls, were thought to be treated exactly as the others, but the spinal cord, as shown by subsequent detailed histological examination, was not injured.

All of the animals were sacrificed by intracardiac perfusion with 10% formalin made from paraformaldehyde according to the method described by Pease ('64). The caudal two-thirds of the vertebral column was removed and washed briefly in water. The spinal cord was dissected out, washed overnight in running water, dehydrated, embedded in paraffin, and sectioned at 7 μ. After removal of paraffin and staining by the periodic-acid Schiff technique, the slides were coated with either Kodak NTB3 nuclear emulsion or Ilford L-4 emulsion and exposed at 4°C for 3 to 12 weeks. Following three to five minute development in Kodak Microdol X the slides were additionally stained with hematoxylin or with hematoxylin and eosin, dehydrated, and mounted in Permount.

The sections chosen for study came from the central part of the spinal cord, and most of them contained segments of central canal. One complete section from each animal was scanned under oil im-

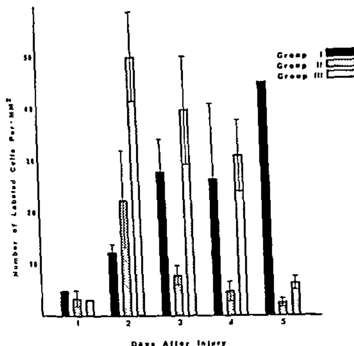


Fig. 1 Mean density of radioactive cells for each group on each of the five days after injury. The bar indicates the standard error of the mean. Controls (not shown) all had less than two labeled cells per mm². There was a progressive increase over the five day period in the number of pre-labled cells (group I). The number of cells undergoing DNA synthesis in the spinal cord (groups II and III) reached a maximum on the second and third days after injury and by the fifth day was again near control levels.

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DISCUSSION

The experiments described in this pape have been directed toward the followin question: Does the DNA synthesis whicl occurs in nervous tissue following a loca injury occur only in mononuclear leuko cytes that have infiltrated the nervous tis sue? The experimental approach to thi question was based on the assumption tha if a population of cells is distributed ove a defined area of tissue in a manner tha is the same in all animals treated the sam way, then two independent and unbiase samples of this population should hav similar distributions. The population c cells in question was the population c mononuclear leukocytes which infiltrat the spinal cord following a penetrating ir

RESULTS

In most of the sections examined the wound was located near the center of the section, although in some of the animals it was more caudally placed. In the animals killed the first day after injury the lesion contained mostly erythrocytes, necrotic debris and polymorphonuclear leukocytes. Cells with dark staining, irregular nuclei and macrophages containing erythrocytic debris were prominent on the second day, and mitotic figures were easily found near the wound on the second and third days after injury. By the fourth and fifth days the central core of the lesion was densely packed with nuclei, and the polymorphonuclear leukocytes had virtually disappeared.

The mean density of radioactive cells (as determined by dividing the total number of radioactive cells per section by the area of the section) is shown for each group on each day of the experiment in figure 1. The density of labeled cells in group I increased over the five day period. The radioactive cell densities in groups II and III reached maximum values on the second day after injury and on the fifth day were only slightly above control levels. The sham operated animals in groups II and III all had less than two radioactive cells per square millimeter of spinal cord.

Examples of the distribution of radioactive cells in the spinal cord sections from group I are shown in figure 2. The large concentration of labeled cells at the lesion, which appears to reach its peak on the third and fourth days after injury, is readily apparent. This distribution of labeled cells should be compared with that found in group II (fig. 3). Here on the second and third days after injury the labeled cells were more diffusely distributed. On the other days more of the labeled cells were located near the lesion, but the difference in radioactive cell density at the lesion from that over the rest of the section is not as high in group II as in group I. In group III the distribution of labeled cells (fig. 4) is similar to that in group II except for an increase in the total number of labeled cells and somewhat greater concentrations at the lesion on days three and four. Again, the labeled cells are seen to be distributed throughout the section on the second day after injury.

The distribution curves which were determined from the maps of all animals in each group are shown in figures 5 through 9. In all cases group I has the highest value at the lesion with a sharp decline as one moves away from the lesion. Group II tends to be much less sharply peaked [at] the lesion, and group III is intermedia[te] between the other two groups in this respect.

To empasize the greater concentration of labeled cells at the lesion in group I as compared with the other two groups, the ratio of the density of labeled cells in the row passing through the lesion to the density of labeled cells over the entire section was calculated for each spinal cord. In group I this ratio ranged from 6.5 to 20.7. In group II the range was from 0.0 to 15.6 and in group III, from 0.8 to 14.6. Figure 10 shows the mean value of this ratio plotted for each group on each of the five days after injury. The values for groups II and III were not significantly different from each other. However, on the second and third days after injury the values for group I were significantly higher than the values for group II. (Days one and five were not included in the statistical analysis, since there was but a single animal in group I on each of these days).

Table 1 shows the values calculated for slope and y-intercept of the straight line for each animal. When these values were ranked and tested by the method of Kruskal and Wallis ('52), it was found that there is a very low probability that either the slopes ($p < 0.05$) or the y-intercepts ($p < 0.01$) from the three groups of straight lines represent samples from the same population. When group I was compared against group II, there were significant differences in both the slope ($p < 0.05$) and the y-intercept ($p < 0.01$). When group I was compared against group III, only the difference between the y-intercepts was significant ($p < 0.01$). In neither the slope nor the y-intercept were there significant differences when groups II and III were compared.

Table 2 shows the results of the nuclear classification of labeled cells that were indistinguishable from glial cells. In all

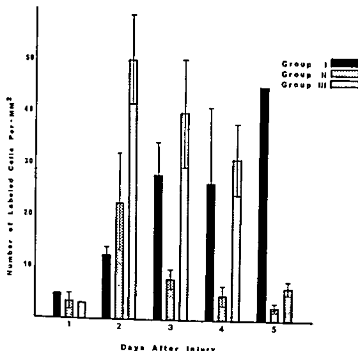


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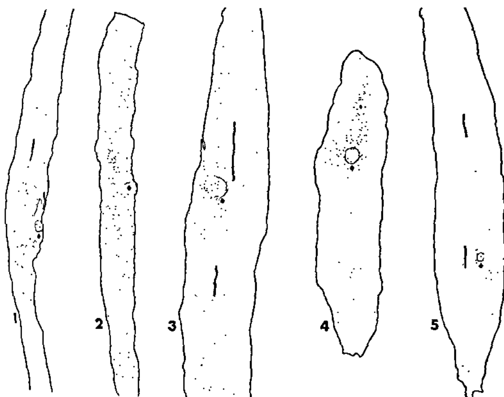


Fig. 3 Representative spinal cord sections from animals in group II. Here the black dots indicate the positions of cells synthesizing DNA at the time of injection, one hour before sacrifice. Numbers indicate the day after injury that the animal was sacrificed.

those which were in the blood, the vast majority of the labeled cells found in group I must be hematogenous. This method of tracing labeled leukocytes into tissues where there are normally only a low number of cells synthesizing DNA has been used previously in the study of wounded muscle (Bintliff and Walker, '60), injured spinal cord (Adrian and Walker, '62; Walker, '63) and injured brain (Konigsmark and Sidman, '63; Huntington and Terry, '66). That these infiltrating mononuclear leukocytes definitely do divide in nervous tissue is demonstrated by the numerous labeled mitotic figures found in group I (figs. 11 through 14; table 4 and 5).

The labeled leukocytes that were found within the nervous tissue were located within the neuropil and were not confined to vessels. The animals in all the experimental groups were sacrificed by perfusion fixation, and the vessels were washed clear of cells. Only occasionally were labeled cells found in relation to vessel walls.

The labeled cells found in the spinal cords from the animals in group II, which received a single injection of tritiated thymidine one hour before sacrifice, represent sites of local DNA synthesis. The much higher number of labeled cells found in the spinal cords of the injured animals over that found in the sham-operated animals should be cells that have synthesized DNA in response to the injury. They cannot be cells that have acquired their label elsewhere before migrating into the nervous tissue, because there was insufficient time between the injection of the labeled thymidine and sacrifice for significant migration of labeled cells to occur.

When the drawings of the spinal cord sections from the animals in group I were

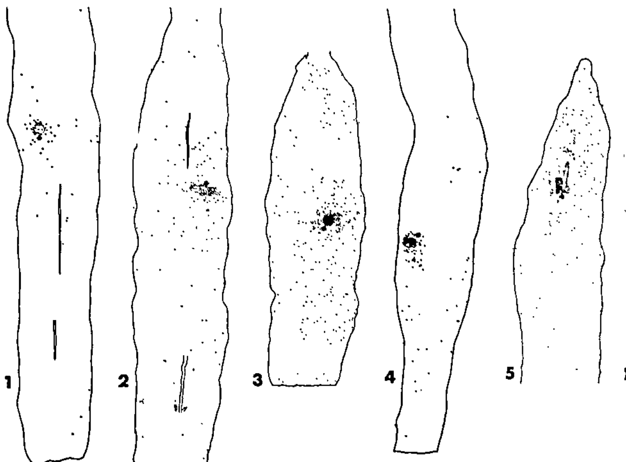


Fig. 2 Outline drawings of spinal cord sections from representative animals in the group injected with tritiated thymidine before injury (group I). Numbers indicate the day after injury when the animal was killed. Black dots show positions of radioactive cells, most of which entered the spinal cord after becoming labeled. Arrows indicate the position of the wounds.

jury. It has been shown that a substantial proportion of this cell population can be labeled by injecting tritiated thymidine several hours prior to injury (Walker, '63). If mononuclear leukocytes are the only cells in the spinal cord engaged in DNA synthesis following injury, then the cells labeled by injecting tritiated thymidine after injury should represent a second, independent sample of the population of mononuclear leukocytes. If the distributions of these two groups of labeled cells are dissimilar, then either the assumption stated above is false, or the hypothesis that only the leukocytes undergo DNA synthesis is false.

Tritiated thymidine is generally considered to label only DNA that is being synthesized in preparation for division (Cronkite et al., '59; Gall and Johnson, '60), and it is incorporated for only a short time following injection (Hughes et al., '58). In normal adult nervous tissue, the number of labeled cells following tritiated thymidine injection is very low (Smart and Leblond, '61; Adrian and Walker, '62). In the spinal cord sections from the animals in group I, which received three injections of tritiated thymidine at 24, 20 and 16 hours before injury, most of the labeled cells at the wound must be due to an influx of labeled mononuclear leukocytes. Since the last injection of labeled thymidine was given 16 hours before the injury, none of it would have been available in the blood to label cells which began DNA synthesis after the injury. Therefore, the population of labeled cells which appeared at the site of injury must have migrated in from outside the nervous tissue. Since there was no large population of labeled cells close to the wound, except

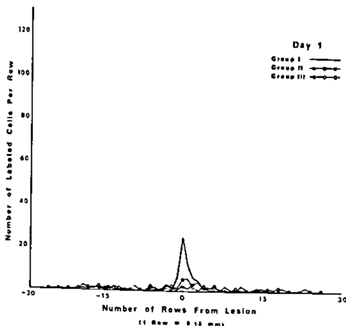


Fig. 5 Comparison of the distribution curves for the three groups of experimental animals sacrificed on the first day after injury. Each point was determined by averaging the individual values for a given row from all the animals in the group. Beyond three rows from the lesion the curves are indistinguishable. In the immediate vicinity of the lesion, the values for group I are much higher than those for the other two groups.

center of the wound. The number of labeled cells in the row passing through the lesion ranged from 25 on the first day to 213 on the third day after injury and averaged 91 for the eleven animals in the group. At a distance of more than four rows (0.7 mm) from the lesion the number of labeled cells per row did not exceed 15 on the first four days after injury and was usually less than ten. In contrast, the number of cells in the row through the lesion in group II ranged from zero to 21 and averaged 11. On the second day after injury the number of cells in DNA synthesis was greater than the number of labeled leukocytes at all places other than in the one or two rows nearest the lesion. On the other days, at a distance from the lesion, the number of cells in DNA synthesis was of comparable magnitude to the number of labeled leukocytes, except on the fifth day when there were more labeled cells in group I than in group II.

Since the exact percentages of the total cell population which the two samples of labeled cells constituted were unknown, it seemed more meaningful to compare the distributions of the two samples about the lesion than to make a statistical comparison of absolute numbers of labeled cells in the two groups. To transform the data from the two samples to comparable forms, the number of labeled cells at a given distance from the lesion in each spinal cord section was expressed as a fraction of the number found in an equivalent area passing through the lesion. By this means the number of labeled cells at the lesion for every graph was set equal to 1.0, and the number of cells per row at a distance from the lesion was expressed as a fractional value of 1.0. When the best fitting straight line was computed for each set of these ratios, it was found that there was a very low probability that the family of lines from group I and the family of lines from group II represented samples from the same population. The distribution of labeled mononuclear leukocytes [illegible] to a spinal cord lesion was signifi-

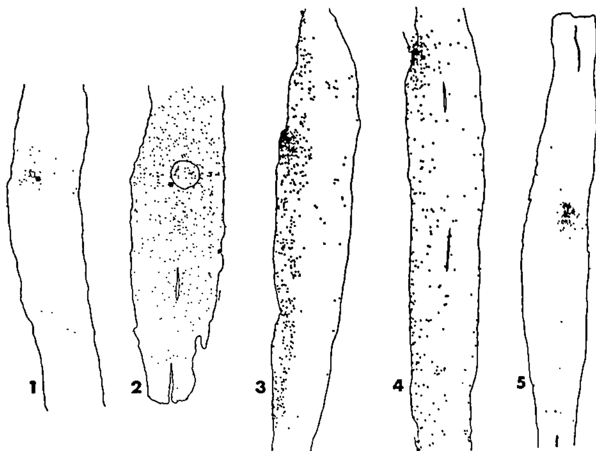


Fig. 4 Spinal cord sections from the animals in group III, which received four injections of tı tiated thymidine during the 24 hour period prior to sacrifice. Although most of the dots represeı cells that became labeled in the nervous tissue, there are probably some cells present that entered tl nervous tissue after acquiring their label.

compared to those from the animals in group II, the patterns of distribution of radioactive cells were seen to be distinctly different. In all the sections from group I there was a large concentration of labeled leukocytes at the lesion compared to the numbers of such cells found at even short distances away. In most of the sections from group II there was an even distribution of cells in DNA synthesis over the entire section. At the wound the number of labeled infiltrating cells was very much greater than the number of cells in DNA synthesis on all five days. On the second day after injury there were more cells in DNA synthesis away from the wound than there were labeled leukocytes from the blood. By the fourth and fifth days after injury, while at the lesion there were still many more labeled leukocytes than there were cells in DNA synthesis, away from the wound the number of leukocytes wa of the same order of magnitude as th number of cells in DNA synthesis.

Although each spinal cord section wa obviously only a two dimensional samp of a three dimensional distribution of la beled cells, the sections were comparab from animal to animal in that the plane o section was always at right angles to th needle track and the sections chosen fo study all came from the central region o the cord. When the two dimensional map were reduced to one dimensional graph showing the number of labeled cells in band or row across the width of the se tion plotted against the longitudin dimension of the cord, the differences a parent on the maps were even more pr nounced. All of the graphs from group had an extremely sharp peak that wa limited to the one or two rows nearest th

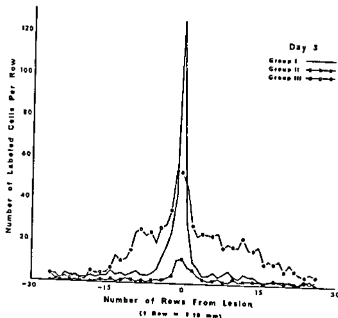


Fig. 7 Comparison of distribution curves on the third day after injury. The curve for group I is an average of the values from four animals. That for group II is an average of the values from five animals, and that for group III is an average from three animals. The curve for group III has become more sharply peaked at the lesion, but its values still remain high at a distance from the lesion. Most of the labeled cells in group I are within six rows of the center of the lesion, and the total number of labeled cells has increased over that found on the second day.

the spinal cord in groups II and III, they were seen to be very different in the area near the lesion. The straight lines plotted for group I were shown to be different from those plotted for either of the other groups. At the lesion there was a much greater difference in the density of leukocytes and the density of cells synthesizing DNA than there was elsewhere on the sections. A possible explanation for this difference in distribution of the leukocyte population and the population of cells synthesizing DNA was that the leukocytes had less tendency to divide near the lesion than elsewhere. To rule out this possibility, mitotic counts were made on several of the sections from group I. Table 4 shows that there is no significant difference in the percentage of leukocytes found in mitosis in the area within 0.7 mm from the lesion and the percentage found in mitosis between 0.7 mm and 3.0 mm from the lesion. Therefore, the leukocytes have just as much ability to divide, and presumably to synthesize DNA, near the lesion as at a considerable distance from it. On the other hand, the leukocytes account for a significantly higher percentage of the total number of mitotic figures in the area within 0.7 mm from the lesion than they do at a greater distance from the lesion (table 5), which would indicate that some other type of cell is also dividing.

On the second day after injury it was only in the immediate vicinity of the lesion that there were a sufficient number of mononuclear leukocytes to account for the observed DNA synthesis in either group II or group III. Beyond a distance of 1.5 mm from the lesion there were roughly six times as many cells in which DNA synthesis was occurring in group II as there were labeled mononuclear leukocytes in group I. Beyond a distance of 0.7 mm from the lesion there were almost ten times as many labeled cells in group III as there were labeled mononuclear leukocytes in group I. If the mononuclear leukocytes

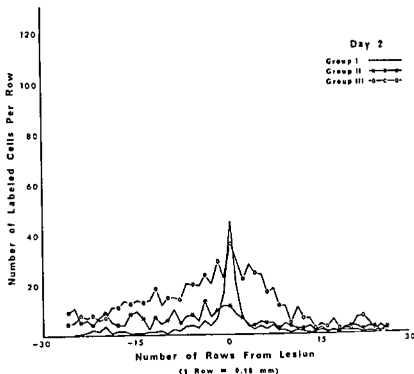


Fig. 6 Comparison of the distribution curves on the second day after injury. Each curve is an average of the values from three animals. Most of the labeled cells in group I are within four rows of the center of the lesion. Groups II and III are more evenly distributed; however, the curve for group III does reach its peak at the lesion.

cantly different from the distribution of cells synthesizing DNA in response to the injury.

The cells labeled in group III should include all of the cells that began DNA synthesis during the 24 hour period starting 30–32 hours before sacrifice, provided that the period of DNA synthesis in these cells is of the same duration (six to eight hours) as that found in most other mammalian cell types (Cameron and Greulich, '63). In the spinal cord this population is principally composed of cells that were in the spinal cord at the time DNA synthesis began. In addition, some mononuclear leukocytes which acquired their label elsewhere are probably among the labeled cells in group III, since a large percentage of these cells are labeled in the blood within 12 hours after beginning tritiated thymidine injection (Bintliff and Walker, '60). The relative magnitude of this infiltration of labeled cells in group III may be estimated by comparing the distribution curves for groups II and III on each of the five days. The cells labeled in group II are known to be cells that were synthe[sizing] DNA in the spinal cord shortly be[fore] sacrifice. Since the period of DN[A] synthesis is roughly six to eight hour[s] there should be about three or four time[s] as many of these cells labeled in group II[I] as are found labeled in group II. Th[e] number of labeled cells in group III i[n] excess of three or four times the numbe[r] in group II could represent either an in[crease] due to division of some of the earl[i]est cells to be labeled or an infiltration o[f] labeled leukocytes. Only on days three an[d] four were there more than four times a[s] many labeled cells in group III as were i[n] group II. There was a significant diffe[r]ence between the y-intercepts of th[e] straight lines computed for groups I an[d] III, which would indicate that the compo[nent] of labeled infiltrated cells in grou[p] III was probably quite small relative t[o] the component of cells that had undergon[e] DNA synthesis in the nervous tissue.

When the distribution of labeled leuko[cytes] in group I was compared to the di[s]tribution of the cells synthesizing DNA i[n]

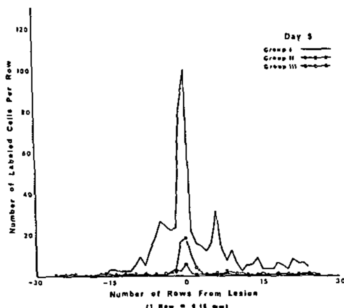


Fig. 9 Comparison of the distribution curves on the fifth day after injury. The curve for group I is based on a single animal. The curves for groups II and III are each based on four animals. The total number of labeled cells has increased in group I. Although most of them are still found close to the lesion, many more are found throughout the section than were found at the earlier times. The number of labeled cells in groups II and III has become quite small, and most of these are near the lesion.

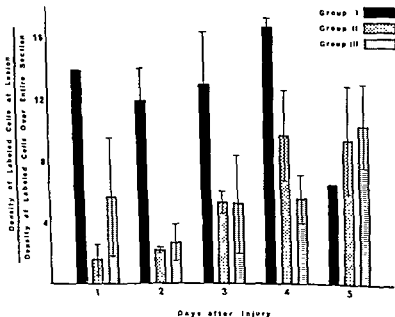


Fig. 10 Relative density of labeled cells at the lesion. The ratio of the density of labeled cells in the row passing through the lesion to the density of labeled cells throughout the entire spinal cord section is shown for each group on each day. Brackets indicate the standard error of the mean. The values for groups II and III are not significantly different from each other. On the second and third days after injury the values for group I are significantly higher than the values for group II ($p < 0.05$).

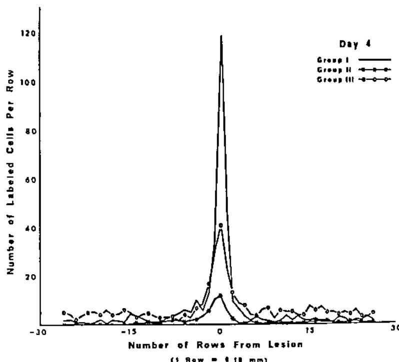


Fig. 8 Comparison of distribution curves on the fourth day after injury. The curve for group I is based on two animals. The curves for groups II and III are each based on four animals. There is little change in the distribution and in the number of labeled cells in group I from that found on the third day. The total number of labeled cells in group III is much smaller than on the previous two days, and the curve has become much more sharply peaked at the lesion.

were the only cells in the spinal cord undergoing DNA synthesis on the second day after injury, no more than 10% of them could have been labeled in group I, and all of them would have had to divide at least once during the second day after injury. However, from the work of others it is clear that a much higher percentage of the total population of mononuclear leukocytes must be labeled in group I by the second day after injury (Bintliff and Walker, '60; Konigsmark and Sidman, '63; Huntington and Terry, '66). Therefore, some type or types of cells other than the mononuclear leukocytes present in such large numbers at the lesion must also have been synthesizing DNA in the spinal cord on the second day after injury.

The exact nature of this cell has not been determined by this study. The possibility that some of the DNA synthesis may occur in small lymphocytes, in which the percentage labeled in the peripheral blood is known to be low, has not been completely excluded. Walker ('63) has shown that these cells probably enter injured spinal cord but that they do not collect at the wound area in large numbers. Mononuclear leukocytes normally present in the nervous tissue, as opposed to those that infiltrated because of injury, may also be sites of DNA synthesis. Roessmann and Friede ('67) found labeled leukocytes in normal nervous tissue of recipient animals following transfusion of blood from animals that had received tritiated thymidine previously, indicating that some of these cells are in nervous tissue at all times. The number of such labeled cells was increased following injury. That endothelial cells must account for a portion of the nonleukocytic DNA synthesis is clear from the fact that a small number of endothelial cells were seen to be labeled in groups II and III throughout most of the five day period. However, the labeled endothelial cells were found almost entirely in the area of the lesion. Endothelial proliferation following nervous tissue injury has been previously reported by Bender ('25),

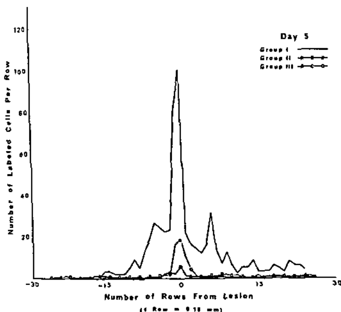


Fig. 9 Comparison of the distribution curves on the fifth day after injury. The curve for group I is based on a single animal. The curves for groups II and III are each based on four animals. The total number of labeled cells has increased in group I. Although most of them are still found close to the lesion, many more are found throughout the section than were found at the earlier times. The number of labeled cells in groups II and III has become quite small, and most of these are near the lesion.

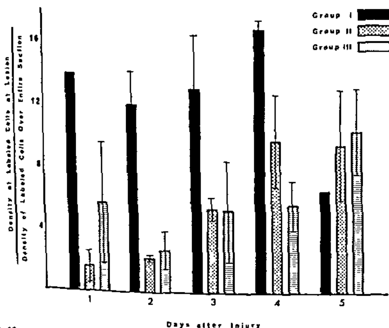


Fig. 10 Relative density of labeled cells at the lesion. The ratio of the density of labeled ells in the row passing through the lesion to the density of labeled cells throughout the ntire spinal cord section is shown for each group on each day. Brackets indicate the andard error of the mean. The values for groups II and III are not significantly different om each other. On the second and third days after injury the values for group I are significantly higher than the values for group II ($p < 0.05$).

TABLE 1

Values for slope and y-intercept of the straight lines calculated for each animal

Day after injury	Group I Slope	Group I Y-intercept	Group II Slope	Group II Y-intercept	Group III Slope	Group III Y-intercept
1	−0.07	0.32	0.20	1.40	−0.07	0.35
			−0.06	1.62	0.12	0.05
2	−0.02	0.21	0.02	0.76	−0.02	0.62
	−0.09	0.43	−0.01	0.99	−0.06	1.10
	−0.02	0.17	−0.13	0.92	−0.03	1.27
3	−0.03	0.15	−0.33	1.46	−0.18	1.03
	−0.08	0.45	−0.08	0.53	−0.10	0.47
	−0.14	0.69	−0.18	0.96	−0.11	0.85
	−0.05	0.25	−0.12	0.53		
			−0.15	0.81		
4	−0.06	0.29	−0.18	1.09	−0.20	1.18
	−0.04	0.24	−0.12	0.51	−0.11	0.54
			−0.11	0.50	−0.16	0.78
			−0.14	0.86	−0.14	0.77
5	−0.01	0.25	−0.15	0.75	−0.08	0.36
			−0.07	0.44	−0.08	0.35
					−0.07	0.34
					−0.06	0.29
Mean	−0.06	0.31	−0.10	0.88	−0.08	0.65

P y-intercepts < 0.01, P slopes < 0.05.

TABLE 2

Classification of radioactive nuclei indistinguishable from glial nuclei in 1.25 mm² around lesion

Group	No. of animals	Total area examined	Total [1] no. of nuclei	Total R.A.	Types of radioactive nuclei: Round, oval or irregular — Dark	Round, oval or irregular — Light	Elongated
		mm^2			%	%	%
I	11	13.75	14,639	2,119	82.9 ± 9.5	9.5 ± 9.5	7.4 ± 5.9
II	17	21.25	22,573	500	61.8 ± 15.3	25.3 ± 10.5	12.9 ± 11.2
III	16	20.00	16,948	1,511	79.4 ± 10.9	11.0 ± 10.2	9.6 ± 7.6

[1] Excluding neurons, ependymal cells, polymorphonuclear leukocytes, and endothelial cells.

TABLE 3

Radioactive endothelial cells in 1.25 mm² around lesion

Group	Total no. of endothelial nuclei	Radioactive endothelial nuclei no.	Radioactive endothelial nuclei %
I	1,736	22	1.26
II	4,064	61	1.50
III	3,457	57	1.65

Baggenstoss et al. ('43) Klatzo et al. ('58), Levine and Stypulkowski ('59), and Cammermeyer ('65). Since the leukocytes in group I were distinguishable from the glial nuclei among which they were found only by their label, the nuclear morphology of the labeled cells in groups II and III is of little value in distinguishing leukocytes from glial cells. A slightly higher percentage of the labeled cells in groups II and III fell into the categories of elongated and light staining nuclei than in group I, and it is possible that connective tissue cells and astrocytes may be responsible for some of the DNA synthesis that occurred. Several authors have maintained that a large portion of the population of macrophages that invade brain wounds are derived from vascular adventitial cells, and proliferation of these cells was usu-

TABLE 4

Labeled mitotic figures shown relative to the total number of labeled cells in group I

	Within 0.7 mm from lesion	Between 0.7 mm and 3.0 mm from lesion	Total
Labeled mitoses	19	6	25
Other labeled cells	914	312	1226
Total labeled cells	933	318	1251

$\chi^2 = 0.005, p > 0.95$.

TABLE 5

Labeled mitotic figures shown relative to the total number of mitotic figures in group I

	Within 0.7 mm from lesion	Between 0.7 mm and 3.0 mm from lesion	Total
Labeled mitoses	19	6	25
Unlabeled mitoses	5	13	18
Total mitoses	24	19	43

$\chi^2 = 8.01, p < 0.01$.

ally thought to be a necessary preliminary event (Cone, '28; Carmichael et al., '39; Baggenstoss et al., '43; Silver and Walker, '47). Following tritiated thymidine injection, Konigsmark and Sidman ('64) found a few labeled nuclei in injured brains that stained with gold chloride sublimate and which they identified as astrocytes. The other reports of labeled glial cells following tritiated thymidine injection into animals with nervous tissue injuries have assumed either that no leukocytes were present to be labeled or that the glial cells could be distinguished from leukocytes on the basis of nuclear morphology (Koenig, '60; Koenig et al., '62; Altman, '62; Adrian and Walker, '62; Samorajski et al., '64; Hassler, '66).

On the basis of the results thus far presented it is concluded that although mononuclear leukocytes do freely invade the nervous tissue and synthesize DNA in and around sites of injury, they are not the only cells in which such DNA synthesis occurs. A substantial number of cells already present in the nervous tissue must engage in DNA synthesis during the five day period following injury. This synthesis begins near the end of the first day after injury and is occurring in a large number of cells by the end of the second day. Many of these cells are located at relatively long distances from the lesion on that day. The number of such cells is considerably diminished by the fifth day after injury.

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11
12
13
14

PLATE 1

EXPLANATION OF FIGURES

Examples of mitotic figures found in the spinal cord sections from the animals in group I, which were injected with tritiated thymidine before injury and sacrificed two (fig. 14) or three (figs. 11–13) days after injury.

11 Two prophases that were located near the lesion. Hematoxylin and eosin. × 1050.

12 A prophase and a metaphase found near the lesion. Both mitotic figures were weakly labeled. Hematoxylin and eosin. × 1550.

13 A labeled prophase and an unlabeled metaphase (arrow) found near the lesion. Hematoxylin and eosin. × 1550.

14 Unlabeled anaphase (arrow) found at a considerable distance from the lesion on the second day after injury. Three neuronal nuclei and a capillary distended by the perfusion are also visible. Periodic acid Schiff hematoxylin. × 1050.

Light and Electron Microscopic Observations on the Spleen and the Splenic Leukocytes of the Newt *Triturus cristatus*

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ABSTRACT The newt spleen has an outer region, the red pulp, that consists of reticular fibres, reticular cells, and fixed macrophages forming a framework within which erythropoiesis occurs, and an inner region, the white pulp, consisting of a reticular framework containing predominantly lymphocytes. The fine structure of the following leucocytes, namely lymphoid cells, reticular cells, macrophage, haemocytoblasts, plasma cells and neutrophilic, basophilic and eosinophilic granulocytes is, in general, similar to that of the corresponding mammalian blood cells. However, in contrast, all the nuclei contain large numbers of interchromatin granules and a differentiated region termed the nuclear light-staining zone because of its electron-scattering properties. Furthermore the specific granules of the eosinophilic granulocytes are structureless whereas those of the basophilic granulocyte have crystalline regions. The structure and distribution of the granular, fibrillar, tubular and vesicular components of the cytoplasm of each cell type is described.

We have previously described the fine structure of the chromosomes in mitotic cells in the spleen of the newt *Triturus cristatus* (Davies and Tooze, '64, '66) and the cellular changes occurring during erythrocyte maturation in this organ (Tooze and Davies, '67). In this paper we describe electron microscopic observations of the fine structure of the non-erythrocytic cells in the newt spleen. We also examined these cells by light microscopy and as far as possible attempted to correlate the electron microscopic observations with our own and the earlier light microscopy of Jordan and Spiedel (see review Jordan, '38).

The early comparative haematologists (see especially Jordan, '32, '38; Jordan and Speidel, '24, '30; and reviews by Foxon, '64; Andrew, '65) established the importance of the spleen as the primary haematopoietic organ in lower vertebrates. In adult urodeles, the spleen is the exclusive site of erythropoiesis except in a few species, where most of the stem cells, the so-called haemocytoblasts, leave the spleen and differentiate in the circulating blood (Jordan, '32). Development of thrombocytes also occurs in the spleen (Jordan and Speidel, '30), whereas granulocytes usually develop in the subcapsular region of the liver, entering the spleen presumably via the circulation (Jordan and Speidel, '24).

Jordan ('32) commented on the striking similarity between the types of blood cells occurring in amphibians and mammals. Our observations on the fine structure of newt blood cells can be compared with those from many studies on mammalian, and from a few studies on avian, haematopoietic organs and blood cells, for example, Braunsteiner and Pakesch ('55); Low and Freeman ('58); Policard, Collet and Pregermain ('59); Bessis ('61); Bernhard and Granboulan ('60); Granboulan ('60); Sorenson ('60a,b); and Han ('61).

MATERIALS AND METHODS

The observations were made on spleens from the newt *Triturus cristatus*. The methods of preparation and study have been described in detail elsewhere (Davies and Tooze, '66; Tooze and Davies, '67). Briefly, smears were examined after staining in May-Grünwald-Giemsa (M.G.G.). Whole spleens or pieces were fixed in glutaraldehyde, post-fixed in OsO_4 (Sabatini, Bensch, and Barrnett, '63) and embedded in araldite (Glauert and Glauert, '58; Luft, '61; Grimley, Albrecht and Michelitch, '65). Sections were stained with both uranyl

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cur. Reticular cells are not seen in smears and presumably remain behind in the reticular framework when the spleen is smeared as do most of the macrophages. Plasma cells have not been identified in smears. The three types of granulocytes are easily recognized. Neutrophilic granulocytes (fig. 9) have a highly polymorphic nucleus; the cytoplasm is unstained. Eosinophilic granulocytes (fig. 7) also have a highly lobed nucleus, and an extensive cytoplasm filled with orange staining eosinophilic granules. Basophilic granulocytes (fig. 8) can sometimes be seen to have a bilobed nucleus easily distinguishable from that of the eosinophil, and a cytoplasm filled with granules staining a deep bluish red.

Electron microscopy

1. *Lymphoid cells*

In the electron microscope a series of lymphoid cells with varying structure are seen. Typically small or medium sized lymphocytes appear round or slightly oval in cross section (fig. 14). Their nuclei, usually with several indentations, contain one or more large blocks of chromatin and many small regions of chromatin ranging in size from about 2500A downwards, that is at about the limits of resolution of the light microscope. Chromatin blocks line the interior of the nuclear envelope. Also inside the nucleus are small lightly-stained regions partly bordered by chromatin, the structure of which is not well defined (fig. 14). Similar regions were found in the nuclei of erythroblasts (Tooze and Davies, '67) and were named nuclear light-staining zones. The nuclear sap contains numerous granules, referred to as interchromatin granules (Bernhard and Granboulan, '63), ranging in diameter from about 400A downwards. These interchromatin granules are common in all the cell types described below, as well as in erythroblasts from *Triturus cristatus* (Davies and Tooze, '66; Tooze and Davies, '67). Occasionally groups of very large granules, about 1200A in diameter, sometimes with hollow centres or U-shapes (figs. 14, 15), lie within the nuclear sap. Small nucleoli are common.

The cytoplasm forms a narrow rim about the nucleus and the surface is often thrown into finger-like projections. Ribosomes are not very numerous; a few occur attached to endoplasmic reticulum, whilst smooth-surfaced endoplasmic reticulum in the form of flattened sheets and vesicles, is fairly common. There are a few mitochondria per section and usually granules, smaller than mitochondria, packed with homogeneous dark staining material (fig. 14).

The size and shape of the nucleus, the numbers of free ribosomes per unit area of cytoplasm and the amount of cytoplasm all vary in these lymphoid cells. Some of the lymphocyte-like cells that have elongated nuclei may correspond to the thrombocytes seen in the light microscope.

We have seen other cells which have a pronouncedly kidney-shaped nucleus. Centrioles as well as Golgi material almost invariably occur within the nuclear indentation forming a cytocentrum. Mitochondria and somewhat smaller electron-dense granules are also present within the cytocentrum. These cells may correspond to the monocytes seen in smears.

Another cell (fig. 16) with easily recognisable characteristics is found infrequently and we believe it corresponds to the haemocytoblast seen in smears. This cell is characterised by a more abundant cytoplasm than is typical of the small lymphocytes; the cytoplasm is packed with ribosomal aggregates (polysomes) and many mitochondria. There is little granular or smooth-surfaced endoplasmic reticulum, but there is a Golgi zone, centrioles with pericentriolar bodies (Bernard and de Harven, '58; Bessis, '61; and Gall, '61) and a few microtubules about 230A in diameter (see fig. 17). A few membrane-limited densely staining granules, smaller than mitochondria, also occur in the cytoplasm. The plasma membrane is often thrown into numerous projections. Much of the nuclear chromatin is distributed throughout the nucleus in small (≤ 2500A) blocks, and there is a narrow rim of condensed chromatin on the nuclear envelope (fig. 16). Nucleoli occur in these cells, as well as nuclear light-staining zones. In some cells, which have the cytoplasmic characteristics of haemocytoblasts, the nuclear chromatin does not occur in small (≤ 2500A diameter) blocks

acetate and lead citrate (Reynolds, '63) for electron microscopy or with various stains for light microscopy. The sections were examined in either a Zeiss E.M.9 or a Siemens Elmiskop I.

The problems of recognising the different types of blood cells in the electron microscope are well known (see e.g., Bernhard and Granboulan, '60) and we have discussed them previously (Tooze and Davies, '67).

RESULTS

Light microscopy

1. *The structure of the spleen*

The spleen of *Triturus cristatus* (figs. 1, 10) has a capsule consisting of an outer mesothelial cellular layer and an inner connective tissue sheath. The connective tissue extends inwards as trabeculae which join a reticular network (figs. 10, 11, 23, 24), so producing the framework of the organ. This reticular framework consists of groups of reticular fibres (figs. 11, 24) surrounded but often not enclosed by the cytoplasm of reticular cells, together with fixed macrophages (fig. 11). The extracellular reticular fibres each have a diameter of about 3000A and an ill-defined banding (fig. 12). There is hardly any amorphous material surrounding these fibres, but there are fine threads lying between them (figs. 12, 13).

The distribution of red and white cells (fig. 1) is similar to that in the spleen of the newt *Triturus* (formerly *Diemyctylus*) *viridescens* described by Jordan and Speidel ('24). The outer region of the organ, the red pulp, consists partly of well defined venous sinuses, that is groups of red blood cells at various stages of development together with cells from circulating blood, such as lymphocytes and granulocytes, lying in irregularly shaped cavities outlined by the reticular framework. The venous sinuses (fig. 10) are interspersed among other areas of indefinitely arranged reticular fibres, reticular cells, macrophages and red and white blood cells, areas that may correspond to the cords thought to exist in the human spleen (Bloom and Fawcett, '62). In the outer regions of the red pulp the red blood cells are generally closely packed together (fig. 1). Well defined blood vessels, circular or elliptical in section, that contain loosely packed erythrocytes are also present. The inner region of the spleen, the white pulp, consists of loosely arranged reticular fibres and reticular cells enclosing masses of lymphocytes and a few erythrocytes (see fig. 1).

As we reported elsewhere (Tooze and Davies, '67) the size of the spleen and the relative numbers of red and white cells varies greatly from animal to animal.

2. *May-Grünwald-Giemsa smears*

The various types of blood cells in the spleen were identified in M.G.G. stained smears. Lymphocytes of various sizes (figs. 2, 4) are characterized by a reddish, sometimes slightly reddish-blue (purple) nucleus usually containing well defined blocks of chromatin and a narrow band of cytoplasm, either hardly discernible or staining a light blue. In another type of cell (fig. 2), comparable in size with the largest lymphocytes, the nucleus is a definite purple and the cytoplasm is a distinct blue. Usually the nucleus appears more homogeneous in structure than that of the lymphocyte (compare figs. 2, 4) but blocks of chromatin are occasionally discernible (fig. 3). This cell appears to be similar to the haemocytoblast described by Jordan and Speidel (30); haemocytoblasts occur infrequently. There are also cells intermediate in appearance between haemocytoblasts and the lymphocytes.

Cells identified as thrombocytes from criteria described by Jordan and Speidel ('30) and Andrew ('65) occur in clumps (fig. 5). Their nuclei, usually oval in shape, stain deep purple and occasionally appear grooved; the cytoplasm has numerous small projections and is very weakly stained: when the cells are in clumps their cytoplasm appears fused. Under oil-immersion some thrombocytes are seen to contain faint pink granules usually situated at one or both poles of the nucleus.

Monocytes (fig. 6) are readily distinguishable. They have an eccentrically placed nucleus, with a characteristic kidney-shape, staining a reddish colour, and a very weakly basophilic cytoplasm. Macrophages are seen very occasionally in smears, and are identified by the variety of their inclusion bodies, often green in

many groups of flattened sacs and circular vesicles.

4. *Plasma cells*

In the spleens we studied plasma cells were not common. They are characterised by a cytoplasm with enlarged cisternae of granular endoplasmic reticulum containing lightly-staining material (fig. 29). The ribosomes are usually more frequent on the membrane surfaces facing mitochondria (fig. 31) and this, no doubt, has physiological significance. There is extensive Golgi material and centrioles occur with pericentriolar bodies (fig. 29) as well as microtubules each about 230 Å diameter (fig. 30). Membrane-bound granules containing electron dense material are seen near the Golgi zone and scattered throughout the cytoplasm (fig. 29).

In the cells we examined the nucleus was eccentrically placed, contained clumped chromatin with a relatively large area of nuclear sap, and a small nucleolus. Interchromatin granules are prominent.

5. *Granulocytes*

(a) *Neutrophilic granulocytes.* The neutrophilic granulocytes are characterised by a highly polymorphic nucleus (fig. 9) causing the cell to appear multinucleate in thin sections (fig. 32). There are extensive regions of condensed chromatin on the interior surface of the nuclear envelope as well as small ($\leq$ 2500A) blocks of chromatin throughout the nuclear sap. Small nucleoli and interchromatin granules are present.

Scattered throughout the cytoplasm are the characteristic neutrophilic granules. Like those in mammalian neutrophils (Palade, '55; Low and Freeman, '58; and Bessis, '61) they are shaped like rice grains or dumbbells and range in diameter from about 800–2000A. They are membrane limited (fig. 37) and contain almost homogeneous material. There are also structures, interpreted as vesicles or sacs flattened or collapsed for most of their length, (figs. 32, 33, 35, 36), in which the two inner parts of the unit-membranes have fused to produce a darker line (fig. 35). These sacs have about the same circumference as the membranes limiting the neutrophilic granules. In some cells (fig. 33) empty sacs, which are not collapsed, occur suggesting an intermediate stage between fully collapsed sacs and the neutrophilic granules.

According to Palade ('55) there is less endoplasmic reticulum in the neutrophilic granulocytes than in the basophilic and eosinophilic granulocytes of the rat. In those neutrophilic granulocytes of the newt, that we observed, however, there is more smooth and granular endoplasmic reticulum than in the other granulocytes. Occasionally (fig. 33) extensive regions of oriented granular endoplasmic reticulum occur. Scattered free throughout the cytoplasm are densely stained particles about the size of the ribosomes attached to the endoplasmic reticulum. Since after treatment with periodic acid and lead citrate some of these free particles had a granular substructure, resembling that of the glycogen granules in reticular cells, at least some of them may be small glycogen granules rather than ribosomes.

Like mammalian granulocytes (Policard and Bessis, '55) these cells have a large Golgi zone, with centrioles and microtubules about 230A diameter (fig. 34) lying in this region. Bundles of fibrillar material, each fibril about 70A diameter, are scattered throughout the cytoplasm (fig. 38).

(b) *Eosinophilic and Basophilic granulocytes.* The granules of eosinophilic granulocytes are generally understood to contain crystalline cores, whereas those of basophilic granulocytes are thought to be homogeneous (Low and Freeman, '58; Miller, de Harven and Palade, '66). Exceptionally, the granules in the basophilic granulocytes of guinea pig have been shown to contain lamellar structures (Pease, '55; Osaka, '59; Winquist, '60, '63; Fedorko and Hirsch, '65); similar crystalloid structure was found in human mast cells (Fedorko and Hirsch, '65). In *Triturus cristatus*, apart from the neutrophilic granulocytes, we see in the electron microscope two types of granulocytes. In one type (fig. 39) the cytoplasm contains dense homogeneous granules which are about 2–3 μ in diameter, larger than mitochondria, and approximately ovoid in shape. In cross section the cell (fig. 39) appears multinucleate presumably because the nucleus is lobed. These cells, as seen in the

but is in larger aggregates: these are however smaller than the large chromatin blocks that occur in lymphocyte nuclei.

2. *Reticular cells*

The characteristic features of the reticular cells are their association with the extra-cellular fibres of the reticular network and their highly indented nuclei, giving the surface of the nucleus a corrugated appearance (fig. 18). The chromatin is distributed partly in a thin layer on the inner surface of the nuclear envelope, but is mainly scattered in small clumps throughout the interior of the nucleus with occasionally a few larger blocks. There are small nuclear light-staining zones, large nucleoli and interchromatin granules.

The cytoplasm usually forms a fairly thin layer about the nucleus, and characteristically projects over long distances (fig. 23) bounding but not completely enclosing reticular fibres (figs. 11, 24). A striking feature of the cytoplasm is the large amount of fibrillar material it contains (fig. 18, 19), which is often aggregated into bundles and sometimes forms a continuous layer about the cell nucleus just within the plasma membrane. The individual fibrils are about 70A thick (figs. 19, 20). There is a well-developed Golgi region with paired centrioles and pericentriolar bodies (fig. 18). Many profiles of granular endoplasmic reticulum are scattered throughout the cytoplasm, together with free ribosomes, mitochondria and few membrane-bounded granules, smaller than mitochondria, packed with densely-staining material.

The cytoplasm of the reticular cell also contains large amounts of glycogen which we identified with the periodic acid/lead citrate technique of Perry and Waddington ('66). In material fixed in glutaraldehyde and osmium tetroxide and stained with uranyl acetate and lead citrate, the glycogen in these cells normally appears as densely stained homogeneous granules about 250A in diameter (see fig. 19) but very occasionally they show a granular substructure (see fig. 21). After periodic acid treatment and staining in lead citrate however all the granules have this substructure (fig. 22).

3. *Macrophages*

These large cells are readily identified in the electron microscope by the presence in the cytoplasm of dense granules and phagocytosed material (fig. 24). The nucleus appears elongate, and sometimes has deep identations so that in thin sections it may appear bilobed. A layer of condensed chromatin is associated with the nuclear envelope; the interior of the nucleus is filled with many small regions of condensed chromatin and a few larger blocks. Light-staining zones are often seen, the one shown in figure 25 being rather larger than usual. Small nucleoli and interchromatin granules are present. Dense granules (figs. 25, 27) about 500A diameter, which lie embedded in, or closely adjacent to, chromatin, but separated from it by a less dense annulus, are presumably perichromatin granules (Swift, '62; Watson, '62a,b).

The cytoplasmic inclusion bodies show a wide range of size and structure; some are recognisably phagocytosed erythrocytes which themselves contain cytolysomes or polar bodies (fig. 24), while other bodies contain extensive myelin figures (fig. 25). There are at least four classes of roughly ovoid bodies or granules distinguishable within the cytoplasm after staining with uranyl acetate and lead citrate; large granules (g1) each of which contains variable concentrations of dense particles, presumably ferritin, the regions free from ferritin appearing dark-grey in stained sections (fig. 24; see the region of the granule g3 surrounding the eccentric dense granule, fig. 26); ovoid granules of varying size (g2) which appear electron-opaque (fig. 24) and sometimes membrane limited (fig. 24, 28); granules (g3) like g1 granules but containing within them very dense structures similar to g2 granules (fig. 26); smaller dark-grey granules (g4) without ferritin and sometimes seen to be membrane limited (figs. 25, 28). In some cells small electron dense particles, presumably ferritin, are scattered throughout the cytoplasm and nucleus, being excluded however from mitochondria (fig. 26) and chromatin.

The cytoplasm contains extensive profiles of smooth and granular endoplasmic reticulum and a few free ribosomes. The Golgi zone is often extensive consisting of

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electron microscope, closely resemble the eosinophilic granulocytes seen in M.G.G. stained smears (compare figs. 39, 7). For this reason we conclude that they are eosinophilic granulocytes even though the fine structure of their granules differs from the generally understood structure of eosinophilic granules (Low and Freeman, '58; Miller et al., '66).

Most of the chromatin in these eosinophils is condensed on the inner surface of the nuclear envelope, but sometimes (fig. 39) both large and small (2500A) blocks of chromatin occur in the nuclear sap. Nucleoli have not been seen but interchromatin granules are numerous (fig. 39).

The eosinophils have little endoplasmic reticulum but very small vesicles occur throughout the cytoplasm (these are not easily seen in low power micrographs such as fig. 39). Aggregates of free ribosomes are scattered throughout the cell and there are a few mitochondria. Extensive Golgi zones, centrioles and microtubules have been seen; frequently there are finger-like projections from the cytoplasm.

The third type of granulocytes occurs less frequently than the neutrophilic and eosinophilic granulocytes; the shape of the nucleus in thin sections (fig. 40) is easily distinguished from that of the eosinophils and is what would be expected from sectioning the bilobed nucleus of the basophilic granulocytes seen in smears (fig. 8); therefore we identify these cells as basophilic granulocytes. The large dense granules characteristic of the cytoplasm contain crystalline regions (fig. 41) seen as spaced lines (dark lines of variable intensity separated by lighter stained spacings about 60–90A wide). We did not attempt to elucidate their structure further.

DISCUSSION

The fine structural features of the cells termed haemocytoblasts (fig. 16), namely the lack of a well developed endoplasmic reticulum, numerous free ribosomes, moderate numbers of mitochondria and the absence of any specialised cell structures are characteristic of undifferentiated cells (Porter, '54). The structure of these so-called haemocytoblasts in *Triturus cristatus* is similar to that of the primitive cells in popliteal nodes of rabbits described by Sorenson ('60b), to cells in erythropoietic tissues of certain mammals (Sorenson, '60a, '63; Grasso, Swift and Ackerman, '62), and to the lymphoblasts in mammalian lymph nodes (Bernhard and Granboulan, '60). These cells also probably correspond to the haemocytoblasts or lymphoid haemoblasts described by Jordan and Speidel in light microscopic studies of newt spleen (see review Jordan, '38). These variously named cells are similar in appearance, and are thought to give rise to the different types of blood cell. According to the Unitarian theory of haematopoiesis which Jordan held, a single stem cell the haemocytoblast is multipotent, its differentiation into different types of blood cells depending on environmental factors; lymphocytes, although morphologically distinct from haemocytoblasts, also have multiple developmental potency according to this theory. Thus Jordan concluded from the presence of intermediate stages that in the urodele spleen haemocytoblasts or lymphocytes give rise to erythrocytes, thrombocytes and monocytes. Although we have seen a wide variety of lymphoid cells, some intermediate in appearance between the easily recognised lymphocyte (fig. 14) and the so-called haemocytoblast (fig. 16) our data are insufficient to support the theories of Jordan and Speidel.

The fine structure of the white blood cells found in the spleen of *Triturus cristatus*, in general, resembles that of their counterparts in mammalian tissues; this is seen when mammalian plasma cells (Braunsteiner, Fellinger and Pakesch, '53; Braunsteiner and Pakesch, '55; Thiery, '60; Bernhard and Granboulan, '60), small lymphocytes (Low and Freeman, '58; Bernhard and Granboulan, '60), and macrophages (Policard, Bessis and Breton-Gorius, '57; Sorenson, '60b; Han, '61; Thiery, '62) are compared with the same cell types of the newt. Interesting exceptions are eosinophilic and basophilic granulocytes.

In mammalian bone marrow (Policard and Bessis, '58; Bessis, '58, '59) reticular cells and macrophages occur almost exclusively surrounded by erythroblasts forming the so-called erythroblastic islands, and according to some authors (Bessis, '61; Sorenson, '62) there appears to be a direct transfer of material from the macrophage

to the surrounding cells. In newt spleen there are no recognisable cell islands, but basophilic erythroblasts usually occur close to the macrophage and reticular cells of the reticular framework (Tooze and Davies, '67); however, as we have discussed elsewhere (Tooze and Davies, '67), no evidence was seen of a transfer of material such as ferritin from the reticular cells or fixed macrophages to erythroblasts.

A variable amount of fine fibrillar material occurs in the cytoplasm of several types of mammalian blood cells including macrophages, monocytes, neutrophilic and eosinophilic granulocytes, and various blood cells in leukemic animals (de Petris, Karlsbad and Pernis, '62); this material also occurs in basophilic erythroblasts of the newt (Tooze and Davies, '67). The reported diameters of these fibrils vary from about 40–80A in these different cell types and usually they are aggregated into bundles. We have not seen such fibrils in macrophages of newt, but they are occasionally present in neutrophilic granulocytes and are a striking feature of the cytoplasm of reticular cells.

We have observed membrane-bounded granules containing densely staining material in all the cell types recognised in the electron microscope. In lymphocytes, haemocytoblasts, reticular cells and plasma cells these granules are usally smaller than mitochondria and their contents appear homogeneous. These structures may be storage lysosomes similar to those thought to occur in immature erythroblasts (Tooze and Davies, '67) but evidence that they contain lysosomal enzymes is required to confirm this interpretation. We have been able to classify the cytoplasmic granules of macrophage into four distinct types on the basis of their fine structure. Since biochemical studies have shown that macrophages contain lysosomes (Cohn, Hirsch and Wiener, '63) it is likely that some if not all of the types of granules we identify are lysosomes, but whether differences in structure reflect differences in function remains to be seen. Polymorphonuclear granulocytes are known to be rich in hydrolytic enzymes (Cohn et al., '63) and Archer and Hirsch ('63) have shown several lysosomal enzymes in the specific granules of mammalian eosinophilic granulocytes. According to Miller et al. ('66) it is probable that the crystalline cores in the granules of mammalian eosinophilic granulocytes consist of a specific peroxidase. We did not observe crystals in the cytoplasmic granules of newt eosinophilic granulocytes but they are present in the granules of newt basophilic granulocytes. In this respect these cells resemble the basophilic granulocytes of the guinea pig which, alone amongst the mammalian basophilic granulocytes so far studied, have cytoplasmic granules with crystalline cores (Winquist, '60; Fedorko and Hirsch, '65). As Miller et al. ('66) suggest, the presence of crystals within such granules may depend on the types and amounts of enzymes that they contain.

The presence of many interchromatin granules appears to be a characteristic feature of the nuclei of amphibian cells, and we have observed them in all the cell types described here; they also occur in pancreatic cells of the salamander (Swift, '59), in newt erythroblasts and mature erythrocytes of the newt and the frog *Rana esculenta* (Tooze and Davies, '67). We have discussed the electron staining properties and the nature of these granules elsewhere (Davies and Tooze, '66; Tooze and Davies, '67). Because of the large number of interchromatin granules it was sometimes difficult to decide whether granules associated with chromatin in the special manner of perichromatin granules (Swift, '62; Watson, '62a,b) occur in most of the cells described here; however, they are certainly present in macrophages. The significance of the so-called light-staining zones found in the nuclei of many types of leucocytes as well as erythroblasts (Tooze and Davies, '67) is unknown.

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Abbreviations

be, basophilic erythroblast
c, cytoplasm
ce, centriole
e, erythrocyte
er, endoplasmic reticulum
fe, ferritin
fi, fibrils
gb, basophilic granule
gc, cytoplasmic granules
ge, eosinophilic granule
gi, interchromatin granules
gn, neutrophilic granules
go, Golgi substance
gp, perichromatin granules
g1, g2, g3, g4, granules of macrophage cytoplasm
l, lymphocyte
lz, nuclear light-staining zone
m, mitochondrion
ma, macrophage
M.G.G., May-Grünwald-Giemsa
mm, membrane material
mt, microtubules
nu, nucleolus
pe, polychromatic erythroblast
r, ribosomes
re, reticular cell
rf, reticular fibres
rp, red pulp
si, venous sinus
vc, collapsed vesicles
wp, white pulp

All sectioned material, unless otherwise stated, was fixed in glutaraldehyde + OsO_4, embedded in araldite and stained in uranyl acetate, plus *lead citrate.*

PLATE 1

EXPLANATION OF FIGURES

1 Light micrograph of 1 µ section through a segment of spleen fixed in glutaraldehyde only, stained with MaCullum's stain followed by toluidine blue. The outer regions, the red pulp, stain more densely than the inner region, the white pulp. The flattened nuclei of the outer mesothelial cell layer (arrowed) and the underlying connective tissue sheath are seen. × 120.

Figs. 2–9 Light micrographs of cells from a spleen smear stained with May-Grünwald-Giemsa. × 1540.

2 The large cell on the right is a so-called haemocytoblast with a small lymphocyte on the left.

3 Another cell with the staining properties of the haemocytoblast but with a somewhat more granular nucleus.

4 A large lymphocyte, the nucleus containing well-defined chromatin blocks.

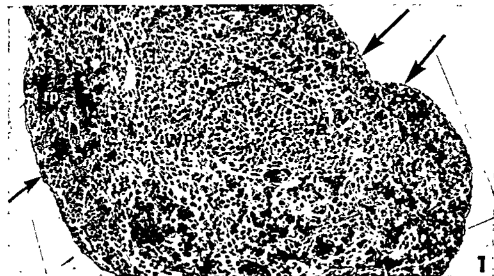
5 A group of *thombocytes with their cytoplasms apparently fused* together.

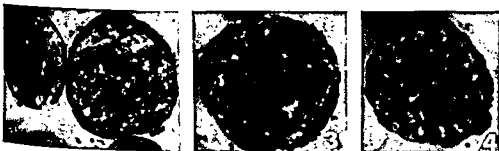
6 A monocyte with characteristic kidney shaped nucleus and lightly stained cytoplasm.

7 An eosinophilic granulocyte with highly lobed nucleus.

8 A basophilic granulocyte with a characteristic bilobed nucleus and cytoplasmic granules.

9 A neutrophilic granulocyte with a highly lobed nucleus and unstained cytoplasm.


rp
WP
1


3
4


5
6


7
8
9

Abbreviations

be, basophilic erythroblast
c, cytoplasm
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er, endoplasmic reticulum
fe, ferritin
fi, fibrils
gb, basophilic granule
gc, cytoplasmic granules
ge, eosinophilic granule
gi, interchromatin granules
gn, neutrophilic granules
go, Golgi substance
gp, perichromatin granules
g1, g2, g3, g4, granules of macrophage cytoplasm
l, lymphocyte
lz, nuclear light-staining zone
m, mitochondrion
ma, macrophage
M.G.G., May-Grünwald-Giemsa
mm, membrane material
mt, microtubules
nu, nucleolus
pe, polychromatic erythroblast
r, ribosomes
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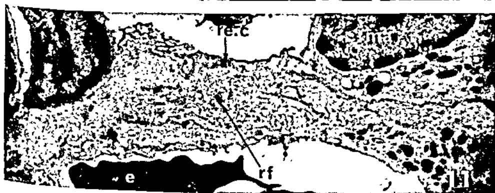
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re.c
m
rf
e
11



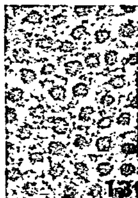


PLATE 2

EXPLANATION OF FIGURES

10 Light micrograph of a 1 μ section through the red pulp of the spleen. The venous sinuses are clearly outlined by lightly stained reticular fibres. The arrows mark cells of the mesothelial layer and the underlying connective tissue sheath. A mitotic polychromatic erythroblast is arrowed (white). × 500.

11 Electron micrograph of part of the reticular framework of the red pulp. Numerous reticular fibres cut transversely and longitudinally are surrounded by reticular cell cytoplasm. × 5,000.

12 Electron micrograph of reticular fibres cut longitudinally. × 90,000.

13 Electron micrograph of reticular fibres cut transversely. × 108,000.


15
e
e
14

PLATE 3

EXPLANATION OF FIGURES

14 Electron micrograph of a small or medium-sized lymphocyte, the cytoplasm of which abuts on that of other cells, probably a macrophage and two erythrocytic cells. Very large (1200Å) granules within the nucleus are arrowed. × 14,000.

15 Electron micrograph of part of a nucleus of a lymphocyte which shows adjacent groups of very large (~ 1200Å) granules (arrow) and interchromatin granules (~ 400Å). × 15,000.


15
e
e
14

PLATE 4

EXPLANATION OF FIGURES

16 Electron micrograph of a haemocytoblast, the cytoplasm of which abuts on that of a macrophage and a polychromatic erythroblast. The haemocytoblast cytoplasm is packed with ribosomes and contains numerous mitochondria and a few membrane-limited densely staining granules. × 10,500.

17 Electron micrograph of the microtubule arrowed in figure 16. × 33,000.


gc
pe
m
g2
ma
17
16

PLATE 5

EXPLANATION OF FIGURE

18 Electron micrograph of a reticular cell showing centrioles lying within Golgi material. Groups of glycogen granules in the cytoplasm are arrowed (see also figs. 19, 21, 22). Profiles of ribosome-studded endoplasmic reticulum are prominent at the bottom of the micrograph. The surface of the nucleus has the characteristic corrugated appearance. × 14,000.


be
pe
fi
gl
nu
lz
er
fi
l

PLATE 6

EXPLANATION OF FIGURES

19 Electron micrograph of part of a reticular cell showing intracellular fibrils lying near the cell periphery (*see* fig. 18). Clusters of glycogen granules about 250Å diameter are arrowed. × 108,000.

20 Electron micrograph of transversely sectioned fibrils in the cytoplasm of a reticular cell. × 125,000.

21 *Electron micrograph of part of a reticular cell with clusters of glycogen* granules: each granule contains small particles about 40Å diameter. × 125,000.

22 Electron micrograph of a reticular cell showing glycogen granules after treatment with periodic acid and lead citrate. × 120,000.

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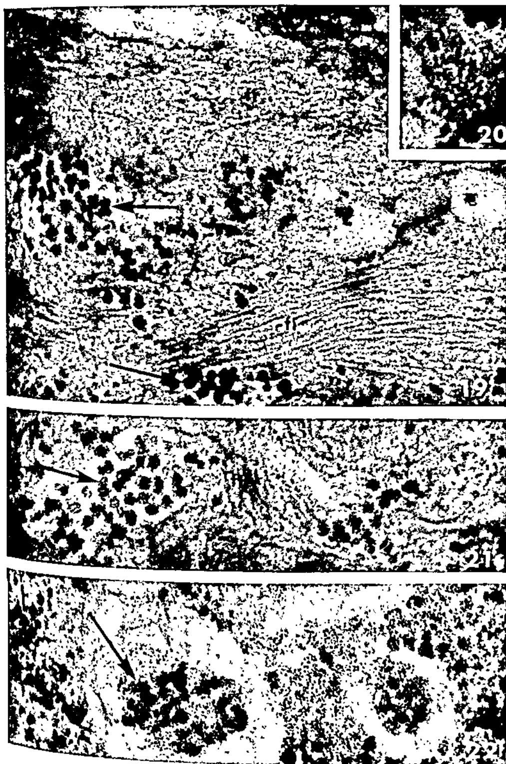


PLATE 6

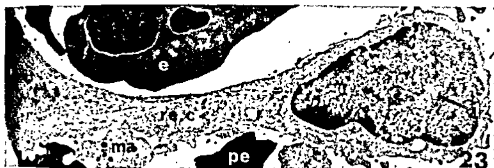
EXPLANATION OF FIGURES

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20 Electron micrograph of transversely sectioned fibrils in the cytoplasm of a reticular cell. × 125,000.

21 Electron micrograph of part of a reticular cell with clusters of glycogen granules: each granule contains small particles about 40Å diameter. × 125,000.

22 Electron micrograph of a reticular cell showing glycogen granules after treatment with periodic acid and lead citrate. × 120,000.


e
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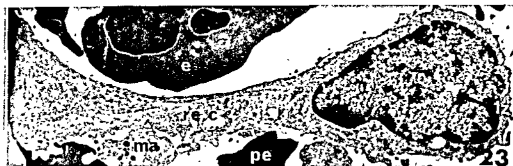
pe
e
g2
e
g1
re.c
rf
l
g2
pe
24

PLATE 7

EXPLANATION OF FIGURES

23 Electron micrograph of part of a reticular cell showing a cytoplasmic extension. × 5,000.

24 Electron micrograph of a macrophage containing a phagocytosed erythrocyte. A cytolysome (polar body) arrow is present within the erythrocyte. Within the cytoplasm of the macrophage lie large dark-grey ferritin-containing granules (g1) and electron opaque granules (g2) (see fig. 28), g1 granules at higher magnification appear like the g3 granule shown in figure 26, but without the large dense inclusion. Note the reticular fibres and associated reticular cell cytoplasm *which form the framework of the spleen.* × 7,300.



pe
e
g2
e
g1
re.c
l
g2
pe
24

PLATE 8

EXPLANATION OF FIGURES

25 Electron micrograph of a macrophage showing cytoplasmic bodies containing extensive myelin formations (arrows) and adjacent g4 and g2 granules (see fig. 28). The cell has long finger-like cytoplasmic extensions. × 15,000.

26 Electron micrograph of part of a macrophage showing a granule (type g3, see text) also containing profiles of stacked membranes (arrow). Note that ferritin is excluded from the mitochondrion. × 88,000.

27 Electron micrograph of part of a macrophage nucleus showing perichromatin granules (arrow) and smaller interchromatin granules (gi). × 42,000.

28 Electron micrograph of an almost electron-opaque granule (g2) with surrounding membrane: the smaller dark-grey granules (g4) are also membrane limited. × 40,000.


fe
m
26


gi
gp
27

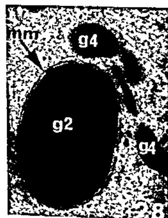

mm
g4
g2
g4
28

PLATE 8

EXPLANATION OF FIGURES

25 *Electron micrograph of a macrophage showing cytoplasmic bodies* containing extensive myelin formations (arrows) and adjacent g4 and g2 granules (see fig. 28). The cell has long finger-like cytoplasmic extensions. × 15,000.

26 Electron micrograph of part of a macrophage showing a granule (type g3, see text) also containing profiles of stacked membranes (arrow). Note that ferritin is excluded from the mitochondrion. × 88,000.

27 Electron micrograph of part of a macrophage nucleus showing perichromatin granules (arrow) and smaller interchromatin granules (*gi*). × *42,000.*

28 Electron micrograph of an almost electron-opaque granule (g2) with surrounding membrane: the smaller dark-grey granules (g4) are also membrane limited. × 40,000.


gc
m
g
30


m
r
m
er
31

PLATE 9

EXPLANATION OF FIGURES

29 Electron micrograph of a plasma cell showing a centriole embedded in Golgi material and numerous extended sacs of endoplasmic reticulum. *The boxed area is enlarged in fig. 30.* × *15,000.*

30 Electron micrograph of part (see box) of the cytoplasm of the cell shown in figure 29 showing microtubules and a membrane-bound cytoplasmic granule (gc). × 15,000.

31 Electron micrograph of part of a plasma cell showing distended endoplasmic reticulum, the cavity of which contains electron scattering material; the membranes which face and partially enclose the mitochondria are preferentially studded with ribosomes. × 57,000.


gn
vc
er
m
pe
gn
32

PLATE 10

EXPLANATION OF FIGURE

32 Electron micrograph of a neutrophilic granulocyte, the specific granules (see figs. 33, 37) of which have only stained faintly: flattened vesicles (see figs. 35, 36) are numerous. × 13,000.


mt
34


35

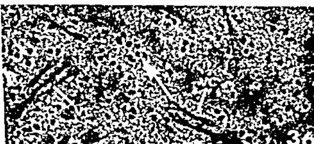






PLATE 11

EXPLANATION OF FIGURES

33 Electron micrograph of a neutrophilic granulocyte showing specific granules (see fig. 37), empty sacs or vesicles (arrows), collapsed vesicles (see figs. 35, 36) and an array of ribosome-studded endoplasmic reticulum. × 37,000.

34 Electron micrograph of the microtubule at site marked mt in figure 33. × 60,000.

35 Electron micrograph of a collapsed vesicle from a neutrophil showing tripartite structure. × 360,000.

36 Electron micrograph showing collapsed vesicles and particles, possibly glycogen (arrowed) lying in the cytoplasm of a neutrophil. × 120,000.

37 Electron micrograph of neutrophilic granules showing limiting membrane (arrows). × 120,000.

38 Electron micrograph of a bundle of fibrils lying in the cytoplasm of a neutrophilic granulocyte. × 120,000.


ge
39

PLATE 12

EXPLANATION OF FIGURE

39 Electron micrograph of an eosinophilic granulocyte. Cytoplasmic vesicles are arrowed. × 9,000.

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gb

40

41

PLATE 13

EXPLANATION OF FIGURES

40 Electron micrograph of a basophilic granulocyte. A crystalline region in one of the granules is arrowed. × 6,000.

41 Electron micrograph of a granule from a basophilic granulocyte which contains a crystalline region (arrow). × 128,000.

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